Emergency and Continuous Exposure Limits for Selected Airborne Contaminants

Volume 2

20030115019

OTIC FILE COPY

Committee on Toxicology Board on Toxicology and Environmental Health Hazards Commission on Life Sciences National Research Council



DISTRIBUTION STATEMENT A 2
Approved for public releases
Distribution Unlimited

REPORT DOCUMENTATION PAGE	1. REPORT NO.	2	3. Recipient's Accession No.	
4. Title and Subtitle			S. Repart Date	
Fmervency and Conti	nuous Exposure Limits for S	elected Airborne	October 1984	
Contamints, Volume			6.	
7. Author(s)			8. Performing Organization R	ept. No.
3. Performing Organization Name a			10. Project/Task/Work Unit N	No.
National Research Co				
Commission on Life S			11. Contract(C) or Grant(G) N	
2101 Constitution A			(c) N00014-80-C-16	
Washington, DC 204	10		AMD-17-82-C-302	
12. Sponsoring Organization Name a	nd Address		13. Type of Report & Period (Covered
U.S. Army, Air Force	e, Navy		Final	•
			14.	
15. Supplementary Notes		\\		
16. Abstract (Limit: 200 words)				
These are emergency	exposure limits recommende	d by the NRC Commi	ittee on Toxicology	y
for chemicals of in	terest to sponsors because	they may affect po	ersonnel during va	rious
operations.	•			
<u></u>				
The report provides	a short review of the chem	ical and physical	properties of the	se
chemicals and their	toxic-potential in animals	and humans.	Accession For	
		Λ	NTIS GRAAI	lof.
		. /	DTIC TAB	
				A
		Ì	Unanaounced	6
	<u>.</u>	/		6
1. 13	5		Unannounced Justification	6
2 12 133			Unannounced Justification By	6
1 June 1 142 135	5	••••	Unanaounced Justification By Distribution/	Codes
1 June 1 135	3	OF 1C	Unannounced Justification By Distribution/ Availability	-
John A 142 135		Maric Maric 1	Unannounced Justification By Distribution/ Availability Avail and	l/or
Vilume 14,2 13.3		Maric Ome, Marico	Unannounced Justification By Distribution/ Availability	l/or
, , , , , , , , , , , , , , , , , , ,		O TO	Unannounced Justification By Distribution/ Availability Avail and	l/or
17. Document Analysis a. Descripto	rs	posure Limits. CEI	By	l/or
17. Document Analysis a. Descripto Emergency Exposure	rs Limits, EELs, Continuous Ex	posure Limits, CEI	Unannounced Justification By Distribution/ Availability Avail and Dist Special	l/or
17. Document Analysis a. Descriptor Emergency Exposure Chlorine, chlorine	re Limits, EELs, Continuous Ex trifluoride, ethanolamin, F	luorocarbon 11, f	Unanaounced Justification By Distribution/ Availability Avail and Dist Special A.1 Luorecarbon 12,	l/or
17. Document Analysis a. Descriptor Emergency Exposure Chlorine, chlorine fluorocarbon 21, fluorocarbon 21,	rs Limits, EELs, Continuous Extrifluoride, ethanolamin, Facorocarbon 113, fluorocarbo	luorocarbon 11, fl n 114, isopropyl a	Unanaounced Justification By Distribution/ Availability Avail and Dist Special A.1 Luorecarbon 12,	l/or
27. Document Analysis a. Descriptor Emergency Exposure of Chlorine, chlorine fluorocarbon 21, fluorocarbon 21, sodium hydroxide, so	re Limits, EELs, Continuous Ex trifluoride, ethanolamin, F	luorocarbon 11, fl n 114, isopropyl a	Unanaounced Justification By Distribution/ Availability Avail and Dist Special A.1 Luorecarbon 12,	l/or
27. Document Analysis a. Descriptor Emergency Exposure Chlorine, chlorine fluorocarbon 21,	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fuorocarbon 113, fluorocarboulfur dioxide, vinylidene c	luorocarbon 11, find	Unannounced Justification By Distribution/ Availability Avail and Dist Special A.1 Luorecarbon 12, alcohol, phosgene,	l/or
27. Document Analysis a. Descriptor Emergency Exposure of Chlorine, chlorine fluorocarbon 21, fluorocarbon 2	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fuorocarbon 113, fluorocarboulfur dioxide, vinylidene citrifluoride, ethanolamine,	luorocarbon 11, fin 114, isopropyl a hloride, xylene.	Unanaounced Justification By Distribution/ Availability Avail and Dist Special A.1 Luorecarbon 12, alcohol, phosgene, procarbon 11,	l/or
27. Document Analysis a. Descriptor Emergency Exposure of Chlorine, chlorine fluorocarbon 21, fluorocarbon 21, fluorocarbon 21, fluorocarbon Ended Terms Chlorine, chlorine fluorocarbon 12, fluo	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fluorocarbon 113, fluorocarbon 11fur dioxide, vinylidene citrifluoride, ethanolamine, porocarbon 21, fluorocarbon	luorocarbon 11, fin 114, isopropyl a hloride. xylene. kkanalaminex fluorocarbon	Unannounced Justification By Distribution/ Availability Avail and Special A.1 Lucrecarbon 12, alcohol, phosgene, procarbon 11, a 114, isopropyl	l/or
27. Document Analysis a. Descriptor Emergency Exposure of Chlorine, chlorine fluorocarbon 21, fluorocarbon 21, fluorocarbon 21, fluorocarbon Ended Terms Chlorine, chlorine fluorocarbon 12, fluo	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fuorocarbon 113, fluorocarboulfur dioxide, vinylidene citrifluoride, ethanolamine,	luorocarbon 11, fin 114, isopropyl a hloride. xylene. kkanalaminex fluorocarbon	Unannounced Justification By Distribution/ Availability Avail and Special A.1 Lucrecarbon 12, alcohol, phosgene, procarbon 11, a 114, isopropyl	l/or
27. Document Analysis a. Descriptor Emergency Exposure of Chlorine, chlorine fluorocarbon 21, fluorocarbon 21, fluorocarbon 21, fluorocarbon 12, fluorocarbon 12, fluorocarbon 12, fluorocarbon 12, fluorocarbon 12, fluorocarbon, phosgene, c. COSATI Field/Group	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fluorocarbon 113, fluorocarbon 11fur dioxide, vinylidene citrifluoride, ethanolamine, porocarbon 21, fluorocarbon	luorocarbon 11, fin 114, isopropyl a hloride. xylene. kkanalaminex fluorocarbon	Unannounced Justification By Distribution/ Availability Avail and Special A.1 Lucrecarbon 12, alcohol, phosgene, procarbon 11, a 114, isopropyl	l/or
27. Document Analysis a. Descriptor Emergency Exposure Chlorine, chlorine fluorocarbon 21, fluorocarbon 21, fluorocarbon b. Identifiers/Open-Ended Terms Chlorine, chlorine fluorocarbon 12, fluorocarbon, phosgene,	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fluorocarbon 113, fluorocarbon 11fur dioxide, vinylidene citrifluoride, ethanolamine, porocarbon 21, fluorocarbon	luorocarbon 11, fin 114, isopropyl a hloride, xylene. Exkansianins; fluorocarbonoxide, vinylidene 19. Security Class (This	Unannounced Justification By Distribution/ Availability Availability Avail and Special Ls. Luorecarbon 12, alcohol, phosgene, procarbon 11, a 114, isopropyl chloride, xylene. Report) 21. No. of Page 129	i/or
27. Document Analysis a. Descriptor Emergency Exposure of Chlorine, chlorine fluorocarbon 21, fluorocarbon 21, fluorocarbon 21, fluorocarbon 12, fluorocarbon 12, fluorocarbon 12, fluorocarbon 12, fluorocarbon 12, fluorocarbon, phosgene, c. COSATI Field/Group	Limits, EELs, Continuous Extrifluoride, ethanolamin, Fuorocarbon 113, fluorocarboulfur dioxide, vinylidene citrifluoride, ethanolamine, porocarbon 21, fluorocarbon sodium hydroxide, sulfur die	luorocarbon 11, fin 114, isopropyl a hloride, xylene. Exhamakaminax fluorocarbonoxide, vinylidene	Unanaounced Justification By	i/or

.

1000000 OF P5525544

DO NOT PRINT THESE INSTRUCTIONS AS A PAGE IN A REPORT

INSTRUCTIONS

Optional Form 272, Report Documentation Page is based on Guidelines for Format and Production of Scientific and Technical Reports, ANSI 739.18—1974 available from American National Standards Institute, 1430 Broadway, New York, New York 10018. Each separately bound report—for example, each volume in a multivolume set—shall have its unique Report Documentation Page.

- 1. Report Number. Each individually bound report shall carry a unique alphanumeric designation assigned by the performing organization or provided by the sponsoring organization in accordance with American National Standard ANSI Z39.23–1974, Technical Report Number (STRN). For registration of report code, contact NTIS Report Number Clearinghouse, Springfield, VA 22161. Use upglercase letters, Arabic numerals, slashes, and hyphens only, as in the following examples: FASEB/NS-75/87 and FAA/ EO-75/09.
- 2. Leave blank.
- 3. Recipient's Accession Number. Reserved for use by each report recipient.
- 4. Title and Subtitle. Title should indicate clearly and briefly the subject coverage of the report, subordinate subtitle to the main title. When a report is prepared in more than one volume, repeat the primary title, add volume number and include subtitle for the specific volume.
- 5. Report Date. Each report shall carry a date indicating at least month and year. Indicate the basis on which it was selected (e.g., date of issue, date of approval, date of preparation, date published).
- 6. Sponsoring Agency Code. Leave blank.
- 7. Author(s). Give name(s) in conventional order (e.g., John R. Doe, or J. Robert Doe). List author's affiliation if it differs from the performing organization.
- 3. Performing Organization Report Number. Insert if performing organization wishes to assign this number.
- Performing Organization Name and Mailing Address. Give name, street, city, state, and ZIP code. List no more than two levels of
 an organizational hierarchy. Display the name of the organization exactly as it should appear in Government indexes such as
 Government Reports Announcements & Index (GRA & I).
- 10. Project/Task/Work Unit Number. Use the project, task and work unit numbers under which the report was prepared.
- 11. Contract/Grant Number. Insert contract or grant number under which report was prepared.
- 12. Sponsoring Agency Name and Mailing Address. Include ZIP code. Cite main sponsors.
- 13. Type of Report and Period Covered. State interim, final, etc., and, if applicable, inclusive dates.
- 14. Performing Organization Code, Leave blank.
- 15. Supplementary Notes. Enter information not included elsewhere but useful, such as: Prepared in cooperation with . . . Translation of . . . Presented at conference of . . . To be published in . . . When a report is revised, include a statement whether the new report supersedes or supplements the older report.
- 16. Abstract, Include a brief (200 words or less) factual summary of the most significant information contained in the report. If the report contains a significant bibliography or literature survey, mention it here.
- 17. Document Analysis. (a). Descriptors. Select from the Thesaurus of Engineering and Scientific Terms the proper authorized terms that identify the major concept of the research and are sufficiently specific and precise to be used as index entries for cataloging.
 - (b). Identifiers and Open-Ended Terms. Use identifiers for project names, code names, equipment designators, etc. Use openended terms written in descriptor form for those subjects for which no descriptor exists.
 - (c). COSATI Field/Group. Field and Group assignments are to be taken from the 1964 COSATI Subject Category List. Since the majority of documents are multidisciplinary in nature, the primary Field/Group assignment(s) will be the specific discipline, area of human endeavor, or type of physical object. The application(s) will be cross-referenced with secondary Field/Group assignments that will follow the primary posting(s).
- 48. Distribution Statement. Denote public releasability, for example "Release unlimited", or limitation for reasons other than security. Cite any availability to the public, with address, order number and price, if known.
- 19. & 20. Security Classification. Enter U.S. Security Classification in accordance with U.S. Security Regulations (i.e., UNCLASSIFIED).
- 21. Mumber of pages. Insert the total number of pages, including introductory pages, but excluding distribution list, if any.
- 22. Price. Enter price in paper copy (PC) and/or microfiche (MF) if known.

EMERGENCY AND CONTINUOUS EXPOSURE LIMITS FOR SELECTED AIRBORNE CONTAMINANTS

Volume 2

COMMITTEE ON TOXICOLOGY

Board on Toxicology and Environmental Health Hazares Commission on Life Sciences National Research Council

> National Academy Press Washington, D.C. October 1984

NOTICE: The project that is the subject of this report was approved by the Governing Board of the National Research Council, whose members are drawn from the councils of the National Academy of Sciences, the National Academy of Engineering, and the Institute of Medicine. The members of the committee responsible for the report were chosen for their special competences and with regard for appropriate balance.

This report has been reviewed by a group other than the authors according to procedures approved by a Report Review Committee consisting of members of the National Academy of Sciences, the National Academy of Engineering, and the Institute of Medicine.

The National Research Council was established by the National Academy of Sciences in 1916 to associate the broad community of science and technology with the Academy's purposes of furthering knowledge and of advising the federal government. The Council operates in accordance with general policies determined by the Academy under the authority of its congressional charter of 1863, which establishes the Academy as a private, nonprofit, self-governing membership corporation. The Council has become the principal operating agency of both the National Academy of Sciences and the National Academy of Engineering in the conduct of their services to the government, the public, and the scientific and engineering communities. It is administered jointly by both Academies and the Institute of Medicine. The National Academy of Engineering and the Institute of Medicine were established in 1964 and 1970, respectively, under the charter of the National Academy of Sciences.

This study was prepared under Contract N00014-80-C-0161 between the National Academy of Sciences and the Office of Naval Research and Contract DAMD-17-82-C-3028 between the National Academy of Sciences and the Department of the Army.

COMMITTEE ON TOXICOLOGY

Roger O. McClellan, Lovelace Inhalation Toxicology Research Institute, Albuquerque, New Mexico, Chairman

Rose Dagirmanjian, University of Louisville, Louisville, Kentucky David W. Gaylor, National Center for Toxicological Research, Jefferson, Arkansas

Richard Griesemer, Oak Ridge National Laboratory, Oak Ridge, Tennessee William Halperin, National Institute for Occupational Safety and Health, Cincinnati, Ohio

Clark W. Heath, Jr., Emory University School of Medicine, Atlanta, Georgia Meryl Karol, Graduate School of Public Health, Pittsburgh, Pennsylvania Ronald J. Spanggord, SRI International, Menlo Park, California

Thomas R. Tephly, The University of Iowa, Iowa City, Iowa

Lloyd B. Tepper, Air Products & Chemicals, Inc., Allentown, Pennsylvania Clarence J. Terhaar, Eastman Kodak Company, Rochester, New York

National Research Council Staff

Francis N. Marzulli, Project Director Gary R. Keilson, Project Director, until September 1983 Kulbir S. Bakshi, Staff Officer Catherine L. St. Hilaire, Staff Officer, until May 1983 Norman Grossblatt, Editor Beulah S. Bresler, Administrative Secretary

BOARD ON TOXICOLOGY AND ENVIRONMENTAL HEALTH HAZARDS

Gerald N. Wogan, Massachusetts Institute of Technology, Cambridge, Massachusetts, Chairman

Donald Hornig, Harvard University, Boston, Massachusetts, Co-Vice-Chairman

Philip Landrigan, National Institute for Occupational Safety and Health, Cincinnati, Ohio, Co-Vice-Chairman

John Doull, University of Kansas Medical Center, Kansas City, Kansas Herman N. Eisen, Massachusetts Institute of Technology, Cambridge, Massachusetts

Emmanuel Farber, University of Toronto, Toronto, Ontario, Canada David G. Hoel, National Institute of Environmental Health Sciences, Research Triangle Park, North Carolina

Richard Merrill, University of Virginia Law School, Charlottesville, Virginia

Vaun A. Newill, Exxon Corporation, New York, New York
Emil Pfitzer, Hoffmann-La Roche Inc., Nutley, New Jersey
Joseph V. Rodricks, Environ Corporation, Washington, D. C.
Liane B. Russell, Oak Ridge National Laboratory, Oak Ridge, Tennessee
Ellen Silbergeld, Environmental Defense Fund, Washington, D. C.
Peter Spencer, Albert Einstein College of Medicine, Bronx, New York

National Research Council Staff

tratian in the contract of the contract of

Devra Lee Davis, Executive Director

ACKNOWLEDGMENTS

This document reflects a continuing effort by the Committee on Toxicology to review and update earlier recommendations that were made with regard to exposure to a variety of airborne contaminants primarily of concern to the Department of Defense and to the National Aeronautics and Space Administration. The preparation of this document has been possible only because of the dedicated efforts of the current and many past members of the Committee on Toxicology.

The document has been evaluated in total by the current members of the Committee; however, much of the work was initially done by former members. The contributions of the following are particularly noted: Richard R. Bates, Health Effects Institute, Cambridge, Mass.; Donald Ecobichon, McGill University; Lawrence Fishbein, National Center for Toxicological Research, Jefferson, Ark.; Peter Greenwald, National Cancer Institute, Bethesda, Md.; Ian Higgins, University of Michigan, Ann Arbor, Mich.; Wendell Kilgore, University of California, Davis, Cal.; Leonard T. Kurland, Mayo Clinic, Rochester, Minn.; Howard Maibach, University of California, San Francisco, Cal.; H. George Mandel, The George Washington University, Washington, D.C.; Robert E. Menzer, University of Maryland, College Park, Md.; Charles Reinhardt, E.I. duPont de Nemours and Company, Newark, Del.; Joseph Rodricks, Environ Corporation, Washington, D.C.; Ronald C. Shank, University of California, Irvine, Cal.; Edward A. Smuckler, University of California, San Francisco, Cal.; Robert Snyder, Rutgers University, Piscataway, N. J.; Peter Spencer, Albert Einstein College of Medicine, Bronx, N. Y.; and Philip Watanabe, Dow Chemical USA, Midland, Mich.

ere perengan dipere del perengan del est del perengan del perengan del perengan de perengan de perengan de la c

CONTENTS

Introduction	1
Summary	3
Chlorine	. 5
Chlorine trifluoride	12
Ethanolamine	17
Fluorocarbon 11	26
Fluorocarbon 12	34
Fluorocarbon 21	41
Fluorocarbon 113	46
Fluorocarbon 114	51
Isopropyl Alcohol	56
Phosgene	69
Sodium Hydroxide	87
Sulfur Dioxide	95
Vinylidene Chloride	103
Kylene	113

INTRODUCTION

The National Research Council's Committee on Toxicolczy recommends emergency exposure limits (EELs), short-term public limits (STPLs), and short-term public emergency limits (SPELs--formerly called public emergency limits, or PELs) for a variety of chemicals of concern to its sponsoring agencies. The definitions and applicability of these limits and the criteria used to establish them were originally outlined in two documents prepared by the Committee (National Research Council, 1964, 1971). In a revision of these documents (National Research Council, 1979), the Committee summarized the principles used to establish exposure limits for short durations. The Committee has also recommended continuous exposure limits (CELs) in response to specific sponsor requests.

This document is one in a series prepared by the Committee that form the basis of the recommendations for EELs and CELs for selected chemicals. Since the Committee began recommending EELs and CELs for its military sponsors (U.S. Army, Navy, and Air Force), the scope of its recommendations has been expanded in response to requests by the U.S. Coast Guard and the National Aeronautics and Space Administration. The CELs grew out of a Navy request for exposure limits for atmospheric contaminants in submarines. The EELs and CELs have been used as design criteria by the sponsors in considering the suitability of materials for particular missions (as in a submarine or a spacecraft) and in assessing the habitability of particular enclosed environments. They are recommended for narrowly defined occupational groups and are not intended for application in general industrial settings or as exposure limits for the general public. These recommended values do not take into consideration the possible effects of exposure of hypersensitive persons.

The EEL is defined as a ceiling limit for an unpredictable single exposure, usually lasting 60 min or less, and never more than 24 h-an occurrence expected to be rare in the lifetime of any person. It reflects an acceptance of the statistical likelihood of the occurrence of a nonincapacitating, reversible effect in an exposed population. It is designed to avoid substantial decrements in performance during emergencies and might contain no uncertainty factor.

The CEL is recommended in specific situations where there may be exposure to a chemical continuously for up to 90 d. It is defined as a ceiling limit designed to avoid adverse health effects, either immediate or delayed, and to avoid degradation in crew performance that might endanger the objectives of a particular mission. Because human data on continuous exposures are rarely available, uncertainty factors might be used, the magnitude depending on the judgment of the Committee.

(min-)

こうしょう じょうじゅう ましんりゅうじょ 神をあるを見る 美国のならなられる かんかいさんき かんかいかいじょ しょうしょ

REFERENCES

National Research Council, Ad Hoc Committee of the Committee on Toxicology. 1964. Basis for Establishing Emergency Inhalation Exposure Limits Applicable to Military and Space Chemicals. Washington, D.C.: National Academy of Sciences. [5 p.]

National Research Council, Committee on Toxicology. 1971. Basis for Establishing Guides for Short-Term Exposures of the Public to Air Pollutants. Washington, D.C.: National Academy of Sciences. [15 p.]

National Research Council, Committee on Toxicology. 1979. Criteria for Short-Term Exposures to Air Pollutants. Washington, D.C.: National Academy of Sciences. [15 p.]

SUMMARY

Because this report deals with diverse substances, a conventional summary would be inappropriate. Table 1 summarizes previously recommended and currently recommended emergency exposure limits (EELs) for up to 24 h and continuous exposure limits (CELs) for 90 d for chemicals suggested for review by military sponsors of the Committee on Toxicology.

TABLE 1

Emergency and Continuous Exposure Limits

Substance Exposure Year Limit Recommendations		Duration of	Describer	Dana mandahi	
Chlorine 60 min 1971 3 ppm 3 ppm 0.5 ppm 90 d 0.1 ppm 0.1 ppm 0.1 ppm 0.1 ppm Chlorine trifluoride 10 min 1968 7 ppm 7 ppm 3 ppm 3 ppm 3 ppm 1 ppm 1 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm 0.5 ppm		•			
24 h 1 ppm 0.5 ppm 90 d 0.1 ppm 0.1 ppm Chlorine trifluoride 10 min 1968 7 ppm 7 ppm 30 min 3 ppm 3 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm	Substance	Exposure	Year	Limit	Recommendations
24 h 1 ppm 0.5 ppm 90 d 0.1 ppm 0.1 ppm Chlorine trifluoride 10 min 1968 7 ppm 7 ppm 3 ppm 3 ppm 60 min 1 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm	Chlorine	60 min	1971	mag E	3 ppm
90 d 0.1 ppm 0.1 ppm Chlorine trifluoride 10 min 1968 7 ppm 7 ppm 3 ppm 3 ppm 60 min 1 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 3 ppm 24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm		24 h			0.5 ppm
30 min 3 ppm 3 ppm 60 min 1 ppm 1 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm		90 d			
60 min 1 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm	Chlorine trifluoride	10 min	1968	7 ppm	7 pρ m
60 min 1 ppm 1 ppm Ethanolamine 60 min 1967 50 ppm 50 ppm 24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm		30 min		3 ррт	3 ppm
24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm		60 min			
24 h 3 ppm 3 ppm 90 d 0.5 ppm 0.5 ppm	Ethanolamine	60 min	1967	50 ppma	50 ppm
90 d 0.5 ppm 0.5 ppm		24 h			
		90 d			- -
Trichlorofluoromethane 60 min 1966 30,000 ppm 1,500 ppm	Trichlorofluoromethane	60 min	1966	30,000 ppm	1,500 ppm
(FC 11) 24 h 20,000 ppm 500 ppm	(FC 11)	24 h		20,000 ppm	500 ppm
90 d 1,000 ppm 100 ppm		90 d		1,000 ppm	
Dichlorodifluoromethane 60 min 1966 30,000 ppm 10,000 ppm	Dichlorodifluoromethane	60 min	1966	30,000 ppm	10,000 ppm
(FC 12) 24 h 20,000 ppm 1,000 ppm	(FC 12)	24 h		20,000 ppm	1,000 ppm
90 d 1,000 ppm 100 ppm		90 d			100 ppm
Dichlorofluoromethane 60 min 100 ppm	Dichlorofluoromethane	60 min			100 ppm
(FC 21) 24 h 3 ppm	(FC 21)	24 h			3 ppm
90 d 1 ppm	·	90 d			
Trichlorotrifluoroethane 60 min 1969 1,500 ppm 1,500 ppm	Trichlorotrifluoroethane	60 min	1969	1,500 ppm	1,500 ppm
(FC 113) 24 h 200 ppm 500 ppm	(FC 113)	24 h			
90 d 100 ppm 100 ppm					
Dichlorotetrafluoroethane 60 min 1966 30,000 ppm 10,000 ppm	Dichlorotetrafluoroethane	60 min	1966	30.000 ppm	10.000 ppm
(FC 114) 24 h 20,000 ppm 1,000 ppm					
90 d 1,000 ppm 100 ppm	· · · · · · · · · · · · · · · · · · ·				

TOTAL STATES OF THE STATES OF

TABLE 1 (continued)

	Duration of	Previou	s Recommendations	Current
Substance	Exposure	Year	Limit	Recommendations
Isopropyl alcohol	60 min	1966	400 ppm	400 ppm
	24 h		200 ppm	200 ppm
	90 d		50 ppm	1 ppm
Phosgene	60 min	1966	1 ppm	0.2 ppm
	24 h		0.1 ppm	0.02 ppm
	90 d		0.05 ppm	0.01 ppm
Sodium hydroxide	10 min	1965	4 mg/m^3	2 mg/m^3
	30 min		4 mg/m^3	2 mg/m^3
	60 min		2 mg/m^3	2 mg/m^3
Sulfur dicxide	10 min	1966	30 ppm	30 ppm
	30 min		20 ppm	20 ppm
	60 min		10 ppm	10 ppm
	24 h		5 ppm	5 ppm
	90 d		1 ppm	1 ppm
Vinylidene chloride	24 h	1966	25 ppm	10 ppm
	90 d		2 ppm	0.15 ppm
Xylene	60 min	1966	200 ppm	200 ppm
	24 h		100 ppm	100 ppm
	90 d		50 ppm	50 ppm

CHLORINE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: Cl₂
Molecular weight: 70.906
CAS number: 7782-50-5

Boiling point: -34.05°C

Density: 1.4085 (20°C)

General characteristics: Greenish-yellow gas with suffocating

odor

Conversion factors: 1 ppm = 2.89 mg/m^3

 $1 \text{ mg/m}^3 = 0.34 \text{ ppm}$

OCCURRENCE AND USE

Chlorine is abundant in combined form in the earth's crust (0.19%) and in seawater (3%). It is usually produced by electrolysis of chlorides. Contaminants include hexachloroethane, hexachlorobenzene, and water. Chlorine was among the first of the war gases used in World War I. It is now used as a bleaching agent, as a germicidal agent for purifying water, and in the manufacture of chlorinated hydrocarbons and chlorine-containing chemicals. About 10 million tons of chlorine is produced annually for industrial use. Massive exposure to fumes may occur during transportation accidents, but the development of treatment procedures has resulted in low incidences of morbidity and mortality in industrial operations, as opposed to nonindustrial exposures (ACGIH, 1980; Lawson, 1981; Stecher et al., 1968; Ploysongsang et al., 1982).

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

An epidemiologic study by Patil et al. (1970) evaluated a population of 500 diaphragm-cell workers exposed to c'horine at a time-weighted average concentration of 0.146 ± 0.287 ppm (range, 0.006-1.42 ppm). It was concluded that results of chest x rays, ECGs, and pulmonary-function tests were no different between exposed and 382 control workers.

A chlorine study performed on 30 students at the University of Michigan by Anglen et al. (1980) disclosed some sensation of odor, throat irritation, and urge to cough in subjects exposed for 4 h at 0.5 and 1.0 ppm. Exposure at 2 ppm was reported to be much more irritating than that at 0.5 and 1.0 ppm for the same period. The effects were confirmed by a physician.

A study of pulmonary function on four healthy adults accidentally exposed for 2-5 min to chlorine (at unknown concentrations) showed

temporary lung-function impairment, which cleared, with no residual lung damage, after 1 mo. Fourteen to sixteen hours after exposure, patients were symptomatic, with cough, chest tightness, and shortness of breath. All had restrictive ventilating defects with impaired diffusing capacity and evidence of obstruction of small airways (Ploysongsang at al., 1982).

Results from a study sponsored by the Chlorine Institute (Rotman et al., 1983) indicate that an 8-h exposure of humans to chlorine at 1 ppm resulted in sensory irritation and changes in pulmonary functions. The literature on the health effects of chlorine has recently been reviewed (National Research Council, 1975).

Data published on airborne exposures of humans to chlorine are summarized in Table 2.

EFFECTS ON ANIMALS

ì

Barrow and Smith (1975) and Barrow et al. (1977) demonstrated that chlorine exposure caused alterations of pulmonary function in rabbits and reduced respiratory rate in mice. The concentration of chlorine to which exposure for 10 min was required to decrease respiratory rate in mice by 50% (RD50) was about 10 ppm. The authors suggested that exposure to a chemical at a concentration that reduced respiratory rate in mice by 50% would be intolerable and incapacitating to humans and that one-tenth of the RD50 might create some discomfort, but would be tolerable. Although this assumption appears to be true for chlorine, studies with other substances have challenged its general applicability. Potts and Lederer (1978) have shown that the pyrolysis products of red oak at concentrations that reduced respiratory rate in mice by 50% did not incapacitate humans. Therefore, use of the RD50 in mice for predicting sensory irritation in humans may very well be compound-specific.

Barrow et al. (1978) also reported studies of male and female Fischer 344 rats (10 of each sex) exposed to chlorine at 1, 3, or 9 ppm for 6 h/d, 5 d/wk, for 6 wk. The results showed decreased body weights in females at all concentrations and in males at 3 and 9 ppm. Three females died before the end of the study. Urinalysis, hematologic tests, and clinical-chemistry measurements were completed for the surviving animals. The urinary specific gravity was increased in females at all exposure concentrations and ir males at 3 and 9 ppm. The hematocrit and white-blood-cell count were increased in females exposed at 9 ppm. Clinical-chemistry results included increases in alkaline phosphatase, blood urea nitrogen (BUN), Y-glutamyl transpeptidase (GGTP), and serum glutamic pyruvic transaminase (SGPT) at 9 ppm and in alkaline phosphatase at 3 ppm.

Pathologic examination of rats exposed at 9 ppm showed gross evidence of inflammatory reactions of the upper and lower respiratory tract, including hyperemia and accumulation of inflammatory material in the nasal passages. There were also various degrees of pulmonary atelectasis or consolidation. These observations were also made, but to a much smaller degree, in rats exposed at 3 ppm. The kidneys of rats exposed at 9 ppm were found to be darkened. These data indicated that repeated exposures of rats to chlorine at 3 and 9 ppm resulted in gross pathologic changes of the respiratory tract, significantly

decreased body weight, and altered kidney function and revealed a greater sensitivity of females. Although the results suggested that repeated exposure to chlorine at 1 ppm may have produced some toxicity, personal communication with the authors has revealed that chloramine may have been formed from chlorine and ammonia in the inhalation chamber during exposure. Thus, it was not certain whether repeated exposure to chlorine at 1 ppm alone was responsible for the toxic effects observed.

Chlorine itself is not absorbed. The chloride content of the plasma increases for a few hours after gassing, and urinary chloride excretion is increased on the second day after gassing.

In living tissues, chlorine rapidly converts to hypochlorous acid (Zillich, 1972), which easily penetrates the cell wall and reacts with cytoplasmic proteins to form N-chloro derivatives that destroy cell structure (National Research Council, 1975).

Data on animals exposed to chlorine are summarized in Table 3.

INHALATION EXPOSURE LIMITS

The American Conference of Governmental Industrial Hygienists (1980, 1983) has established a TLV-TWA for chlorine of 1 ppm and a 15-min TLV-STEL of 3 ppm. The TLV-TWA of 1 ppm was recommended "to minimize chronic changes in the lungs, accelerated aging, and erosion of the teeth." The Occupational Safety and Health Administration (1983) adopted a ceiling limit of 1 ppm as the federal workplace standard for chlorine.

COMMITTEE RECOMMENDATIONS

Emergency exposure limits were set by the Committee in 1966 and 1971 primarily on the basis of the chlorine concentrations that produced nasal and eye irritation. Having reviewed available toxicity data on chlorine, the Committee believes that no compelling new information warrants revision of the previously established 60-min EEL and 90-d CEL. However, on the basis of human data that suggest sensory irritation and changes in pulmonary function as results of 8-h exposure to chlorine at 1 ppm, the Committee believes that the 24-h EEL should be lowered to 0.5 ppm.

The present Committee's recommended EELs and CEL for chlorine and the limits proposed in 1966 and 1971 are shown below.

	1966/1971	1984
60-min EEL	3 ppm	3 ppm
24-h EEL	1 ppm	0.5 ppm
90-d CEL	0.1 ppm	0.1 ppm

TABLE 2
Human Exposure to Chlorine

Concentration, ppm	Comments	Reference
34-51	Lethal in 1-1.5 h	Freitag, 1941
14-21	Dangerous within 0.5-1 h	Heyroth, 1963
7	Recommended 5-min EEL®	Zielhuis, 1970
5	Severe irritation of eyes, nose, and respiratory tract; intolerable after a few minutes	Zielhuis, 1970
3–6	Stinging or burning in eyes, nose, and throat; perhaps headache due to sinus irritation; watering of eyes; sneezing; coughing; bloody nose; blood-tinged sputum	Heyroth, 1963
5 .	Recommended 15-min EEL ⁸	Zielhuis, 1970
4	Recommended 30-min EEL ^a	Zielhuis, 1970
3.5	Odor threshold	ACGIH, 1980
3	Short-term (5-min) limit	Pa. Dept. of Health undated
3	Permissible exposure (15 min)	ACGIH, 1983
3	Recommended 60-min EELa	Zielhuis, 1970
1-3	Slight irritation	Zielhuis, 1970
1-2	Men can work without interruption	Heyroth, 1963
1	ACGIH TLV-TWA and OSHA standard	ACGIH, 1983 OSHA, 1983
0.146 (Lean)	No effects on chest x ray, ECG, or pulmonary-function after 30 yr of exposure	Patil et al., 1970

A For emergency in manufacturing area and its neighborhood, in storage facilities and surrounding areas, and during transport; suggested by Zielhuis in 1970.

TABLE 3

Animal Exposure to Chlorine

Species	Concentration, ppm	Duration	Effects	Reference
Rabbit	50, 100, 200	30 min	Alteration of pulmonary function	Barrow and Smith, 1975
Mouse	0.7, 38.4	10 min	RD ₅₀ : 10 ppm	Barrow <u>et al</u> ., 1977
Rat	9	6 h/d, 5 d/wk, 6 wk	Decreased body weight; increased urinary specific gravity; increased alkaline phosphatase, GGTP, SGPT, and BUN; increased hematocrit and WBC (females only); pulmonary inflammation and darkened kidneys at autopsy	Barrow <u>et al</u> ., 1978
Rat	3	6 h/d, 5 d/wk, 6 wk	Decreased body weight; increased urinary speci- fic gravity; increased alkaline phosphatase; les pulmonary inflammation that 9 ppm	
Rat	1	6 h/d, 5 d/wk, 6 wk	Decreased body weight (females only); increased urinary specifi gravity (females only) ^a	Barrow <u>et al</u> ., 1978

A Whether effects are due to chlorine is questionable, in that chloramine may have been present in inhalation chambers.

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Chlorine. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists, Inc. p. 80.

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Anglen, D.M., Smith, R.G., Byers, D.H., and Hecker, L.H. 1980. Sensory response of human subjects to low levels of chlorine in air. American Industrial Hygiene Conference Abstracts, May 27-June 1, 1980. Chicago, Ill. p. 92, abstr. no. 159.

Barrow, C.S., Alarie, Y., Warrick, J.C., and Stock, M.F. 1977. Comparison of the sensory irritation response in mice to chlorine and hydrogen chloride. Arch. Environ. Health 32:68-76.

Barrow, C.S., Kociba, R.J., and Rampy, L.W. 1978. A thirty-day inhalation toxicity study of chlorine in Fischer 344 rats. Toxicol. Appl. Pharmacol. 45:290, abstr. no. 162.

Barrow, R.E. and Smith, R.G. 1975. Chlorine-induced pulmonary function changes in rabbits. Am. Ind. Hyg. Assoc. J. 36:398-403.

Freitag. 1941. Danger of chlorine gas. Z. Gesamte Schiess Sprengstoffwes.

Heyroth, F.F. 1963. Halogens. In: Fassett, D.W., and Irish, D.E., eds. Patty's Industrial Hygiene and Toxicology. Vol. II. Toxicology. 2nd revised ed. New York: Interscience Publishers. p. 831-857.

Lawson, J.J. 1981. Chlorine exposure: A challenge to the physician. Am. Fam. Physician 23:135-138.

National Research Council. 1975. Chlorine and Hydrogen Chloride. Washington, D.C.: National Academy of Sciences. 282 p.

Occupational Safety and Health Administration. 1982. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Patil, L.R.S., Smith, R.G., Vorwald, A.J., and Mooney, T.F., Jr. 1970. The health of diaphragm cell workers exposed to chlorine. Am. Ind. Hyg. Assoc., J. 31:678 686.

Pennsylvania Department of Health. Supplement #1 to Short Term Limits for Exposure to Airborne Contaminants. A Documentation 1967-1969. Harrisburg, Pennsylvania: Pennsylvania Dept. Health, Division of Occupational Health. [151] p.

Ploysongsang, Y., Beach, B.C., and DiLisio, R.E. 1982. Pulmonary function changes after acute inhalation of chlorine gas. South. Med. J. 75:23-26.

Potts, W.J., and Lederer, T.S. 1978. Some limitations in the use of the sensory irritation method as an end-point in measurement of smoke toxicity. J. Combust. Toxicol. 5:182-195. [Chem. Abs. 89:141361t, 1978]

Rotman, H.H., Flergelman, M.J., Moore, T., Smith, R.J., Anglen, D.M., Kowalski, C.J., and Weg, J., 1983. Effects of low concentrations of chlorine on pulmonary function. Japp Phys. 54:1120-1124.

Stecher, P.G., Windholz, M., and Leahy, D.S., eds. 1968. The Merck Index: An Encyclopedia of Chemicals and Drugs. 8th ed. Rahway, New Jersey: Merck & Co., Inc. p. 236-237.

Zielhuis, R.L. 1970. Tentative emergency exposure limits for sulphur dioxide, sulphuric acid, chlorine and phosgene. Ann. Occup. Hyg. 13:171-176.

Zillich, J.A. 1972. Toxicity of combined chlorine residuals to fresh water fish. J. Water Pollution Control. Fed. 44:212-220.

CHLORINE TRIFLUORIDE

BATHER DUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: C1F3 Molecular weight: 92.46

CAS number: 7790-91-2 Melting point: -80 to -83°C Boiling point: 11.2-11.75°C

Density: 1.8403 g/m1 (15°C)

General characteristics: May exist as white solid, colorless

to pale yellow-green liquid, or practically colorless gas; sweet suffocating odor; odor threshold very low, but not accurately known (Cloyd and Murphy, 1965); vapor phase decomposes into Cl2, ClF, C10F, C102, and HF, depending on

availability of water

1 ppm = 3.78 mg/m^3 1 mg/m³ = 0.26 ppmConversion factors:

OCCURRENCE AND USE

Chlorine trifluoride is used as a fluorinating agent, in nuclear-fuel processing, as an incendiary, and as an igniter and propellant for rockets (Windholz et al., 1976). It is prepared by reaction of chlorine and fluorine at 200°C and condensation of the product at -80°C. The product obtained is 99.0% pure (Hawley, 1977). Dost et al. (1974) noted the instability of chlorine trifluoride and the potential diversity of its hydrolysis products.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

At high concentations (detectable by odor), exposure can cause gasping, swelling of the eyes and eyelids, cloudiness of the cornea, lacrimation, severe salivation, coughing, breathing difficulties, and convulsions within a few minutes (Cloyd and Murphy, 1965). Damage to the eyes is said to be permanent (Leins, undated). Concentrations high enough to be fatal would be so irritating to eyes, throat, and lungs as to be intolerable. The halogen-pungent odor of chlorine trifluoride can be detected at sufficiently low concentrations that exposed personnel may not experience adverse effects if they evacuate the area immediately (Cloyd and Murphy, 1965). Contact with skin causes severe burns and ulcers that are difficult to heal (Cloyd and Murphy, 1965; Leins, undated). The tissue destruction is caused by

oxidation of tissues, thermal damage from the heat of oxidation, and the effects of HF formed (Cloyd and Murphy, 1965).

In the only documented human exposure, a worker was exposed to the effluent of a chlorine trifluoride-charcoal reactor when he was eating lunch approximately 200 ft downwind from the disposal system. The exposure lasted about 1-2 min. He reported to the medical clinic with symptoms of frontal headache, a bad taste, abdominal pain, and breathing difficulty that persisted for some 2 h. A physician treated him with oxygen therapy for 0.5 h and with APCs (a mixture of aspirin, phenacetin, and caffeine) and the symptoms were relieved. No systemic or local effects were found. The patient reported for work the next day, with no apparent after-effects except fatigue (Longley et al., 1965).

EFFECTS ON ANIMALS

Rats exposed in a dynamic-flow chamber to chlorine trifluoride at 400 ppm died within 40 min; at 800 ppm, the LT50 (lethal time for 50% of the animals) was 15 min. Inhalation exposure of rats at 800 ppm for 15 min was always lethal, whereas exposure at 400 ppm was usually lethal after 35 min of exposure (Dost et al., 1974). In another experiment, 1-h exposure to chlorine trifluoride led to LC50s of 299 (260-344) ppm in male rats and 178 (169-187) ppm in mice (Vernot et al., 1977).

In extensively described experiments, rats were exposed at 480, 96, and 21 ppm; two dogs were also exposed at 21 ppm (Horn and Weir, 1955). The LT50 at 480 ppm was 40 min; at 96 ppm, it was 3.7 h. At 480 ppm, the rats immediately exhibited increased activity. After 2 min, rhinorrhea was noted. From then on, symptoms of respiratory difficulty, eye irritation, and excessive salivation developed. Within 20 min, all were in "acute distress." In several rats, the cornea looked dull and milky-white where it was not protected by the eyelid. All rats died within 70 min; death was usually preceded by excitement and occasionally by convulsions and coma. The same signs developed, at a lower rate, in the rats exposed at 96 ppm. After 4.5 h of exposure, 70% of the animals were dead. Shortly after the survivors were removed from the chamber, two more died (total mortality, 80%). Some showed excitement and "tonic movements" before death. The gross pathology of the lungs from the animals exposed at 480 and 96 ppm was the same: emphysema, pulmonary edema, vascular congestion, and fusion of the lining cells of the bronchi into a hyaline membrane. The livers exhibited hydropic degeneration and marked vascular congestion. In the 96-ppm group, only the gastrointestinal tract was markedly distended with gas.

THE PERSONAL PROPERTY OF THE P

Rats and dogs were exposed at 21 ppm 6 h/d for 2 d. About 10 min after exposure began, the dogs had rhinorrhea and lacrimation and kept their eyes tightly closed. During the first day, they coughed up mucous material and had rapid respiration and excessive salivation. When removed from the chamber, they refused to eat or drink, and they kept their eyes tightly closed. The conjunctivae were markedly injected. The animals' fur felt as though it had been "singed." Toxic signs also appeared early in the rats, with preening and rhinorrhea. By the end of the first day, rhinorrhea and lacrimation

 C_{ij} (with C_{ij}) and C_{ij}

were observed. By the next morning, the animals were essentially normal, except that the dogs' eyes were still markedly inflamed. The second day of the exposure followed the same course as the first up to 4.5 h after starting. The exposure was then halted, because the exhaust system had become plugged and air flow had decreased. The concentration at which the animals had been exposed was not determined, but certainly had risen somewhat. After removal from the chamber, one dog developed severe bilateral corneal ulcers, which had re-epithelialized after a month. Within a few days, all the animals were essentially normal, except for the presence of corneal ulcers in one dog.

Horn and Weir (1955) also exposed two dogs and 20 rats to chlorine trifluoride at an average concentration of 5.15 ppm for 6 h/d, 5 d/wk, for 6 wk (31 exposures). The same signs developed over the first several days as had after acute exposure, but they were less severe. By the midpoint of the experiment, some respiratory distress developed; one dog died after 17 d, and another on the twenty-sixth day. Both apparently died of pneumonia, although penicillin therapy was given to the second. Only one rat died during the experiment; its death was preceded by a convulsion. All others appeared to be in poor health. All the animals that died had bronchopneumonia, bronchiectasis, purulent bronchiolitis, and lung abscesses. Those killed at the end of the experiment had lung hyperemia, hemorrhage, and edema. The kidneys and livers showed vascular congestion.

In a chronic exposure study, 20 rats and two dogs were exposed to chlorine trifluoride at an average concentration of 3.15 ppm for 6 h/d, 5 d/wk, for 6 mo (Horn and Weir, 1956). At the start of the experiment, the dogs exhibited the early signs of irritation, but the rats appeared unaffected and quiet. Recovery was complete overnight. By the ninth day, the rats started preening immediately after exposure began; they then became "depressed" and remained so for the rest of the exposure period. One-fourth the rats died during the experiment. Autopsy of these rats showed almonary edema and bronchopneumonia. The animals that survived showed essentially the same signs as were seen in the subacute exposures. On autopsy, pulmonary irritation was observed. One dog died on day 115 of the experiment of purulent bronchitis and pulmonary abscesses. The other had alveolar hemorrhage, interstitial edema, and pulmonary irritation on autopsy.

INHALATION EXPOSURE LIMITS

そうしょうしゅ やくりいいいし 野木のじのいは 一般ないのななな じゃくらんじょ

The ACGIH has established a ceiling limit for chlorine trifluoride of 0.1 ppm (approximately 0.4 mg/m³) (ACGIH, 1980, 1983). Its recommendation noted that this limit is probably low enough to prevent development of serious injury, but the suitability of the limit requires further evaluation in controlled worker exposure.

COMMITTEE RECOMMENDATIONS

The toxicology of chlorine trifluoride was reviewed originally by the Committee on Toxicology in 1962 and updated in 1968.

The only new information that was not available to the Committee at the previous updating deals with the acute toxicity of the compound by inhalation. The determined values are consistent with previous data. Therefore, there sppears to be no reason to change the previously established exposure limits. However, as stated by ACGIH, data are still needed on controlled exposure of workers in occupational settings.

The present Committee's recommended EELs for chlorine trifluoride and the limits proposed in 1968 are shown below.

・シストランド かいかいこう かいまい かいまい かいしいいい まないしょうきゅう 一部のののできる はいしゅう はいしゅう はいしゅう ないしゅう ないしゅう ないない アプランド できる アプランド

	1968	1984
10-min EEL	7 ppm	7 ppm
30-min EEL	3 ppm	3 ppm
60-min EEL	1 ppm	1 ppm

TO A CONTROL OF A CASE OF THE STATE OF THE SECOND

. De la compressa de la compress

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Chlorine trifluoride. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 81-82.

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R). Threshold Limit Values for Chemical Substances and Physical Agents in Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Cloyd, D.R., and Murphy, W.J. 1965. Handling of Hazardous Materials. Technology Survey. Washington, D.C.: National Aeronautics and Space Administration. 93 p.

Dost, F.N., Reed, D.J., Smith, V.N., and Wang, C.H. 1974. Toxic properties of chlorine trifluoride. Toxicol. Appl. Pharmacol. 27: 527-536.

Hawley, G.G. 1977. The Condensed Chemical Dictionary. 9th ed. New York: Van Nostrand Reinhold Co. p. 192.

Horn, H.J., and Weir, R.J. 1955. Inhalation toxicology of chlorine trifluoride. I. Acute and subacute toxicity. AMA Arch. Ind. Health 12:515-521.

Horn, H.J., and Weir, R.J. 1956. Inhalation toxicology of chlorine trifluoride. II. Chronic toxicity. AMA Arch. Ind. Health 13:340-345.

Leins, J.A. 1961. Aero-jet General Corporation Test Operations. Safety Procedure No. 26. Aero-jet General Corp., California. 9 p.

Longley, M.Y., Pierce, J.F., and Griesemer, E.C. 1965. (U) A toxic hazard study of selected missile propellants (1965). AMRL-TR-65-99. Wright-Patterson Air Force Base, Ohio: Aerospace Medical Research Laboratories. 87 p.

Vernot, E.H., MacEwen, J.D., Haun, C.C., and Kinkead, E.R. 1977. Acute toxicity and skin corrosion data for some organic and inorganic compounds and aqueous solutions. Toxicol. Appl. Pharmacol. 42:417-423.

Windholz, M., Budavari, S., Stroumtsos, L.Y., and Fertig, M.N., eds. 1976. The Merck Index: An Encyclopedia of Chemicals and Drugs. 9th ed. Rahway, New Jersey: Merck & Co., Inc. p. 266.

ETHANOLAMINE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: C₂H₇NO Molecular weight: 61.08

Chemical Names: 2-Aminoethanol, ethylolamine,

B-aminoethyl alcohol, glycinol,

2-hydroxyethylamine,

B-hydroxyethylamine, monoethylamine

THE PARTY AND THE PROPERTY OF THE PROPERTY OF

Synonyms: Olamine, colamine

CAS number: 141-43-5 Melting point: 10.5°C

Boiling point: 170.5°C (760 mm Hg)

Flash point (open cup): 200°F

Specific gravity: 1.0179

Vapor pressure: 0.4 mm Hg (20°C)
Vapor density: 2.1 (air = 1)

Solubility in water: Complete

Odor threshold (sensation): 2.6 ppm (Weeks et al., 1960)
Odor threshold (describable): 25 ppm (Weeks et al., 1960)
General characteristics: Colorless liquid with mildly

ammoniacal odor and relatively

strong base

Conversion factors: 1 ppm = 2.5 mg/m³

 $1 \text{ mg/m}^3 = 0.4 \text{ ppm}$

OCCURRENCE AND USE

Ethauolamine is used to remove carbon dioxide from submarines. It is also used to scrub natural gas; as a dispersant for agricultural products; as a softening agent for hides; as an accelerator (after reaction with other substances) in the production of antibiotics, polishes, and waving solutions for hair; as a corrosion inhibitor; as a rubber accelerator; an intermediate in the production of emulsifiers, soaps, and detergents; and in some hair-care products. Dow, Olin, Texaco, and Union Carbide are the primary producers in the United States. Ethanolamine may contain isopropylamine, diethanolamine, triethanolamine, water, and ammonia.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

The I.G. Elberfeld Toxicology Irdex of 1931, as quoted by Browning (1953), stated that pure ethanolamine produced marked redness and infiltration when it was applied to human skin on gauze and allowed to

remain for 1.5 h. There is little other information on the effects of ethanolamine in man. An anonymous report in Bioenvironmental Safety News Letter (1972) described a case in which a drop of amine fell into the right eye of a sailor aboard a submarine. Within a minute, he was able to get to an eye bath, and he flushed his eye for 30 min. He had prompt medical attention, but vision in his right eye deteriorated from 20/20 to 20/200. The report stated that "he will have partial permanent disability." The same report referred to an earlier, similar incident.

Paustovskaia et al. (1977) stated that "persons working in conditions of increased concentrations of derivatives of cyclo- and dicyclohexylamine and MEA inhibitors of atmospheric corrosion of metals frequently showed changes of the central nervous system, myocardium and hepatobiliary system. Changes in the heart and liver result from the toxic effect of amine derivations on the tissues and also of indirect influence via the central nervous system. Estimation of lactic dehydrogenase isoenzymes is a valuable differential—diagnostic index of myocardial and hepatic pathology due to the effect of amines of the polymethylene series."*

The maximal permissible concentration of ethanolawine suggested by Sidorov and Timofeevskaya (1979) was $0.5~\text{mg/m}^3$ (approximately 0.2~ppm). They stated that workers exposed to ethanolamine at concentrations of greater than 1 mg/m^3 suffer from chronic bronchitis, disorders of the liver, asthenic syndrome, and dystonia.

No controlled-exposure or epidemiologic studies with ethanolamine have been reported.

EFFECTS ON ANIMALS

LD₅₀ values are given in Table 4. Grant (1962) referred to Carpenter and Smyth (1946) and stated that a drop of ethanolamine applied to rabbit eyes causes injury similar to that caused by ammonia, but slightly less severe (graded 9 on a scale of 1-10 after 24 h). Union Carbide Corporation (1970) reported that the dermal LD₅₀ of ethanolamine in rabbits is 1.00 ml/kg (24-h covered exposure). It was judged a moderate primary skin irritant, on the basis of the response to application of 0.01-ml amounts to uncovered rabbit skin 24 h later. Hinglais (1947) reported that ethanolamine had considerable necrotic action on the skin.

In a study of inhalation toxicity, Sidorov et al. (1968) exposed mice and cats to vapors of athanolamine (heated to 80°C) for 2 h. The maximal vapor and condensation aerosol concentration achieved was 970 ppm. The cats "displayed vomiting tendencies."* No other signs were noted. A single 8-h exposure to "concentrated vapors" did not kill any of six rats (Union Carbide Corporation, 1970). Guinea pigs survived a 15-min exposure to ethanolamine st 193 ppm, but four of six died after 1-h exposure at 233 ppm (Treon et al., 1957).

^{*} Translation.

Smyth et al. (1951) obtained the following results in a 90-d dietary feeding study in rats in which the minimal daily dose was 160 mg/kg and the maximal, 2,670 mg/kg:

Maximal daily dose with no effect:

320 mg/kg

Minimal daily dose that produced

altered liver or kidney weight:

640 mg/kg

Minimal daily dose that produced histopathologic lesions and

deaths:

1,280 mg/kg

Investigators at the Kettering Laboratory (Treon et al., 1957) conducted a series of animal inhalation exposures to ethanolamine. The exact amount of free ethanolamine delivered to the animals is unknown, because carbon dioxide in the stream of dried air converted some of the ethanolamine to a carbonate. Dogs and cats survived concentrations of 990 ppm for 7 h on each of four consecutive days. Rats, rabbits, and mice were less susceptible than guinea pigs, but more suspectible than cats or dogs. Of 61 animals, 60 survived exposure at either 104 or 108 ppm for 7 h on each of five consecutive days. All but one of 26 mice survived exposure at 51 ppm for 7 h on each of 25 d over 5 wk. At autopsy, the dogs and cats appeared normal. Animals of the other species that died showed acute pulmonary irritation. The survivors had acute bronchitis and pneumonia superimposed on the pulmonary lesions.

Rats, guinea pigs, and dogs were exposed to ethanolamine for 24 h/d for 1-90 d at mean concentrations of 5-102 ppm (Weeks et al., 1960). High concentrations (66-102 ppm in dogs, 66-75 ppm in rodents) for 30 d caused extensive damage (to skin, liver, and kidney) and some deaths. Dogs exposed at 102 ppm showed some biochemical and hematologic changes; these changes were not seen in the other animals. Intermediate concentrations (12-26 ppm) for 90 d caused no fatalities during the 90-d study. Dogs exposed at 12 or 26 ppm and rodents exposed at 12-15 ppm became lethargic early in the study; the dogs recovered, but the rodents did not. The skin of all animals became irritated. Male dogs and immature rats of both sexes were exposed at 5 or 6 ppm for 40 d (rats) or 60 d (dogs). Some decrease in activity was observed. Skin irritation was also noted, although much less than in the animals exposed at high concentrations.

No data were found on chronic, carcinogenic, teratogenic, or reproductive effects of exposure to ethanolamine. This compound does not appear to be mutagenic in the Salmonella typhimurium reversemutation bioassay with and without S-9 using strains TA 1535 and TA 100 (Hedenstedt, 1978). Inoue et al. (1982) reported that ethanolamine was very toxic at $500~\mu\text{g/ml}$ in a cell-transformation assay; it did not produce any morphologic transformation at 25-500 $\mu\text{g/L}$.

PHARMACOKINETICS

Ethanolamine is a normal constituent of mammalian urine (Table 5), and 6-48% of ethanolamine given to rats orally is recovered in the urine

(Luck and Wilcox, 1953). In rats, 8 h after an intraperitoneal injection of \$^{14}\$C-labeled ethanolamine, 54% of the \$^{14}\$C was found in the liver, spleen, kidneys, brain, and diaphragm, and 11.5% was accounted for as expired \$^{14}\$CO2. The liver contained approximately 50% of the \$^{14}\$C, nearly all in the lipid fraction. The main metabolic pathway is incorporation into the phospholipid fraction. The maximal rate of respiratory excretion of \$^{14}\$CO2 was observed between 1 and 2 h after administration; that indicates rapid oxidation of ethanolamine. The rate of excretion then dropped rapidly, and that suggests either dilution of the compound by endogenous ethanolamine or conversion of ethanolamine into compounds not readily catabolized (Taylor and Richardson, 1967).

Weeks et al. (1960) cited unpublished data from the Mellon Institute that showed that, in dogs treated with Nembutal (sodium pentobarbital), ethanolamine either stimulated (at low doses) or depressed (at lethal doses) the central nervous system.

INHALATION EXPOSURE LIMITS

The ACGIH (1983) recommended an 8-h TLV-TWA of 3 ppm and a 15-min TLV-STEL of 6 ppm. The OSHA (1983) standard is an 8-h PEL (permissible exposure limit) of 3 ppm. Sidorov and Timofeevskaya (1979) suggested an occupational exposure limit of 0.2 ppm.

COMMITTEE RECOMMENDATIONS

The toxicity of ethanolamine was studied by the Committee on Toxicology in 1967. The study of Weeks et al. (1960) indicated that skin and eye irritation and immediate signs of irritability and restlessness followed by central nervous system depression are the major adverse effects seen in unanesthetized experimental animals exposed to ethanolamine at 12-26 ppm for 24 h. Continuous exposure at 5-6 ppm produced some behavioral changes in animals, but only after 2-3 wk of exposure. On the basis of these data, the committee believes that the previous recommendations are adequate. However, we note that additional data could help to refine these exposure limits. For example, reports concerning dose- and time-dependent central nervous system changes after ethanolamine exposure need to be confirmed and elaborated. It would be desirable to have better quantitative data on these changes. Although behavioral-toxicology studies are relatively new and their results are difficult to interpret, some of them might. be incorporated into the studies suggested above or run separately. It would be most useful if more data on accidental exposure to humans or 40-h/wk occupational exposure could be gathered, including estimated concentrations, symptoms (if any), clinical findings, and sequelae.

The present Committee's recommended EELs and CEL for ethanolamine and the limits proposed in 1967 are shown below.

	<u>1967</u>	<u>1984</u>
60-min EEL	50 ppm	50 ppm
24-h EEL	3 ppm	3 ppm
90-d CEL	0.5 ppm	0.5 ppm

TABLE 4 LD50 Values of Ethanolamine

Species (Sex)a	Route	LD ₅₀ ,	mg/kg ^b	Reference
Rat (male)	Ora1	1,970	(1,430-1,710)	Vernot et al., 1977
Rat (female)	Oral	1,720	(1,150-2,540)	Vernot et al., 1977
Rat (male)	Oral	3,320	(1,710-4,070)	Hartung and Cornish, 1968
Rat (male) ^c	Oral	2,740	(2,390-3,150)	Smyth et al., 1951
Rat	Ora1	2,050		Sidorov et al., 1968
Mouse	0ral	1,475		Sidorov <u>et al.</u> , 1968
Rabbit	Oral	1,000		Sidorov <u>et al.</u> , 1968
Guinea pig	Oral	620		Sidorov et al., 1968
Rat	Subcutaneous	1,500		Sidorov et al., 1968
Rat	Intramuscular	1,750		Sidorov et al., 1968
Rat	Intravenous	225		Sidorov et al., 1968
Rat	Intraperitoneal	67		Sidorov et al., 1968

b Ranges in parentheses.
c Animals not fasted.

TABLE 5 Urinary Excretion of Ethanolamine

		Excretion rateb		
Species	N	mg/d	mg/d-kg b.w.	
Human:				
Males	8	12.2	0.162	
		(4.8-22.9)	(0.071-0.293)	
Females	11	29.9	0.491	
Dans		(12.9-49.8)	(0.291-0.930)	
Rat: Males	12	0.41	1.46	
		(0.33-0.52)	(1.21-1.89)	
Females	11	0.40	1.26	
		(0.36-0.45)	(0.81-1.62)	
Cat	2	(1.53-1.91)	(0.443-0.465)	
Rabbit	2	(3.02-3.10)	(0.80-1.01)	
Dog	1	3.10	0.80	

a Data from Luck and Wilcox, 1953. b Ranges in parentheses.

REFERENCES

American Conference of Governmental Industrial Hygienists. 1983. TLVs(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Bioenvironmental Safety News Letter. 1972. Second monoethanolauine injury reported. p.40 (April).

Browning, E. 1953. Toxicity of Industrial Organic Solvents. New York: Chemical Publishing Co. 411 p.

Carpenter, C.P., and Smyth, H.F., Jr. 1946. Chemical burns of the rabbit-cornea. Am. J. Ophthalmol. 29:1363-1372.

Grant, W.M. 1962. Toxicology of the Eye. Springfield, Ill.: Charles C Thomas. 641 p.

Hartung, R., and Cornish, H.H. 1968. Cholinesterase inhibition in the acute toxicity of alkyl-substituted 2-aminoethanols. Toxicol. Appl. Pharmacol. 12:486-494.

Hedenstedt, A. 1978. Mutagenicity screening of industrial chemicals: Seven aliphatic amines and one amide tested in the Salmonella/microsomal assay. Mutat. Res. 53:198-199, abstr. no. 90.

Hinglais, H. 1947. Method of measuring irritation and necrosis produced by chemical derivatives [amino alcohols] on the skin and mucous membranes. Produits Pharm. 2:445-449.

Inoue, K., Sunakawa, T., Okamoto, K., and Tanaka, Y. 1982. Mutagenicity tests and in vitro transformation assays on triethanolamine. Mutat. Res. 101:305-313.

Luck, J.M., and Wilcox, A. 1953. On determination of ethanolamine in urine and factors affecting its daily output. J. Biol. Chem. 205:859-866.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29CFR 1910.1000.

Paustovskaia, V.V., Krashniuk, E.P., Onikienko, F.A., and Vasiliuk, L.M. 1977. Effect of working conditions on health of persons working with inhibitors of atmospheric corrosion of metals. Vrach Delo. Number 4: 121-124.

Sidorov, K.K., Gorban', G.M., and Tikhonova, G.P. 1968. Comparative toxicological characteristics of some regenerable absorbers of carbon dioxide. Environ. Space Sci. 2:289-292. [Trans. from: Kosmicheskaya Biologiya i Meditsina 2:44-49]

Sidorov, K.K., and Timofeevskaya, L.A. 1979. Data for substantiation of the maximum permissible concentration of monoethanolamine in the working environment. Gig. Tr. Prof. Zabol. No. 9:55

Smyth, H.F., Jr. Carpenter, C.P., and Weil, C.S. 1951. Range-finding toxicity data: List IV. AMA Arch. Ind. Hyg. Occup. Med. 4:119-122.

Sutton, W.L. 1963. Aliphatic and alicyclic amines: Ethanolamine (2-aminoethano1). In Patty, F.A., ed. Industrial Hygiene and Toxicology. Vol. II. Toxicology, Fassett, D.W., and Irish, D.D., eds. 2nd rev. ed. New York: Interscience Publishers. p. 2064-2066.

Taylor, R.J., Jr., and Richardson, K.E. 1967. Ethanolamine metabolism in the rat. Proc. Soc. Exp. Biol. Med. 124:247-252.

Treon, J.F., Cleveland, F.P., Stemmer, K.L., Cappel, J., Larson, E.E., and Shaffer, F. 1957. Toxicity of monoethanolamine in air. Cincinnati: Kettering Laboratory.

Union Carbide Corporation. 1970. Ethanolamines. [Technical booklet] New York: Union Carbide Corporation, Chemicals and Plastics. p. 30.

Vernot, E.H., MacEwen, J.D., Haun, C.C., and Kinkead, E.R. 1977. Acute toxicity and skin corrosion data for some organic and inorganic compounds and aqueous solutions. Toxicol. Appl. Pharmacol. 42:417-423.

Weeks, M.H., Downing, T.O., Musselman, N.P., Carson, T.R., and Groff, W.A., 1960. The effects of continuous exposure of animals to ethanolamine vapor. Am. Ind. Hyg. Assoc. J. 21:374-381.

FLUOROCARBON 11

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: CClaF Molecular weight: 137.38

Chemical names: Trichlorofluoromethane,

fluorotrichloromethane

Synonyms: FC-11, Freon 11

CAS number: 75-69-4 -111°C Freezing point:

Physical state: Liquid below 23.7°C Specific gravity: 1.494 (17.2°C) Vapor density: 5.04 (air = 1)

Vapor pressure: 792 torr (25°C)

Solubility: Insoluble in water; soluble in

ethanol or alcohol

General characteristics: At ordinary ambient temperatures, a

colorless, nonflammable liquid or gas

1 ppm = 5.6 mg/m^3 1 mg/m³ = 0.18 ppmConversion factors:

OCCURRENCE AND USE

Fluorocarbon 1. (FC-11) has been used primarily as an aerosol propellant, refrigerant, and blowing agent for polymeric feams. Its use is now banned because of its potential effects on the ozone layer.

It is prepared from carbon tetrachloride and antimony trifluoride (Stecher et al., 1968; ACGIH, 1980). FC-11 may be a contaminant of submarine atmospheres.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

Inhalation of fluorocarbons during the years 1960-1970 was a prominent cause of abusive death among teen-agers. Severe cardiac arrhythmia--resulting from light plane anesthesia and intensified by hypercapnia, stress, or activity-was suggested as an explanation for 110 cases of sudden sniffing death (Bass, 1970). Typically, a person would spray the Freon into a paper bag from a commercial aerosol product and inhale it; after a few breaths and a short excitement period, death might occur. Fluorocarbons are thought to sensitize the heart to asphyxia-induced sinus bradycardia, atrioventricular block, and ventricular T-wave depression (Haj et al., 1980).

Accidental ingestion of FC-11 occurred when a healthy man mistook a bottle in a refrigerator for a bottle of plain water. This resulted in freezing, tissue necrosis, and multiple perforations of the

stomach. The patient recovered after surgery to remove the damaged tissue (Haj et al., 1980).

Labeled FC-11 administered to four healthy males by inhalation of a single breath held for 5 s was eliminated from the body rapidly. Results in humans appeared to parallel those in rats in more detailed studies (Williams et al., 1974). The investigators found rapid transfer of FC-11 to blood followed by distribution to fat, from which release was slow. Mergner et al. (1975) exposed a male and a female volunteer to radiolabeled FC-11 at 1,000 ppm for 7-17 min. Recovery of administered radioactivity in exhaled air was essentially complete (99% and 79%). Errors in collection of rapidly eliminated gases account for the differences from 100%. Only a very small fraction of the administered radioactivity (less than 0.2%) was exhaled as 14CO2 or excreted as nonvolatile urinary activity. The impurities in FC-11-namely, chloroform and carbon tetrachloride-known to be metabolized could account for all the radioactivity found in urine and exhaled CO2 after exposure to FC-11.

Cardiac effects have been studied in healthy subjects and patients with bronchopulmonary disease. None of the subjects exhibited cardiotoxic effects (Fabel et al., 1972).

Human volunteers were exposed to FC-113 (similar to FC-11) at 500 or 1,000 ppm for 6 h/d, 5 d/wk during a 2-wk period. No adverse changes were seen in performance of complex mental tasks, clinical status, or results of biochemical tests. Breath analysis did not reveal a significant buildup of FC-113 (Reinhardt et al., 1971).

EFFECTS ON ANIMALS

FC-11 has not shown appreciable oral toxicity in rats and dogs in either acute or chronic studies (Haskell Laboratory, 1970; NCI, 1978). The chronic investigations include 1-mo, 90-d, and 2-yr studies. FC-11 was tested on the intact skin of mice. It was well tolerated by the skin, but retarded the recovery of wounds and burns and regrowth of hair (Quevauviller, 1960; Quevauviller et al., 1963). Dermal application of FC-11 to rabbit skin did not produce any lesions (Scholz, 1962). Transient conjunctival irritation was observed after application of FC-11 solution to the rabbit eye. No irreversible eye damage was seen (Haskell Laboratory, 1970; Kudo et al., 1971).

The LC50 of FC-11 for rats in a 4-n exposure is 26,200 ppm (Haskell Laboratory, 1970). A 30-min exposure of rats at 50,000 ppm caused no signs of intoxication. Similar exposure at higher concentrations caused clinical signs of central nervous system depression. Concentrations of 100,000 ppm or more were fatal after less than 30 min (Lester and Greenberg, 1950). Acute exposure of other species of laboratory animals produced similar effects (Caujolle, 1964; Haskell Laboratory, 1970; Nuckolls, 1933; Scholz, 1962).

Rats, guinea pigs, monkeys, and dogs were continuously (24 h/d) exposed to FC-11 at approximately 1,000 ppm for 90 d. One monkey died on day 78, but its death was not definitely linked to exposure to FC-11. No other animals were affected. No compound-related pathologic changes were observed. Another group of animals was

exposed at 10,250 ppm, 8 h/d, 5 d/wk for 6 wk without adverse effects (Jenkins et al., 1970). In another study, dogs, cats, guinea pigs, and rats were exposed to FC-11 for 3.5 h/d, 5 d/wk for 4 wk; the dogs were exposed at 12,500 ppm, and the other animals at 25,000 ppm. No microscopic evidence of damage to the lungs, heart, spleen, liver, or kidneys was seen (Scholz, 1962).

Rats and mice exposed to FC-11 at 1,000 or 5,000 ppm for lifetime showed no evidence of carcinogenicity or other adverse health effects

(C. Maltoni, unpublished).

FC-11, like other chlorofluorocarbons and hydrocarbons, was capable of sensitizing the beagle heart to exogenous epinephrine in standard 5-min cardiac-sensitization screening studies. A 5-min cardiac-sensitization screening test consists of a control intravenous injection of epinephrine at 8 $\mu g/kg$, followed later by a 5-min exposure to fluorocarbon and then a challenge with 8 $\mu g/kg$ intravenously. Manifestation of arrhythmia (multiple consecutive ventricular beats), which is considered to pose a serious threat to life, or cardiac arrest (ventricular fibrillation) constitutes a positive test. The lowest concentration that elicited a marked response in exposed dogs was 5,000 ppm. A concentration of 1,000 ppm was ineffective. Dogs exposed while running on a treadmill (to increase their circulating epinephrine) were not sensitized at concentrations up to 10,000 ppm (Mullin et al., 1972).

Azar et al. (1973) studied nonanesthetized dogs and reported that the average blood concentrations of FC-11 associated with cardiac sensitization were 28.6 μ g/L in arterial and 19.7 μ g/L in venous blood.

Belej and Aviado (1975) studied cardiopulmonary toxicity of propellants in anesthetized dogs. They concluded that FC-11, unlike eight other halocarbon propellants studied, produced bronchodilation, rather than bronchoconstriction. It also reduced pulmonary compliance and respiratory minute volume. FC-11 had the greatest tachycardiac effect of all compounds studied. Effects of FC-11 on the circulatory system were summarized by Aviado (1975, 1978).

In a bioassay supported by the National Cancer Institute (1978), oral FC-11 was not carcinogenic in rats or mice. No significant increase in tumor formation was seen in a study that used subcutaneous injection (Epstein et al., 1967). Additionally, FC-11 has not been shown to be mutagenic in the Salmonella typhimurium reverse-mutation bioassay (C.F. Reinhardt, Haskell Laboratory, personal communication). No embryotoxic, fetotoxic, or teratogenic effect of FC-11 was shown in a study with pregnant rats and rabbits; the animals were exposed for 2 h/d to a 200,000 ppm of a FC-11/FC-12 (1:9) mixture from day 4 to 16 of gestation for rats and from day 5 to 20 for rabbits (Paulet, 1976). Blake and Mergner (1974) studied the biotransformation of 14C-labelled FC-11 (8,000-12,000 ppm) in male and female beagles after a short (6-20 min) inhalation. Essentially all the inhaled fluorocarbon was recovered in the exhaled air within 1 h. Only traces of radioactivity were found in urine or exhaled CO2. The investigators concluded that FC-il is relatively refractory to biotransformation after a short inhalation exposure and that it is rapidly exhaled chemically unaltered.

engin en la proposition de la company de la company

INHALATION EXPOSURE LIMITS

The American Conference of Governmental Industrial Hygienists (1980, 1983) recommended a ceiling of 1,000 ppm. The Occupational Safety and Health Administration's (1983) permissible exposure limit currently in effect for FC-11 is a ceiling of 1,000 ppm.

1992年 1997年 1998年 1988年 1988

COMMITTEE RECOMMENDATIONS

The previous EELs and CEL were established by the Committee on Toxicology in 1966. No adverse effects have been observed in dogs, monkeys, guinea pigs, or rats continuously exposed to FC-11 at 1,000 ppm for 90 d or in a similar group repeatedly exposed at 10,250 ppm, 8 h/d, 5 d/wk for 6 wk (Jenkins et al., 1970). Dogs exposed at 12,500 ppm and cats, guinea pigs, and rats at 25,000 ppm for 4 wk were not affected (Scholz, 1962). Human exposure to FC-113 (a compound similar to FC-11) at 1.500 ppm produced no adverse effects after 2.75 h. Signs of central nervous system involvement were seen after exposure to FC-113 at 2,500 ppm for 30 min. These effects were reversible, and the volunteers appeared normal 15 min after cessation of the experiment (C.F. Reinhardt, Haskell Laboratory, personal communication). FC-11 can sensitize the mammalian heart to epinephrine and result in serious cardiac arrhythmia. However, the possible combined effects of excitement-stimulated epinephrine release and FC-11 on the heart are not easy to predict. It would therefore be prudent to take a more cautious approach to EEL recommendations than was taken by the Committee in 1966, when it was not aware of the sudden-sniffing-death syndrome. The previous 60-min and 24-h EELs are too high, on the basis of experimental cardiac sensitization of dogs, which occurred when they were exposed at 5,000 ppm and given a large challenge injection of epinephrine. However, no sensitization occurred in resting dogs exposed at 1,000 ppm and given epinephrine or in exercising dogs exposed at 10,000 ppm (Mullin et al., 1972).

Based on the no-observed-adverse-effect concentration of FC-113 in humans (1,500 ppm for 2.5 h), the 10,000-ppm concentration (which did not cause cardiac arrhythmia in exercising dogs), and the results in standard 5-min cardiac-sensitization screening tests in dogs, the Committee recommends a 60-min EEL of 1,500 ppm. It bases its 24-h EEL on the finding in humans that repeated exposure to FC-113 at 500 or 1,000 ppm for 2 wk did not result in adverse effects. Finally, using the no-observed-effect concentration of 1,000 ppm in a continuous-exposure animal study and applying an uncertainty factor of 10, the Committee arrives at a recommended CEL of 100 ppm.

The present Committee's recommended EELs and CEL for FC-11 and the limits proposed in 1966 are shown below.

	1966	1984
60-min EEL	30,000	1,500 ppm
24-hr EEL	20,000	500 ppm
90-d CEL	1,000	100 ppm

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Trichlorofluoromethane. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 408-409.

一分 とない アプランド こうしょうじゅう

かんし いいいいい

うりがいこと は私をおりる さいかいかい ていかんな かんかいかい

一方ののこののです 動いがながななが

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R). Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Aviado, D.M. 1975. Toxicity of aerosol propellants in the respiratory and circulatory systems. IX. Summary of the most toxic: Trichlorofluoromethane (FC11). Toxicology: 3:311-319.

Aviado, D.M. 1978. Effects of fluorocarbons, chlorinated solvents, and inosine on the cardiopulmonary system. Environ. Health Perspect. 26:207-215.

Azar, A., Trochimowicz, H.J., Terrill, J.B., and Mullin, L.S. 1973. Blood levels of fluorocarbon related to cardiac sensitization. Am. Ind. Hyg. Assoc. J. 34:102-109.

Bass, M. 1970. Sudden sniffing death. J. Am. Med. Assoc. 212:2075-2079.

Belej, M.A., and Aviado, D.M. 1975. Cardiopulmonary toxicity of propellants for aerosols. J. Clin. Pharmacol. 15:105-115.

Blake, D.A., and Mergner, G.W. 1974. Inhalation studies on the biotransformation and elimination of [14C]trichlorofluoromethane and [14C]dichlorofluoromethane in beagles. Toxicol. Appl. Pharmacol. 30:396-407.

Caujolle, F. 1964. Comparative toxicity of refrigerants. Bull. Inst. Intern. Froid 44:20-55. [In English and French]

Epstein, S.S., Joshi, S., Andrea, J., Clapp, P., Falk, H., and Manteí, N. 1907. Synergistic toxicity and carcinogenicity of 'freons' and piperonyl butoxide. Nature 214:526-528.

Fabel, H., Wettengel, R., and Hartmann, W. 1972. [Myocardisl ischemia and arrhythmias due to the use of pressurized aerosols in man.] Dtsch. Med. Wochenschr. 97:428-431. (in German)

Haj, M., Burstein, Z., Horn, E., and Stamler, B. 1980. Perforation of the stomach due to trichlorofluoromethane (Freon 11) ingestion. Isr. J. Med. Sci. 16:392-394.

Haskell Laboratory. 1970. Unpublished data. News., Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine.

Jenkins, L.J., Jr., Jones, R.A., Coon, R.A., and Siegel, J. 1970. Repeated and continuous exposures of laboratory animals to trichlorofluoromethane. Toxicol. Appl. Pharmacol. 16:133-142.

Kudo, K., Toida, S., Matsuura, S., Sasaki, T., and Kawamura, H. 1971. Comparison of Freon-11S and Freon-11. Acute, subacute toxicity and irritation of mucous membrane. Toho Igakkai Zasshi 18(2):363-367.

Lester, D., and Greenberg, L.A. 1950. Acute and chronic toxicity of some halogenated derivatives of methane and others. AMA Arch. Ind. Hyg. Occup. Med. 2:335-344.

Mergner, G.W., Blake, D.A., and Helrich, M. 1975. Biotransformation and elimination of ¹⁴C - trichlorofluoromethane (FC-11) and ¹⁴C-dichlorodifluoromethane (FC-12) in man. Anesthesiology 42:345-351.

Mullin, L.S., Azar, A., Reinhardt, C.F., Smith, P.E., Jr., and Fabryka, E.F. 1972. Halogenated hydrocarbon-induced cardiac arrhythmias associated with release of endogenous epinephrine. Am. Ind. Hyg. Assoc. J. 33:389-396.

National Cancer Institute. 1978. Bioassay of Trichlorofluoromethane for Possible Carcinogenicity. NCI-CG-TR-106. Bethesda, Maryland: U.S. Department of Health, Education, and Welfare, Public Health Service, National Institutes of Health. [84] p. [DHEW Publ. No. (NIH) 78-1356]

Nuckolls, A.H. 1933. The comparative life, fire, and explosion hazards of common refrigerants. Chicago, Illinois: National Board of Fire Underwriters, Underwriters' Laboratories. [Misc. Hazard No. 2375] 119 p.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Faulet, G. 1976. Fluorocarbons in question. Eur. J. Toxicol. Environ. Hyg. 9(7 Suppl.):385-407.

Quevauviller, A. 1960. Skin tolerance of chlorofluoromethaues used as propellants in cosmetology. Parfum. Cosmet. Savons 3:228-230. [Chem. Abs. 54:19965e, 1960]

CONTRACTOR SECURE CONTRACTOR SECURITIES

۴,

Quevauviller, A., Chaigneau, M., and Schrenzel, M. 1963. [Experimental studies with mice on tolerance of lungs to chlorofluorinated hydrocarbons.] Ann. Pharm. Franc. 21:727-734.

Reinhardt, C.F., McLaughlin, M., Maxfield, M.E., Mullin, L.S., and Smith, P.E., Jr. 1971. Human exposures to fluorocarbon 113 (1,1,2-trichloro-1,2,2-trifluoroethane). Am. Ind. Hyg. Assoc. J. 32:143-152.

Scholz, J. 1962. New toxicologic investigations of freons used as propellants for aerosols and sprays. Translation of: Fortschr. Biol. Aerosol-Forsch. Jahren 1957-1961, Ber. Aerosol-Kongr. 4:420-429.

Stecher, P.G., Windholz, M., and Leahy, D.S., eds. 1968. The Merck Index: An Encyclopedia of Chemicals and Drugs. 8th ed. Rahway, New Jersey: Merck & Co., Inc. p. 1069.

Williams, F.M., Draffan, G.H., Dollery, C.T., Clark, J.C., Palmer, A.J., and Vernon, P. 1974. Use of ¹⁸F-labelled fluorocarbon-11 to investigate the fate of inhaled fluorocarbons in man and in the rat. Thorax 29:99-103.

FLUOROCARBON 12

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: CCL₂F₂
Molecular weight: 120.92

Chemical name: Dichlorodifluoromethane

Synonyms: FC-12, Freon 12

CAS number: 75-71-8
Melting point: -158°C
Boiling point: -29.8°C

Specific gravity: 1.1834 (57°C)
Vapor pressure: 5.7 atm (20°C)

Solubility: Soluble in alcohol and ether;

insoluble in water

General characteristics: Nonflammable, colorless

gas

Conversion factors: 1 ppm $\frac{1}{2}$ 4.94 mg/m³

 $1 \text{ mg/m}^3 = 0.2 \text{ ppm}$

OCCURRENCE AND USE

Fluorocarbon 12 (FC-12) has been used primarily as an aerosol propellant, refrigerant, and blowing agent for polymeric foams. Air conditioning is the suspected source of FC-12 contamination in submarines.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON IFFIANS

Of 34 case of sudden death reported by the medical examiner of Dallas. Texts, to be due to abusive, deliberate use of products that can be inhated, 16 involved fluorocarbons (Garriott and Petty, 1980). The victims were mainly white males aged 13-30 (mean, 18.4). FC-11 and FC-114 were used in addition to FC-12, and many products contained mixtures of FC-11 and FC-12. The sequence of events has been summarized by Aviado (1978) and others as follows: sensitization of the heart to prearrhythmic effects of epinephrine, depression of myocardial contractility, reduction in cardiac output, and irritation of mucosa in upper and lower respiratory tract that causes an increase in sympathetic and vagal impulses to the heart.

Azar et al. (1972) exposed two human volunteers to FC-12 at 1,000 and 10,000 ppm for 2.5 h. Each volunteer was exposed twice at each concentration. Electrocardiographic monitoring and psychomotor performance tests revealed no adverse effects of exposure at 1,000 ppm. At 10,000 ppm, there was only a 7% reduction in psychomotor-test score. Rapid elimination from the lungs occurred when exposure stopped. The investigators concluded that a single brief exposure

(up to 2.5 h) to FC-12 at 10,000 ppm could be tolerated without adverse health effects.

Kehoe (1943) reported that exposure to FC-12 at up to 60,000 ppm was tolerated for 80 min by one human subject. When exposed at 40,000 ppm for 14 min and then at 20,000 ppm for 66 min, a second subject developed EEG changes and had slurred speech and decreased scores in psychologic tests. At 110,000 ppm, amnesia and cardiac arrhythmia occurred within about 10 min.

Mergner et al. (1975) exposed a male and a female volunteer to radiolabeled FC-12 at 1,000 ppm for 7-17 min. Recovery of administered radioactivity in exhaled air was essentially complete. Radioactivity in urine and exhaled CO_2 together amounted to less than 0.2% of administered radioactivity.

EFFECTS ON ANIMALS

FC-12 has not shown appreciable oral toxicity in laboratory animals in either acute or chronic exposures. These included 1d-wk rat and dog feeding studies (fed at 160-379 mg/kg per day) and 2-yr rat feeding studies (up to 150 mg/kg per day) (Haskell Laboratory, 1955a; Hood, 1956; Sherman, 1974; Sherman and Barnes, 1966).

No skin irritation was observed after contact of the gas with the skin of rats, rabbits, or guinea pigs. Transient eye irritation followed application of a 50% solution to rabbit eyes. Treated eyes were normal 24 h later. No eye irritation was seen after an aerosol mixture containing FC-12 was sprayed into the eyes of rabbits (Haskell Laboratory, 1955b; Hood, 1956).

The lethal concentration of FC-12 for rats in a 3-h exposure was 620,000 ppm (Shugaev, 1963). Two-hour exposure at 600,000 ppm was lethal to rats, but not to guinea pigs (Scholz, 1962). Central nervous system effects were observed in all exposed species. In rats exposed for 30 min, there were no reactions at 200,000 ppm (Lester and Greenberg, 1950). At higher concentrations, the following effects were observed: muscular twitching and tremors at 300,000-400,000 ppm, loss of postural reflex at 500,000 ppm, and loss of righting reflex at 600,000 ppm. Guinea pigs were similarly affected at 200,000-300,000 ppm (Nuckolls, 1933). Mice survived a 24-h exposure at 10,000 ppm, but microscopic examination revealed nonspecific lung changes (Quevauviller et al., 1963).

Rats, guinea pigs, monkeys, rabbits, and dogs were exposed to FC-12 continuously at 810 ppm for 90 d (Prendergast et al., 1967). Although 2 of 15 rats and 1 of 15 guinea pigs died during exposure, there were no visible signs of toxicity. During the course of the experiment—which involved tests of several other chemicals, such as trichloroethylene, carbon tetrachloride, and other fluorocarbons—7 of 304 control rats and 2 of 34 control guinea pigs died. Pathologic examination revealed focal necrosis in the livers of the guinea pigs. This change was thought to be due to the continuous nature of the exposure or the high degree of susceptibility of the guinea pig. No pathologic changes were seen in the tissues of the other four species. In another experiment, test animals were exposed at 840 ppm for 8 h/d, 5 d/wk, for 6 wk (Nuckolls, 1933). No signs of toxicity were seen,

and pathologic examination revealed changes similar to those seen in the continuous study (Nuckolls, 1933). At higher concentrations, toxic signs indicative of CNS effects were observed on repeated exposure (Sayers et al., 1930; Scholz, 1962; Watanabe and Aviado, 1975). In addition, rats and mice exposed to FC-12 at 1,000 and 5,000 ppm for 104 wk showed no evidence of carcinogenicity (C. Maltoni, unpublished).

FC-12, like other chlorofluorocarbons and hydrocarbons, is capable of sensitizing the beagle heart to exogenous epinephrine in standard 5-min cardiac-sensitization screening studies. The concentration needed to elicit marked responses in 50% of exposed dogs is 80,000 ppm (Clark and Tinston, 1973). Dogs exposed to FC-12 while running on a treadmill to increase their own epinephrine concentration were sensitized at concentrations over 100,000 ppm (Mullin, 1970; Mullin et al., 1972).

Lessard et al. (1978) reported that epinephrine perfusion induced cardiac arrhythmia in rabbits and dogs breathing gas mixtures of 79% FC-12 and 21% 02. After 5 min of inhalation, arterial blood contained FC-12 at 0.8 mg/L in rabbits and 0.7 mg/L in dogs.

Blake and Mergner (1974) studied the biotransformation and elimination of FC-12 in beagles after exposure to radiolabeled FC-12 (8,000-12,000 ppm, v/v). Essentially all the inhaled air was recovered in the exhaled air within 1 h. Only traces of radioactivity were found in urine or exhaled CO_2 . All tissues contained measurable concentrations of nonvolatile radioactivity 24 h after exposure, but together represented less than 1% of the administered dose. It was not possible to determine whether this radioactivity was associated with metabolites of FC-12 or with unavoidable radiolabeled impurities present in the administered gas mixture. The investigators concluded that FC-12 is relatively refractory to biotransformation after a short inhalation exposure and that it is rapidly exhaled in its unaltered form.

INHALATION EXPOSURE LIMITS

The ACGIH TLV-TWA (1983) and the OSHA federal standard (1983) for FC-12 are both 1,000 ppm; ACGIH recommended a TLV-STEL for 15-min excursions of 1,250 ppm.

COMMITTEE RECOMMENDATIONS

The previous EELs and CEL were established by the Committee in 1966. On the basis of a small decrement in human psychomotor performance at 10,000 ppm and none at 1,000 ppm for 2.5 h and positive findings in dogs in the 5-min cardiac-sensitization screening test at 80,000 ppm and in exercising dogs at 100,000 ppm, the Committee recommends a 60-min EEL of 10,000 ppm for FC-12. Because FC-12 is rapidly eliminated in expired air, the Committee recommends a 24-h EEL of 1,000 ppm and a CEL of 100 ppm.

The present Committee's recommended EELs and CEL for FC-12 and the limits proposed in 1966 are shown below.

	1966	1984	
60-min EEL	30,000 ppm	10,000 ppm	
24-hr EEL	20,000 ppm	1,000 ppm	
90-d CEL	1,000 ppm	100 ppm	

REFERENCES

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Aviado, D. 1978. Physiological and biochemical responses to specific group of inhalants: concluding remarks. Fed. Proc. Fed. Am. Soc. Exp. Biol. 37:2508-2509.

Azar, A., Reinhardt, C.F., Maxfield, M.E., Smith, P.E., Jr., and Mullin, L.S. 1972. Experimental human exposures to fluorocarbon 12 (dichlorodifluoromethane). Am. Ind. Hyg. Assoc. J. 33:207-216.

Blake, D.A. and Mergner, G.W. 1974. Inhalation studies on the biotransformation and elimination of [14C]trichlorofluoromethane and [14C]dichlorofluoromethane in beagles. Toxicol. Appl. Pharmacol. 30:396-407.

Clark, D.G., and Tinston, D.J. 1973. Correlation of the cardiac sensitizing potential of halogenated hydrocarbons with their physicochemical properties. Br. J. Pharmacol. 49:355-357.

Garriott, J., and Petty, C.S. 1980. Death from inhalant abuse: Toxicological and pathological evaluation of 34 cases. Clin. Toxicol. 16:305-315.

Haskell Laboratory for Toxicology and Industrial Medicine. 1955a. Eye irritation of Freon 12 in rabbits. Newark, Delaware: E.I. du Pont de Nemours & Co. [Unpublished]

Haskell Laboratory for Toxicology and Industrial Medicine. 1955b. Acute oral toxicity of Freon 12, dichlorodifluoromethane. Newark, Delaware: E.I. du Pont de Nemours & Co. [Unpublished]

Hood, D.B. 1956. Skin and eye irritation tests with aerosol hair sprays. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine [3] p. [Unpublished]

Kehoe, R.A. 1943. [Unpublished Data on Dichlorodifluoromethane]. Cincinnati, Ohio: University of Cincinnati, Kettering Laboratory of Applied Physiology, College of Medicine. 22 p.

Lessard, Y., Desbrousses, S., and Paulet, G. 1978. Rôle de l'adrénaline endogène dans le déclenchement de l'arythmie cardiague par le dichlorodifluorométhane (FC 12) chez les mammifères. C.R. Seances Soc. Biol. Paris 172:337-347.

Lester, D., and Greenberg, L.A. 1950. Acute and chronic toxicity of some halogenated derivatives of methane and ethane. AMA Arch. Ind. Hyg. Occup. Med. 2:335-344.

Mergner, G.W., Blake, D.A., and Helrich, M. 1975. Biotransformation and elimination of $^{14}\text{C-trichlorofluoromethane}$ (FC-11) and $^{14}\text{C-dichlorodifluoromethane}$ (FC-12) in man. Anesthesiology 42:345-351.

Mullin, L.S. 1970. Cardiac sensitization - fright exposures. Report No. 81-70. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [5] p. [Unpublished]

Mullin, L.S., Azar, A., Reinhardt, C.F., Smith, P.E., Jr., and Fabryka, E.F. 1972. Halogenated hydrocarbon-induced cardiac arrhythmias associated with release of endogenous epinephrine. Am. Ind. Hyg. Assoc. J. 33:389-396.

Nuckolls, A.H. 1933. The comparative life, fire, and explosion hazards of common refrigerants. Chicago, Illinois: National Board of Fire Underwriters, Underwriters' Laboratories. [Misc. Hazard No. 2375]. 119 p.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Prendergast, J.A., Jones, R.A., Jenkins, L.J., Jr., and Siegel, J. 1967. Effects on experimental animals of long-term inhalation of trichloroethylene, carbon tetrachloride, 1,1,1-trichloroethane, dichlorodifluoromethane, and 1,1-dichloroethylene. Toxicol. Appl. Pharmacol. 10:270-289.

Quevauviller, A., Chaigneau, M., and Schrenzel, M. 1963. [Experimental studies with mice on tolerance of lungs to chlorofluorinated hydrocarbons.] Ann. Pharm. Franc. 21:727-734. (in French)

Sayers, R.R., Yant, W.P., Chornyak, J., and Shoaf, H.W. 1930. Toxicity of dichloro-difluoro methane, a new refrigerant. U.S. Bureau of Mines. Report of Investigations, R.I. 3013. Washington, D.C.: U.S. Bureau of Mines, Department of Commerce. 15 p.

化合物 化多数分离 医多数多数 医多数多数 医多数多数 医多数多数 医多数多数 医多数多数 医多数多数 医多种毒素

Scholz, J. 1962. New toxicologic investigations of freons used as propellants for aerosols and sprays. Translation of: Fortschr. Biol. Aerosol-Forsch. Jahren 1957-1961, Ber. Aerosol-Kongr. 4:420-429.

Sherman, H., and Barnes, J.R. 1966. Feeding studies with dichlorodifluoromethane (Freon-12^(R) food freezant). Report No. 12-66. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [18] p. [Unpublished]

Sherman, H. 1974. Long-term feeding studies in rats and dogs with dichlorodifluoromethane (Freon^(R) 12 food freezant). Report No. 24-74. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [21] p. [Unpublished]

Shugaev, V.A. 1963. The toxicology of refrigerant Freon-12. In: Levine, B.S., ed. U.S.S.R. Literature on Air Pollution and Related Occupational Diseases. Vol. 12. A survey. 1966. p. 161-163. [Translation of: Gig. Sanit. 28(6):95-97, 1963]. Available from Natl. Technical Information Service, Springfield, VA as TT 66-61429.

Stecher, P.G., Windholz, M., and Heahy, D.S., es. 2968. The Merck Index: An Encylopedia of Chemicals and Drugs. 8th ed. Rahway, New Jersey: Merck & Co., Inc. p. 351-352.

Watanabe, T., and Aviado, D.M. 1975. Subacute inhalational toxicity of aerosol propellants. Pharmacologist 17:192, abstr. 097.

and the second of the second o

FLUOROCARBON 21

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: CHCl₂F Molecular weight: 102.92

Chemical name: Dichlorofluoromethane

Synonyms: FC-21, fluorocarbon 21, Freon 21

CAS number: 75-43-4
Melting point: -135°C
Boiling point: 8.9°C

Specific gravity: 1.405 (9°C)

Solubility: Insoluble in water; soluble in

alcohol and ether

General characteristics: Colorless, nearly odorless,

nonflammable, heavy gas

中,他们也是一个一个,我们是一个人的,我们是一个人的,我们是是一个人的,我们也是一个人的,我们也会会会会会会会,我们也会会会会会会会会,这一个人的人的人,也可以

Conversion factors: 1 ppm = 4.2 mg/m³

 $1 \text{ mg/m}^3 = 0.24 \text{ ppm}$

OCCURRENCE AND USE

Dichlorofluoromethane (FC-21) is used primarily as a solvent, as a refrigerant, in aerosol propellants, and in fire extinguishers. Recently, it has found use as a heat-transfer liquid in the space-shuttle program.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

The Committee is aware of no data on human exposure to FC-21.

EFFECTS ON ANIMALS

The lethal concentration of FC-21 for rats in a 4-h exposure was 49,900 ppm (Tappan and Waritz, 1964). A 2-h exposure of guinea pigs at 50,000-52,000 ppm was not fatal, but exposure produced loss of coordination in 3 min and unconsciousness within 30 min (Nuckolls, 1935). Exposure at 100,000 ppm killed rats and guinea pigs within 1 h (Weigand, 1971). Central nervous system effects (loss of coordination, tremors, and narcosis) were observed in both exposed species.

Contact of propylene glycol solutions of FC-21 with the skin of guinea pigs produced mild irritation at FC-21 concentrations of 25-40% (Goodman, 1976; Hood, 1964a). No irritation was seen at 2.5%. No evidence of sensitization potential was observed.

Transient eye irritation was seen after instillation of liquid FC-21 chilled to the temperature of dry ice or of a 40% solution in propylene glycol into the eyes of rabbits (Brittelli, 1976; Hood, 1964b). Mild lacrimation, but no corneal or iritic effect, was seen after FC-21 was sprayed directly into the eyes of rabbits (Hocd, 1964b).

A group of 10 rats exposed at 10,000 ppm, 6 h/d, 5 d/wk, for 2 wk all survived, but pathologic examination revealed liver damage (Trochimowicz et al., 1977b). In a later 90-d study, groups of 54 rats and 4 dogs were similarly exposed at 1,000 and 5,000 ppm (Trochimowicz et al., 1977a). Excessive mortality and bilateral hair loss were seen in both groups of rats. The dogs lost weight at both concentrations. Histopathologic examination revealed cirrhosis in all rats, but only minimal changes in the livers of dogs exposed at 5,000 ppm. No compound-related effects were seen in dogs exposed at 1,000 ppm. Another incomplete 90-d study reported gross pathologic changes in the livers of rats exposed at 500 ppm, probable changes at 200 ppm, but no gross effects at 50 ppm (Allied Chemical, unpublished report to TLV Committee, 1978).

FC-21, like other chlorofluorocarbons and hydrocarbons, is capable of sensitizing the beagle heart to exogenous epinephrine in 5-min cardiac-sensitization screening studies (Mullin, 1975). A concentration of 10,000 ppm produced a marked response in 2 of 12 exposed dogs. No response was seen at 5,000 ppm; at this concentration, FC-21 is considered slightly less cardiotoxic than FC-11, which produced sensitization at 5,000 ppm. In the monkey, respiratory depression and tachycardia were seen after 5 min at 25,000 ppm (Aviado and Smith, 1975).

Kelly et al. (1978) exposed 25 pregnant rats to FC-21 at 10,000 ppm 6 h/d on days 6-15 of gestation. There was an unspecified adverse effect on maternal weight and a preimplantation loss of fertilized ova in 15 of the 25 rats. No teratogenic effects were observed.

INHALATION EXPOSURE LIMITS

The ACGIH (1980, 1983) established a TLV-TWA for FC-21 of 10 ppm (40 mg/m^3) . This represented a downward revision from the previous TLV-TWA of 1,000 ppm and was recommended because data had suggested that FC-21 was considerably more hepatotoxic than closely related fluorinated compounds and was similar in that respect to chloroform (ACGIH, 1980). OSHA (1983) recommended a permissible exposure level (PEL) of 1,000 ppm.

COMMITTEE RECOMMENDATIONS

On the basis of 5-min cardiac-sensitization screening findings in dogs, which were positive at 10,000 ppm and negative at 5,000 ppm; hepatotoxic effects in rats exposed at 10,000 ppm over a 2-wk period; and reproductive changes with exposure at 10,000 ppm at a critical period in gestation, the Committee recommends a 60-min EEL of 100 ppm.

For 24-h and 90-d exposures, the most relevant data are those from the two 90-d studies in rats and dogs. In rats, 50 ppm was a no-observed-adverse-effect concentration; 1,000 ppm was the minimal-effect concentration in dogs, causing weight loss. On the basis of the subchronic data in animals, which suggest that the toxicity of FC-21 is greater than that of other fluorocarbons and similar to that of chloroform, the Committee has applied an uncertainty factor of 1,000 to the minimal-effect concentration in dogs and recommends a 90-d CEL of 1 ppm. For a single 24-h exposure, a somewhat higher concentration would be expected not to have adverse effects, and a 24-h EEL of 3 ppm is recommended.

The Committee's recommended EELs and CEL for FC-21 are shown below.

		1984	
60-min	EEL	100	ppm
24-h	EEL	3	ppon
90-d	CEL	1	ppm

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Dichlorofluoromethane. Documentation of the Threshold Limit Values. 4th ed. Concinnati, Ohio: American Conference of Governmental Industrial Hygienists p. 132.

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Aviado, D.M., and Smith, D.G. 1975. Toxicity of aerosol propellants in the respiratory and circulatory systems. VIII. Respiration and circulation in primates. Toxicology 3:241-252.

Brittelli, M.R. 1976. Eye irritation tests in rabbits with dichlorofluoromethane. Report No. 751-75. Newark, Delaware: E.I. du Pont de Nemours and Co., Haskell Laboratory for Toxicology and Industrial Medicine. 2 p. [Unpublished]

Goodman, N.C. 1976. Primary skin irritation and sensitization tests on guinea pig with methane, dichlorofluoro. Report No. 750-75. Newark, Delaware: E.I. du Pont de Nemours and Co., Haskell Laboratory for Toxicology and Industrial Medicine. 2 p. [Unpublished]

Hood, D.B. 1964a. Freon-21: Eye irritation, skin irritation and sensitization tests. Report No. 105-64. Newark, Delaware: E.I. du Pont de Nemours and Co., Haskell Laboratory for Toxicology and Industrial Medicine. 3 p. [Unpublished]

Hood, D.B. 1964b. Eye irritation tests with Freon-21. Report No. 107-64. Newark, Delaware: E.I. du Pont de Nemours and Co., Haskell Laboratory for Toxicology and Industrial Medicine. 2 p. [Unpublished]

Kelly, D.P., Culik, R., Trochimowicz, H.J., and Fayerweather, W.E. 1978. Inhalation teracology studies on three fluorocarbons. Toxicol. Appl. Pharmacol. 45:293, abstr. no. 170.

Mullin, L.S. 1975. Cardiac sensitization with Freon-21^R. Report No. 707-75. Newark, Delaware: E.I. du Pont de Nemours and Co., Haskell Laboratory for Toxicology and Industrial Medicine. 2 p. [Unpublished]

A MARKON OF THE SECTION OF THE SECTI

Nuckolls, A.H. 1935. Underwriters' Laboratories Report on the comparative life, fire, and explosion hazards of dichloromonofluoromethane (FC-21). Miscellaneous Hazard No. 2630. Chicago: Underwriters' Laboratories. [15] p.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Tappan, C.H., and Waritz, R.S. 1964. Acute inhalation toxicity of Freon-21^R (fluorodichloromethane). Report No. 128-64. Newark, Delaware: E.I. du Pont de Nemours and Co., Haskell Laboratory for Toxicology and Industrial Medicine. 2 p. [Unpublished]

Trochimowicz, H.J. Lyon, J.P., Kelly, D.P., and Chiu, T. 1977a. Ninety-day inhalation toxicity studies on two fluorocarbons. Toxicol. Appl. Pharmacol. 41:200, abstr. no. 164.

Trochimowicz, H.J., Moore, B.L., and Chiu, T. 1977b. Subacute inhalation toxicity studies on eight fluorocarbons. Toxicol. Appl. Pharmacol. 41:198-199, abstr. no. 161.

THE PARTY AND ASSOCIATED BY AND ASSOCIATED TO SECOND PROPERTY AND ASSOCIATED BY ASSOCIATED BY AND ASSOCIATED BY AND ASSOCIATED BY AND ASSOCIATED BY AND ASSOCIATED BY ASSOCIATED BY AND ASSOCIATED BY ASSOCIATED BY

Weigand, W. 1971. [Studies on inhalation toxicity of fluorine derivatives of methane, ethane, and cyclobutane.] Zentralbl. Arbeitsmed. 21:149-156 (in German).

FLUOROCARBON 113

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: CF3CCi3 Molecular weight: 197.5

Chemical name: 1,1,2-trichloro-1,2,2-trifluoroethane

Synonyms: FC-113, fluorocarbon 113, Freon 113 CAS number: 76-13-1

CAS number: 76-13-1
Melting point: -35°C
Boiling point: 47.6°C

Specific gravity: 1.5635 (25°C)
Vapor pressure: 284 mm Hg (20°C)

Solubility: Insoluble in water; soluble in

alcohol, ether, and benzene

General characteristics: Colorless, nonflammable liquid

Conversion factors: 1 ppm = 8.0 mg/m³ 1 mg/m³ = 0.12 ppm

OCCURRENCE AND USE

Ī

Fluorocarbon 113 (FC-113) has been used principally as a dry-cleaning solvent, refrigerant, and blowing agent. It is also used as a cleaning solvent for sensitive electronic parts and in the maintenance of hydraulic piping systems in submarines.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

Exposure of the skin of human subjects to FC-113 resulted in localized numbness followed by transient erythema after exposure stopped (Reinhardt and Schultze, 1968). End-tidal air samples demonstrated that some solvent had been absorbed through the skin.

Human exposure at 1,500 ppm produced no effects after 2.75 h (Stopps and McLaughlin, 1967). Signs indicative of CNS involvement were seen after 30 min of exposure at 2,500 ppm. These slight decreases in psychomotor performance were reversible and disappeared by 15 min after exposure stopped.

Human volunteers were exposed at 500 or 1,000 ppm for 6 h/d, 5 d/wk, during a 2-wk period. No adverse changes were seen in performance of complex mental tasks, clinical status, or results of biochemical tests. Breath analysis did not reveal a significant buildup of FC-113 (Reinhardt et al., 1971).

EFFECTS ON ANIMALS

FC-113 has very low acute oral toxicity, with an LD50 in rats of 43 g/kg (Michaelson and Huntsman, 1964). Its lethal dose after application to the skin of rabbits is greater than 11 g/kg, the largest feasible dose. Mild irritation results from contact with the skin and eyes (Haskell Laboratory, 1963; Reinke, 1962).

Exposure to FC-113 at 50,000-60,000 ppm has proved lethal to rats after 4 h. Signs of toxicity indicative of CNS involvement were incoordination, tremors, irregular respiration, and convulsions (Bodganowicz, 1973; Dashiell, 1971; Sarver, 1971).

Rats were exposed at an average concentration of 2,520 ppm, 7 h/d, 5 d/wk, for 6 wk (Limperos, 1954). No signs of toxicity were seen throughout the experiment.

Other groups of laboratory animals were similarly exposed at up to 5,100 ppm without effect (Carter et al., 1970; Philadelphia Naval Shipyard, 1952; Steinberg et al., 1969). In a limited study involving six rats exposed at 12,000 ppm for up to 24 mc, a slight sleepiness was observed and disappeared immediately after daily exposure stopped (Desoille et al., 1968). Rats were exposed by inhalation for 6 hr a day, 5 d/wk, for 104 wk. Exposures were at 0, 2,000, 10,000, and 20,000 ppm (v/v). No significant toxic effects were observed, and no evidence of carcinogenicity was seen (C. F. Reinhardt, personal communication).

FC-113, like other chlorofluorocarbons and hydrocarbons, is capable of sensitizing the beagle heart to exogenous epinephrine in standard 5-min cardiac-sensitization screening studies. A concentration of 5,000 ppm can sensitize 25-35% of exposed dogs; 2,500 ppm is ineffective (Clark and Tinston, 1973; Reinhardt et al., 1973). However, dogs exposed while running on a treadmill (to increase their own epinephrine concentration) were not sensitized at concentrations up to 20,000 ppm (Mullin et al., 1971; Trochimowicz et al., 1974).

FC-113 is analogous to FC-11 with regard to its pharmacokinetics and metabolism. It has a short half-life in the body, is not metabolized to any significant extent, and is rapidly expelled through the lungs upon removal from exposure (C. F. Reinhardt, personal communication).

INHALATION EXPOSURE LIMITS

The ACGIH TLV-TWA and the OSHA federal standard (ACGIH, 1983; OSHA, 1983) for FC-113 are both 1,000 ppm. ACGIH recommended a TLV-STEL for 15-min excursions of 1,250 ppm (ACGIH, 1980). The TLV-TWA was recommended on the basis of a belief that it would provide a margin of safety for systemic effects and an adequate margin against cardiac sensitization.

COMMITTEE RECOMMENDATIONS

The previous EELs and CEL were established by the Committee in 1969. On the basis of 5-min cardiac-sensitization screening tests with dogs (which showed no effects at 2,500 ppm and positive findings at 5,000 ppm), an absence of effects in unexcited humans exposed at 1,500 ppm for 1.5 h, but changes in manual dexterity at 2,500 ppm for 1.5 h, the

Committee suggests a 60-min EEL of 1,500 ppm. The Committee recommends a 24-h EEL of 500 ppm on the basis of an absence of adverse health effects in unexcitead humans exposed at 500 ppm or 1,0 h/d, 5 d/wk, for 2 wk. In view of absence of adverse effects in rats exposed to 20,000 ppm (v/v) for two years, its short half-life, little metabolism, and rapid removal from the body, the Committee recommends a 90-day CEL of 100 ppm.

The present Committee's recommended EELs and CEL for FC-113 and the limits proposed in 1969 are shown below.

	1969	1984
60-min EEL	1,500 ppm	1,500 ppm
24-h EEL	200 ppm	500 ppm
90-d CEL	100 ppm	100 ppm

REFERENCES

American Conference of Governmental Industrial Hygienists. 1983. 1,1,2-Trichloro-1,2,2-trifluoroethane. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 411-412.

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Bodganowicz, T.K. 1973. Acute inhalation toxicity. Report No. 179-73. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [1] p. [Unpublished]

Carter, V.L., Chikos, P.M., MacEwen, J.D., and Back, K.C. 1970. Effects of inhalation of Freon 113 on laboratory animals. Pp. 309-314 in Proceedings of the 1st Annual Conference on Environmental Toxicology. Report No. AMRL-TR-70-102, Paper No. 20. Wright-Patterson Air Force Base, Ohio. [Also issued as NTIS AD-727 524]

Clark, D.G., and Tinston, D.J. 1973. Correlation of the cardiac sensitizing potential of halogenated hydrocarbons with their physicochemical properties. Br. J. Pharmacol. 49:355-357.

Dashiell, O.L. 1971. Acute inhalation toxicity. Report No. 46-71. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [2] p. [Unpublished]

Desoille, H., Truffert, L., Bourguignon, A., Delavierre, P., Philbert, M., and Girard-Wallon, C. 1968. Etude experimentale de la toxicite du trichlorotrifluoroethane (freon 113). Arch. Mal. Prof. Med. Trav. Secur. Soc. 29:381-388.

Haskell Laboratory for Toxicology and Industrial Medicine. 1963. Acute skin absorption of Freon 113, trifluorotrichloroethane. Newark, Delaware: E.I. du Pont de Nemours & Co. [Unpublished]

Limperos, G. 1954. Inhalation toxicity of "Freon 113" (1,1,2-trichlorotrifluoroethane). Report No. 3-54. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [9] p. [Unpublished]

Michaelson, J.B., and Huntsman, D.J. 1964. Oral toxicity study of 1,2,2-trichloro-1,1,2-trifluoroethane. J. Med. Pharm. Chem. 7:378-379.

Mullin, L.S., Azar, A., Reinhardt, C.F., Smith, P.E., Jr., and Fabryka, E.F. 1971. Halogenated hydrocarbon induced cardiac arrhythmias associated with release of endogenous epinephrine. Report No. 279-71. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [21] p. [Unpublished]

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

関う とうこので書からなった間の、Aののの

Philadelpnia Naval Shipyard. 1952. Non-inflammable solvent cleaner for insulated electrical motor parts; toxicity tests of. (ITL Test No. 2335; Progress Report NS 041 001). Philadelphia, Pennsylvania: Philadelphia Naval Shipyard, Industrial Test Laboratory. 8 p.

Reinhardt, C.F., McLaughlin, M., Maxfield, M.E., Mullin, L.S., and Smith, P.E., Jr. 1971. Human exposures to fluorocarbon 113 (1,1,2-trichloco-1,2,2-trifluoroethane). Am. Ind. Hyg. Assoc. J. 32:143-152.

Reinhardt, C.F., Mullin, L.S. and Maxfield, M.E. 1973. Epinephrine-induced cardiac arrhythmia potential of some common industrial solvents. J. Occup. Med. 15:953-955.

Reinhardt, C.F., and Schultze, L.R. 1968. Human skin absorption studies with trichlorotrifluoroethane, F-113. Report No. 84-68. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [7] p. [Unpublished]

Reinke, R.E. 1962. "Freon 113" (1,1,2-trichlorotrifluoroethane). [Eye irritation experiment]. Report No. 39-62. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [1] p. [Unpublished]

Sarver, J.W. 1971. Four-hour inhalation toxicity. Report No. 104-71. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [2] p. [Unpublished]

Steinberg, M., Boldt, R.E., Renne, R.A., and Weeks, M.H. 1969. Inhalation toxicity of 1,1,2-trichloro-1,2,2-trifluoroethane (TCTFE). Study number 33-18-68/69. Edgewood Arsenal, Maryland: U.S. Army Environmental Hygiene Agency. 20 p.

Stopps, G.J., and McLaughlin, M. 1967. Psychophysiological testing of human subjects exposed to solvent vapors. Am. Ind. Hyg. Assoc. J. 28:43-50.

Trochimowicz, H.J., Azar, A., Terrill, J.B., and Mullin, L.S. 1974. Blood levels of fluorocarbon related to cardiac sensitization: Part II. Am. Ind. Hyg. Assoc. J. 35:632-639.

FLUOROCARBON 114

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: CF4CCl2 Molecular weight: 170.93

Chemical name: 1,2-Dichloro-

1,1,2,2-tetrafluoroethane

Synonyms: FC-114, cryofluorane,

Freon 114

76-14-2 CAS number: Melting point: -94°C

Boiling point: 3.8°C

Specific gravity of liquid: 1.5312 (0°C)

Vapor pressure: 1444 mm Hg (20°C)

Solubility: Soluble in alcohol, ether,

and water (0.01%)

General characteristics: Colorless, noncorrosive,

nonflammable gas

Conversion factors: 1 ppm = 7 mg/m³ 1 mg/m³ = 0.14 ppm

OCCURRENCE AND USE

1,2-Dichloro-1,1,2,2-tetrafluoroethane (FC-114) has been used primarily as an aerosol propellant and refrigerant. Air conditioning is a suspected source of FC-114 contamination in submarines.

SUMMARY OF TOXICITY DATA

EFFECTS ON HUMANS

Human exposures to FC-114 at up to 20,000 ppm have shown it to be poorly absorbed and rapidly excreted. Warning signs of overexposure are dizziness, headache, and a tingling sensation (Dollery et al., 1970; Morgan et al., 1972a,b; Paulet, 1970; Paulet and Desbrousses, 1969; Paulet et al., 1969).

EFFECTS ON ANIMALS

FC-114 has not shown appreciable oral toxicity in laboratory animals in either acute or subchronic studies (Griffith and Sherman, 1969; Haskell Laboratory, 1955a; Quevauviller, 1965; Sherman, 1972). Administration of a foam propelled by FC-114 or of the gas itself to the skin or eyes of guinea pigs and rabbits has resulted in only mild transient irritation (Haskell Laboratory, 1955b; Hood, 1967; Quevauviller et al., 1964).

Underwriters' Laboratories, Inc., (1933) has placed FC-114 in Category 6 of its classification of life hazards. This category contains the least toxic gases and vapors; i.e., they do not appear to produce injury or death in test animals as a result of exposures at 200,000 ppm for about 2 h.

The lethal concentration of FC-114 for rats in a 2-h exposure is greater than 600,000 ppm, and for guinea pigs, greater than 500,000 ppm (Scholz, 1962). The LC50 in mice exposed for 30 min is 700,000 ppm (Paulet and Desbrousses, 1969). A dog survived an 8-h exposure at 200,000 ppm, but a 16-h exposure was lethal to another dog (Yant et al., 1932). Exposure of dogs at 150,000 ppm for 24 h was not lethal and resulted only in a loss of appetite (Yant et al., 1932). Exposure at 200,000 ppm caused pupillary dilation, convulsions, opisthotonus, and unconsciousness in dogs. However, exposure at 400,000-600,000 ppm was necessary to cause mild CNS effects in rats and guinea pigs. Recovery was rapid and complete in all species.

FC-114, like other chlorofluorocarbons and hydrocarbons, is capable of sensitizing the beagle heart to exogenous epinephrine in standard 5-min cardiac-sensitization screening studies. The concentration needed to elicit marked responses in 50% of a group of beagles is 45,000 ppm (Reinhardt et al., 1971). However, dogs exposed to FC-114 while running on a treadmill to increase their own epinephrine concentration were not sensitized until exposures reached 50,000-100,000 ppm. In another endogenous-epinephrine study, a concentration of 800,000 ppm (80% FC-113:20% oxygen) was shown to sensitize the beagle heart after its epinephrine concentration was increased by fright (Mullin et al., 1972).

Rats and mice were exposed at 200,000 ppm for 2.5 h/d, 5 d/wk, for 2 wk (Paulet and Desbrousses, 1969). Slight blood and body-weight changes were seen, as well as slight evidence of lung irritation. Exposure at 100,000 ppm had no effect, even after 2 mo of exposure (Paulet and Desbrousses, 1969). Dogs and guinea pigs were exposed at 141,600 ppm for 8 h/d for 21 d (Yant, 1933). The severity of the clinical signs attributable to CNS effects decreased during the 21 d of exposure. In the dogs, slight hematologic changes were observed, but the measures in question returned to normal 15-17 d after exposure; no gross pathologic changes were seen.

No effects were seen in dogs, cats, guinea pigs, or rats exposed at 100,000 ppm for 3.5 h per exposure for 20 exposures (Scholz, 1962). No effects were seen in rats and rabbits exposed at 10,000 ppm for 2 h/d, 5 d/wk, during 8-9 mo (Desoille et al., 1973).

INHALATION EXPOSURE LIMITS

The ACGIH TLV-TWA (ACGIH, 1980, 1983) and the OSHA Federal Standard (OSHA, 1983) for FC-114 are both 1,000 ppm. ACGIH recommended a TLV-STEL for 15-min excursions of 1,250 ppm (ACGIH, 1983). The TLV-TWA was "recommended as exposure level which should provide a margin of safety in preventing systemic toxicity and an adequate margin in preventing cardiac sensitization" (ACGIH, 1980).

COMMITTEE RECOMMENDATIONS

The Committee on Toxicology previously established EELs and CEL for FC-114 in 1966.

On the basis of positive 5-min cardiac-sensitization studies in dogs at 45,000 ppm and the absence of effects in dogs, cats, guinea pigs, or rats exposed to FC-114 at 10,000 ppm 3.5 h/day for 20 d, the Committee recommends a 60-min EEL of 10,000 ppm and a 24-h EEL of 1,000 ppm. In the absence of long-term exposure data, it appears appropriate to recommend a CEL of 100 ppm for 90 d.

The present Committee's recommended EELs and CEL for FC-114 and the limits proposed in 1966 are shown below.

	1966	1984	
60-min EEL	30,000 ppm	10,000 ppm	
24-h EEL	20,000 ppm	1,000 ppm	
90-d CEL	1,000 ppm	100 ppm	

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Dichlorotetrafluoroethane. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 135.

American Conference of Governmental Industrial Hygienists. 1983-1984. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Workroom Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Desoille, H., Truffert, L., Girard-Wallon, Cl., Ripault, J., and Philbert, M. 1973. Recherche experimentale de la toxicité chronique éventuelle du dichlorotétrafluoroéthane. Arch. Mal. Prof. Med. Trav. Secur. Soc. 34:117-125.

Dollery, C.T., Draffan, G.H., Davies, D.S., Williams, F.M., and Conolly, M.E. 1970. Blood concentrations in man of fluorinated hydrocarbons after inhalation of pressurized aerosols. Lancet 2:1164-1166.

Griffith, F.D., and Sherman, H. 1969. The effect of Freon^R-114 (ethane-1,2-dichlorotetrafluoro) on growth rate, urinary fluoride, and bone fluoride in rats. Report No. 381-69. Nevark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine, [9] p. [Unpublished]

Haskell Laboratory for Toxicology and Industrial Medicine. 1955a. Acute oral toxicity of Freon^R-114, dichlorotetrafluoroethane. Newark, Delaware: E.I. du Pont de Nemours & Co. [Unpublished]

Haskell Laboratory for Toxicology and Industrial Medicine. 1955b. Eye irritation of Freon^R-114 in rabbits. Newark, Delaware: E.I. du Pont de Nemours & Co. [Unpublished]

Hood, D.B. 1967. Skin tests with Rexall facial cleanser containing "Freon-114". Report No. 110-67. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [7] p. [Unpublished]

Morgan, A., Black, A., and Belcher, D.R. 1972a. Studies on the absorption of halogenated hydrocarbons and their excretion in breath using ³⁸Cl tracer techniques. Ann. Occup. Hyg. 15:273-282.

Morgan, A., Black, A., Walsh, M., and Belcher, D.R. 1972b. The absorption and retention of inhaled fluorinated hydrocarbon vapours. Int. J. Appl. Radiat. Isot. 23:285-291.

at a la contrata como como encolorida de alcantera de alcantera de alcantera de alcantera contrata de la compa

Mullin, L.S., Azar, A., Reinhardt, C.F., Smith, P.E., Jr., and Fabryka, E.F. 1972. Halogenated-hydrocarbon induced cardiac arrhythmias associated with release of endogenous epinephrine. Am. Ind. Hyg. Assoc. J. 33:389-396.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Paulet, G. 1970. Action of fluorochlorohydrocarbons used in aerosols. Problems of their retention by the organism after inhalation. Trib. CEBEDEAU 23:487-497. [Chem. Abs. 74:97243v, 1971]

Paulet, G., and Desbrousses, S. 1969. Le dichlorotétrafluoroéthane. Toxicité aiguë et chronique à moyen terme. Arch. Mal. Prof. Med. Trav. Secur. Soc. 30:477-492.

Paulet, G., Chevrier, R., Paulet, J., Duchêne, M., and Chappet, J. 1969. De la retention des fréons par les poumons et les voies aériennes. Étude faite chez l'homme et l'animal. Arch. Mal. Prof. Med. Trav. Secur. Soc. 30:101-120.

Quevauviller, A. 1965. [Hygiene and safety of propellants in medicated aerosols.] Prod. Probl. Pharm. 20:14-17.

Quevauviller, A., Schrenzel, M., and Vu Ngoc Huyen. 1964. Tolérance locale (peau, muqueuses, plaies, brulures chez l'animal, aux hydrocarbures chlorofluores). Therapie 19:247-263. [Index Medicus 5:1067, 1964].

Reinhardt, C.F., Azar, A., Maxfield, M.E., Smith, P.E., Jr., and Mullin, L.S. 1971. Cardiac arrhythmias and aerosol "sniffing." Arch. Environ. Health 22:265-279.

Scholz, J. 1962. New toxicologic investigations of freons used as propellants for aerosols and sprays. Translation of: Fortschr. Biol. Aerosol-Forsch. Jahren 1957-1961, Ber. Aerosol-Kongr. p. 4:420-429.

Sherman, H. 1972. Ninety-day feeding study in rats and dogs with dichloratetrafluoroethane [Freon^R 114]. Report No. 5-72. Newark, Delaware: E.I. du Pont de Nemours & Co., Haskell Laboratory for Toxicology and Industrial Medicine. [13] p. [Unpublished]

Underwriters' Laboratories. 1933. Report on the Comparative Life, Fire and Explosion Hazard of Common Refrigerants Including . . . Dichlorotetrafluoroethane Miscellaneous Hazard no. 2375, Chicago, Nov. 13.

Yant, W.P. 1933. Toxicity of organic fluorides. Am. J. Public Health 23:930-934.

Yant, W.P., Schrenk, H.H., and Patty, F.A. 1932. Toxicity of dichlorotetrafluorosthane. U.S. Bureau of Mines Report of Investigations, R.I. 3185. 10 p.

AND PROPERTY OF A PROPERTY OF A PROPERTY OF A PROPERTY OF A SECOND OF A PROPERTY OF A

ISOPROPYL ALCOHOL

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: Cakgo Molecular weight: 60.09

Chemical names: 2-Propanol, isopropanol,

propan-2-ol

Synonyms: Secondary propyl alcohol,

alcosolve 2

CAS number: 67-63-0 Freezing point: -89.5°C Boiling point: 82.4°C Specific gravity: 0.786

Vapor pressure: 33 mm Hg (20°C)

Flash point: 53°F

Auto ignition temperature: 455.6°C

Solubility: Miscible with water, alcohol,

ether, and chloroform

General characteristics: Colorless liquid with slight

odor resembling that of

rubbing alcohol

Conversion factors: 1 ppm = 2.45 mg/m^3

 $1 \text{ mg/m}^3 = 0.41 \text{ ppm}$

OCCURRENCE AND USE

Isopropyl alcohol is manufactured in the United States by an indirect hydration technique in which a fraction containing 40-60% propylene that is isolated from refinery exhaust gases reacts with sulfuric acid (Lowenheim and Moran, 1975). In an older (strong-acid) process, 88-93% sulfuric acid reacted with propylene gas at 25-60°C for a long time. In a newer (weak-acid) process, which has replaced the strongacid process, propylene gas is absorbed in 60% sulfuric acid at 85°C for a short reaction time (NIOSH, 1976). Estimated annual production capacity for 1981 was 2.8 million pounds (SRI, International, 1982).

Isopropyl alcohol was used primarily in the production of acetone, by dehydrogenating catalytically at 400°C to give acetone and hydrogen (major process) or oxidizing at high pressure to give acetone and hydrogen peroxide. However, the use of isopropyl alcohol in acetone manufacture has been decreasing in recent years. Its second main use is as a solvent: to extract or purify numerous natural products, such as oils, gums, shellacs, waxes, kelp, and pectin; in the manufacture of fish-protein concentrate; as a solvent for synthetic resins, e.g., such coatings as phenolic varnishes and nitrocollulose lacquers; and as a solvent in drug and cosmetic formulations (12 is the major component of rubbing compounds used as solvents and rubefacients) (Lowenheim and Moran, 10/5; Wickson, 1968; National Pormulary Board, 1975). Its use in cosmetics has generally been limited to highly scented or

relatively inexpensive products. The third principal use is in the manufacture of other chemicals, such as isopropyl acetate, isopropylamine, diisopropylamine, herbididal ester, isopropyl xanthate, isopropyl myristate, isopropyl palmitate, isopropyl oleate, aluminum isopropoxide, and isopropyl ether (Wickson, 1968).

Isopropyl alcohol toxicity is of interest to the Navy, because of its presence as an atmospheric contaminant in nuclear submarines.

SUMMARY OF TOXICITY INFORMATION

Table 6 summarizes some data on toxic doses of isopropyl alcohol in animals and man. The International Agency for Research on Cancer has also reviewed toxicity data (IARC, 1977).

EFFECTS ON HUMANS

The documented toxicity of impropyl alcohol in man is confined for the most part to accidental ingestion (not inhalation), with a few cases reported in association with rectal and topical application.

Several deaths have reportedly resulted from ingestion of about 1 pint of 70% isopropyl alcohol (Adelson, 1962). Other persons have survived after ingesting similar amounts (Chapin, 1949; Freireich et al., 1967; Juncos and Taguchi, 1968; King et al., 1970). The lethal dose of isopropyl alcohol is estimated as 160-240 ml (Ashkar and Miller, 1971) and 250 ml (McBay, 1973).

In 1978, 372 Melanesian men consumed a solution of 82% methyl alcohol and 18% isopropyl alcohol in the mistaken belief that the solution was methylated spirits; 18 of them died. A disparity was noted in the amount of solution consumed and the sequelae; for example, 100 ml produced blindness and death in one case, but 500 ml seemed to cause no disability in two other men who claimed to have drunk this high quantity (Scrimgeour, 1980). The rates of ingestion were not specified.

Ballard et al. (1975) reported that 15 of 41 persons working in a drug company became ill and had nausea, vomiting, weakness, and abdominal pain. Their illness was attributed to their exposure to carbon tetrachloride and isopropyl alcohol, inasmuch as 13 of the 15 had been within 25 ft of these chemicals when they were spilled.

In two factories manufacturing isopropyl alcohol by the strong-acid process (involving the formation of isopropyl oils as byproducts), an excess risk of cancers of the paranasal sinuses was found (Eckhardt, 1974; Hueper, 1966; Weil et al., 1952). An excess risk of laryngeal cancer may also have been present. However, disopropyl sulfate, an intermediate substance in the preparation of isopropyl alcohol suspected of being an animal carcinogen, is formed in the strong-acid process.

Zakhari et al. (1977) quoted several studies (Garrison, 1953; Vermeulen, 1966; McFadden and Haddow, 1969; Moss, 1970; Wise, 1969) of coma produced in hospital patients by topical application of isopropyl alcohol during sponge baths intended to reduce fever. Blood isopropyl alcohol concentrations ranged from 10 to 220 mg/100 ml; recovery in all cases was complete in 24-36 h.

Ten volunteers exposed for 3-5 min to isopropyl alcohol vapor at concentrations of 200, 400, and 800 ppm reported mild to moderate irritation of the eyes, nose, and throat at the two higher concentrations (Nelson et al., 1943).

Daily oral intake of low doses of isopropyl alcohol (2.6 or 6.4 mg/kg of body weight) by groups of eight men for 6 wk had no effect on blood cells, serum, or urine and produced no subjective symptoms (Wills et al., 1969).

Fuller and Hunter (1927) reported that dizziness occurred within a short time of oral exposure of seven human subjects to 20-30 cm³ of 50% solution of isopropyl alcohol. They also experienced moderate to severe headache lasting one to three h. The odor threshold for isopropyl alcohol ranges from 40 ppm (May, 1966) to 200 ppm (Scherberger et al., 1958).

Isogropyl alcohol is not a cutaneous irritant (Nixon et al., 1975), although several cases of allergic contact dermatitis have been reported (Fregert et al., 1971; McInnes, 1973; Richardson et al., 1969; Wasilewski, 1968).

EFFECTS ON ANIMALS

The oral LD50 of isopropyl alcohol in rats, rabbits, and dogs is about 5 g/kg (Lehman and Chase, 1944). The dermal LD50 in rabbits is about 13 g/kg, and inhalation at 16,000 ppu for 8 h was lethal to four of six rats (Smyth and Carpenter, 1948). The intravenous lethal dose in cats is 2.5 ml/kg (Macht, 1922). Isopropyl alcohol vapor at maximal saturation in air (i.e., 5.8% at 25°C) is not lethal to mice exposed for less than 1 h. The LC50 administered for 120 min to mice is 10.39 ± 3.68 mg/L (49,120 ppm). The oral and intraperitoneal LD50s are approximately equal in mice and rats. LC50s measured in rats exposed to isopropyl alcohol for 8 h were 19,000 ppm for females and 22,500 ppm for males.

The signs of intoxication after application of isopropyl alcohol are similar to those with ethyl alcohol, although it is 1.5-2 times more coric than ethyl alcohol (International Agency for Research on Cancer, 1977). Death is usually preceded by dizziness, narcosis, deep coma, and shock (Lehman et al., 1945; Morris and Lightbody, 1938). An orally administered dose of 2 g/kg produced narcotic effects in rabbits for about 8 h (Morris and Lightbody, 1938). Augmented hepatotoxicity of various chlorinated hydrocarbons was noted in mice administered isopropyl alcohol at 2.5 ml/kg 18 h before hydrocarbon exposure (Traiger and Plaa, 1974). Lehman and Chase (1944) demonstrated the doses of isopropyl alcohol that produced anesthesia and death in rabbits and dogs:

	Anesthetic Dose	Lethal Dose
Rabbits	3.23 m1/kg	8.23 m1/kg
Dogs	3.35 ml/kg	5.12 m1/kg

.

The respiratory system may be paralyzed by isopropyl alcohol; this is usually the cause of death after isopropyl alcohol ingestion (Zakhari et al., 1977). A concentration of isopropyl alcohol in air of 97.5 mg/L caused respiratory minute-volume depression, bronchoconstriction, hypotension, and bradycardia in rats. Single exposure to atmospheric isopropyl alcohol results in an increase in pulmonary resistance and a decrease in pulmonary compliance. These effects are more pronounced if more than six daily exposures are administered (Zakhari et al., 1977).

In anesthetized dogs, isopropyl alcohol inhalation caused various effects: depression of myocardial contractility at 1.0% (2.45 mg/L), reduction in cardiac output at 2.5% (6.12 mg/L), and systemic hypotension at 7.5% (18.37 mg/L) (Zakhari et al., 1977). Histopathologic examination of rats exposed at 21,000 ppm for 8 h showed typical lesions of chemical pneumonitis and pulmonary edema accompanied by foamy vacuolization of liver cells and severe focal cytoplasmic degradation (Laham et al., 1980).

Baikov et al. (1974) investigated the effects of chronic inhalation of isopropyl alcohol by rats. Groups of 15 animals were exposed to isopropyl alcohol continuously for 24 h/d for 86 d at concentrations of 20, 2.5, and 0.6 mg/m³ (approximately 8.14, 1.02, and 0.24 ppi). The animals inhaling isopropyl alcohol at 20 mg/m³ (8.14 ppm) showed changes in reflex behavior, increases in the retention of BSP, the total leukocyte count, and the number of abnormal fluorescent leukocytes. They also showed a decrease in the blood nucleic acid content, the blood oxidase and catalase activities, and the amount of coproporphyrin in blood. Animals inhaling isopropyl alcohol at 2.5 mg/m³ (1.02 ppm) demonstrated some of the same effects, but none were statistically significant. In animals inhaling isopropyl alcohol at 20 mg/m³ (8.14 ppm), postmortem findings included hyperplasia of the spleen with the development of hemorrhages of the sinuses and erosion of follicular cells, some evidence of liver parenchymal cell dystrophy, hyperplastic ependymal cells, and degenerative changes in the cerebral motor cortex. None of these effects were observed in animals inhaling isopropyl alcohol at 0.6 mg/m^3 (0.24 ppm). On the basis of this continuous exposure study, the authors suggested that 0.6 mg/m^3 (0.24 ppm) be adopted as the maximal daily average concentration. Some of the physiological responses reported in this study, such as the increase in abnormal fluorescent leukocytes, are obscure and are therefore difficult to interpret. Furthermore, they suffer from a lack of experimental details.

Isopropyl alcohol (10%) in the diet of young rats for 30 d had no effect on growth, liver weight, or lipid content (Miyazaki, 1955).

Dogs given daily doses of 1.3 g/kg in the drinking water had the appearance of drunkenness 3-5 h after intake, but no consistent pathologic changes over a 6-mo period (Lehman et al., 1945).

No evidence of carcinogenicity was found when several strains of mice were exposed to isopropyl alcohol in air at 7,700 mg/m³ 3-7 h/d, 5 d/wk, for 5-8 mo. However, the animals were not observed over a normal lifetime and were killed at 8-12 mo of age (Weil et zi., 1952). The authors reported no increases in the incidence of lung tumors in mice given subcutaneous injections of 0.025 ml of isopropyl

alcohol once a week for 20-40 wk. However, in mice sacrificed at 40 wk, effects of lifetime exposure were not assessed. No skin tumors were detected (NIOSH, 1976) in a group of 30 Rockland mice that received skin applications of isopropyl alcohol twice a week for a year. Because of methodologic limitations in the preceding studies, it is difficult to draw a conclusion regarding the carcinogenicity of isopropyl alcohol (IARC, 1977).

Lehman et al. (1945) reported that growth, reproductive function, and embryonic and postnatal development of rats were not affected, except for some retardation of growth early in the life of first-generation offspring when parents and two successive generations of rats were given isopropyl alcohol continuously in the drinking water at 1.5, 1.4, and 1.3 g/kg per day, respectively.

というのでは、一種のなり、このないのでは、このでは

•

ころうり うちゅうしゅうしょうしょうし

ĺ

との とととう あり

Solvent controls of isopropyl alcohol in an assay of the mutagenicity of Fusarium moniliforms were negative when tested in the S. typhimurium reverse-mutation bioassay with strains TA 98 and TA 100 (Bjeldanes and Thomson, 1979).

PHARMACOKINETICS

After oral intake of 0.1-20 g of isopropyl alcohol, none was excreted in the urine of volunteers laring the next 48 h, and no formic acid was detected (Kemal, 1927).

Isopropyl alcohol is absorbed from all segments of the gastro-investinal tract, most rapidly in the small intestine and least rapidly in the stomach. Wax et al. (1949) reported that absorption from isolated canine loops of intestine and stomach was 99% complete at the end of 2 h, 82% being absorbed during the first 30 min. Intravenous injections of ethyl alcohol (0.8 cc/kg) significantly reduced the absorption of is propyl alcohol from the digestive tract.

Before 1940, the toxicity of isopropyl alcohol was compared with that of methyl alcohol, in which the long and cumulative action was attributed to low rates of metabolism and excretion. But most data now point to a fairly rapid disposal of isopropyl alcohol, which is eliminated from the bloodstream of dogs within 24 h after administration.

The elimination of isopropyl alcohol in rats is decreased by simultaneous ingestion of ethyl alcohol or 1-propanol, but not methyl alcohol or tertiary butyl alcohol. These results suggest that isopropyl alcohol is oxidized by alcohol dehydrogenase (Abshagen and Rietbrock, 1970).

Determinacion of blood isopropyl alcohol and its metabolite, acetone, was carried out during and after a single 4-h exposure (concentration, 500-8,000 ppm) in Sprague-Dawley rats. The amounts of acetone and isopropyl alcohol were directly related to the air concentrations of alcohol inhaled. Increase in exposure time to 8 h considerably increased the amount of blood acetone that could be determined even 20 h after exposure. These findings indicate a slow conversion of this alcohol to acetone, which can be used as a biochemical indicator of exposure, but it is a nonspecific indication and may be produced by other compounds (Laham et al., 1980).

Exposure of male Wistar rats to isopropyl alcohol vapor at 12.3 mol/L (300 ppm) for 6 h/d, 5 d/wk, for 5-21 wk with simultaneous ethyl

alcohol administration in drinking water (5% v/v) caused a significant increase in isopropyl alcohol removal as assessed by blood isopropyl alcohol and acetone determinations (Savolainen et al., 1979). This study confirmed the production of acetone from isopropyl alcohol. The novel aspect of this study is the more rapid metabolism of acetone induced by ethyl alcohol. This effect might be attributable to synergistic effects of ethyl alcohol and isopropyl alcohol on aldehyde and ketone dehydrogenation. Neurochemical studies revealed decreased superoxide dismutase and azoreductase activities in cerebellar homogenate at the end of the exposure, whereas increased protein degradation was found in glial cells isolated from rats fed ethyl alcohol. Analyses of spinal cord axon lipid composition showed increases in cholesterol content in relation to lipid phosphorus in animals exposed to isopropyl alcohol or to the combination of isopropyl and ethyl alcohol. Spontaneous behavioral tests indicated minor effects on reactivity from the tenth week on with isopropyl alcohol exposure. Coexposure to isopropyl alcohol vapor and ethyl alcohol abolished the increased excitability. Savolainen et al. concluded that isopropyl alcohol vapor causes significant metabolic and functional chages in rats at relatively low doses -- 300 ppm.

Another synergistic effect has been an increase in the toxicity of carbon tetrachloride caused by pretreatment with isopropyl alcohol (Plaa and Traiger, 1973; Traiger and Plaa, 1973; Traiger and Plaa, 1974). The concentration of isopropyl alcohol in rat saliva 15 and 60 min after exposure was proportional to the exposure concentration and the duration of exposure and was closely correlated with the blood concentration (Tomita, 1980).

Liver alcohol dehydrogenase is the principal enzyme involved in the oxidation of Isopropyl alcohol. The acetone produced by action of the enzyme on isopropyl alcohol is eliminated from the human body in expired air and urine (Zakhari et al., 1977).

Rat liver microsomes are capable of oxidizing branched-chain alcohols, and hydroxyl radicals generated from microsomal electron transfer may have a role in isopropyl alcohol oxidation (Cederbaum et al., 1981).

INHALATION EXPOSURE LIMITS

The OSHA health standards for exposure to air contaminants require that an employee's exposure to isopropyl alcohol not exceed an 8-h TWA of 400 ppm in the working atmosphere in any 8-h shift of a 40-h workweek (OSHA, 1982); a ceiling of 800 ppm was determined during a sampling time of 15 min (NIOSH, 1976). An estimated 141,000 employees may be exposed occupationally to isopropyl alcohol in the United States (NIOSH, 1976).

The American Conference of Governmental Industrial Hygienists established the value of 400 ppm as the TLV for isopropyl alcohol; as described above, this is the TLV currently recommended in the United States (American Conference of Governmental Industrial Hygienists, 1983, 1983). The permissible concentration of isopropyl alcohol was established by the Japan Association of Industrial Health in 1966 at 400 ppm (Japan Association of Industrial Health, 1971). In the

U.S.S.R., the maximal permissible concentration of isopropyl alcohol in a single dose or as a daily average is $0.6~mg/m^3$ (U.S.S.R. Ministry of Public Health Individual Reports, 1971).

COMMITTEE RECOMMENDATIONS

というに重要があるのでも 日からしていると言うないないに 間間 と

1.000 St. 1.000

The National Research Council's Committee on Toxicology (1960) recommended, as a part of the submarine toxicology program, the EELs and CEL for isopropyl alcohol. The basis for these limits was primarily the ACGIH TLV of 400 ppm, which was recommended by the Committee as the 60-min EEL. The Committee stated: "From the data available it would appear that isopropyl alcohol is not an industrial health hazard. The threshold limit value of 400 ppm set by the ACGIH may cause mild irritation of the eyes, nose and throat, especially in those not regularly exposed." The 24-h EEL and 90-d CEL were apparently recommended by the Committee on the basis of its judgment concerning tolerable doses for these periods and extrapolations from the 400-ppm 60-min EEL. No new information has become available to suggest changes in previously proposed EELs for 1 and 24 h. However, data from long-term, low-dose, continuous-exposure studies show adverse effects on behavior, liver, and spleen in rats (Baikov et al., 1974). Although these data are not conclusive, prudence dictates that previously recommended long-term exposure limit (CEL) of 50 ppm be lowered to 1 ppm.

The present Committee's recommended EELs and CEL for isopropyl alcohol and the limits proposed in 1960 and 1966 are shown below:

	1960/1966	1984	
60-min EEL	400 ppm	400 ppm	
24-h EEL	200 ppm	200 ppm	
90-d CEL	50 ppma	1 ppm	

TABLE 6

Acute Toxicity of Isopropyl Alcohol^a

Species	Route	Measure	Value		
Human	Eye	Irritation	20	ppm	
Human	Oral	$\mathtt{TD}_{\mathbf{Lo}}$	15,710	mg/kg	
Human	0ral	$\mathtt{LD_{Lo}}$	8,600	mg/kg	
Human	Inhalation	TC10	400	ppm	
Human	Subcutaneous	$\mathtt{LD}_{\mathbf{Lo}}$	6	mg/kg	
Human	Unknown	$\mathtt{LD}_{\mathbf{Lo}}$	2,770	mg/kg	
Rat	Oral	LD50	5,840	mg/kg	
Rat	Inhalation	LC50	16,000	ppm, 8	h
Mouse	Oral	LC _{Lo}	192	mg/kg	
Mouse	Intraperitoneal	LD50	933	mg/kg	
Mouse	Subcutaneous	$\mathtt{LD_{Lo}}$	6,000	mg/kg	
Dog	Oral	LD50	6,150	mg/kg	
Dog	Intravenous	LD_{Lo}	5,120	mg/kg	
Cat	Intravenous	LD _{I.o}	1,963	mg/kg	
Rabbit	Eye	Irritation	16	mg	
Rabbit	Oral	$\mathtt{LD_{Lo}}$	5,000	mg/kg	
Rabbit	Skin	LD ₅₀	13	g/kg	
Rabbit	Intravenous	LDLo	8,230	mg/kg	

a Data from NIOSH Registry of Toxic Effects of Chemical Substances, 1982.

REFERENCES

Abshagen, U., and Rictbrock, N. 1970. Zum Mechanismus der 2-Propanoloxydation. Interferenzversuche mit niederen aliphatischen Alkoholen in vivo und an der isoliert perfundierten Rattenleber. Naunyn-Schmiedeberg's Arch. Exp. Pathol. Pharmakol. 265:411-424.

Adelson, L. 1962. Fatal intoxication with isopropyl alcohol (rubbing alcohol). Am. J. Clin. Pathol. 38:144-151.

American Conference of Governmental Industrial Hygienists. 1980. Isopropyl alcohol. Documentation of the Threshold Limit Values. 4th ed., with Supplemental Documentation through 1983. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 239(81).

American Conference of Governmental Industiral Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical SUbstances and Physical Agents in the Work Environment with Intended Changes for 1983-1984, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Ashkar, F.S., and Miller, R. 1971. Hospital ketosis in the alcoholic diabetic: a syndrome due to isopropyl alcohol intoxication. South. Med. J. 64:1409-1411.

Baikov, B.K., Gorlova, O.E., Gusev, M.I., Novikov, Yu V., Yudina, T.V. and Sergeev, A.N. 1974. Hygienic standardization of the daily average maximal permissible concentrations of propyl and isopropyl alcohols in the atmosphere. Gig. Sanit. No. 4: 6-13. [English summary]

Ballard, T.W., Walker, P., Ginn, E., Sanders, R.S., McMurray, D., Horst, E.V., and Hutchenson, R.H., Jr. 1975. Renal failure and hepatitis from inhaled carbon tetrachloride and isopropyl alcohol. Morbidity and Mortality (Center for Disease Control) 24:59-60.

Bjeldanes, L.F., and Thomson, S.V. 1979. Mutagenic activity of Fusarium moniliforme isolates in the Salmonella typhimurium assay. Appl. Environ. Microbiol. 37:1118-1121.

Cederbaum, A.I., Qureshi, A., and Messenger, P. 1981. Oxidation of isopropanol by rat liver microsomes: Possible role of hydroxyl radicals. Biochem. Pharmacol. 30:825-831.

Chapin, M.A. 1949. Isopropyl alcohol poisoning with acute renal insufficiency. J. Maine Med. Assoc.. 40:288-290. [Chem. Abs. 44:10137g, 1950]

Eckardt, R.E. 1974. Annals of industry - noncasualties of the work place. J. Occup. Med. 16:472-477.

Fregert, S., Groth, O., Gruvberger, B., Magnusson, B., Mobacken, H. and Rorsman, H. 1971. Hypersensitivity to secondary alcohols. Acta Derm. Venereol. 51:271-272.

Freireich, A.W., Cinque, T.J., Xanthaky, G., and Landau, D. 1967. Hemodialysis for isopropanol poisoning. N. Engl. J. Med. 277:699-700.

Fuller, H.C., and Hunter, O.B. 1927. Isopropyl alcohol--An investigation of its physiologic properties. J. Lab. Clinic Med. 12:326-341.

Garrison, R.F. 1953. Acute poisoning from use of isopropyl alcohol in tepid sponging. J. Am. Med. Assoc. 152:317-318.

Hueper, W.C. 1966. Occupational and Environmental Cancers of the Respiratory System. (Recent Results Cancer Res. 3) New York: Springer-Verlag. p. 105-107, 183.

International Agency for Research on Cancer. 1977. IARC Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Man: Some Fumigants, the Herbicides 2,4-D and 2,4,5-T, Chlorinated Dibenzodioxins and Miscellaneous Industrial Chemicals, Vol. 15. Lyon: International Agency for Research on Cancer. 354 p.

Japanese Association of Industrial Health. 1971. Recommendation for permissible concentrations, etc. Translation of Sangyo Igaku (Jpn. J. Ind. Health) 13:475-484.

Juncos, L., and Taguchi, J.T. 1968. Isopropyl alcohol intoxication: Report of a case associated with myopathy, renal failure, and hemolytic anemia. J. Am. Med. Assoc. 204:732-734.

Kemal, H. 1927. Beitrag zur Kenntnis der Schicksale des Isopropylalkohols im menschlichen Organismus. Biochem. Z. 187:461-466.

King, L.H., Jr., Bradley, K.P., and Shires, D.L., Jr. 1970. Hemodialysis for isopropyl alcohol poisoning. J. Am. Med. Assoc. 211:1855.

Laham, S., Potvin, M., Schrader, K., and Marino, I. 1980. Studies on inhalation toxicity of 2-propanol. Drug Chem. Toxicol. 3:343-60.

Lehman, A.J., and Chase, H.F. 1944. The acute and chronic toxicity of iso-ProH. J. Lab. Clin. Med. 29:561-567. [Chem. Abs. 38:59712, 1944]

Lehman, A.J., Schwerma, H., and Rickards, E. 1945. Isopropyl alcohol. Acquired tolerance in dogs, rate of disappearance from the blood stream in various species, and effects on successive generation of rats. J. Pharmacol. Exp. Ther. 85:61-69.

Lowenheim, F.A., and Moran, M.K. 1975. Faith, Keyes, and Clark's Industrial Chemicals. 4th ed. New York: John Wiley & Sons. p. 496-501.

Macht, D.I. 1922. Pharmacological examination of isopropyl alcohol. Arch. Int. Pharmacodyn. Ther. 26:285-286. [Chem. Abs. 16:2923, 1922]

May, J. 1966. Geruchsschwellen von Lösemitteln zur Bewertung von Lösemittelgerüchen in der Luft. Staub Reinhalt. Luft 26:385-389.

McBay, A.J. 1973. Toxicological findings in fatal poisonings. Clin. Chem. 19:361-365.

McFadden, S.W., and Haddow, J.E. 1969. Coma produced by topical application of isopropanol. Pediatrics 43:622-623.

McInnes, A. 1973. Skin reaction to isopropyl alcohol. Br. Med. J. 1:357.

Miyazaki, M. 1955. Nutritive value of aliphatic alcohols. I. The nutritive value of saturated alcohols with one to five carbon atoms. J. Agric. Chem. Soc. Japan 29:497-501. [Chem. Abs. 49:149421, 1955]

Morris, H.J., and Lightbody, H.D. 1938. The toxicity of isopropanol. J. Ind. Hyg. Toxicol. 20:428-434.

Moss, M.H. 1970. Alcohol-induced hypoglycemia and coma caused by alcohol sponging. Pediatrics 46:445-447.

National Formulary Board. 1975. National Formulary XIV. Washington, D.C.: American Pharmaceutical Association. p. 367-368.

National Institute for Occupational Safety and Health. 1976. Criteria for a Recommended Standard . . . Occupational Exposure to Isopropyl Alcohol. HEW Publication No. (NIOSH) 76-142. Cincinnati, Ohio: U.S. Department of Health, Education and Welfare, Public Health Service, Center for Disease Control. National Institute for Occupational Safety and Health. 119 p.

National Institute for Occupational Safety and Health. 1982.
Registry of Toxic Effects of Chemical Substances. 1980 ed. DHHS
(NIOSH) Publication No. 81-116. Rockville, Maryland: U.S. Department of Health and Human Services, Public Health Service, Centers for Disease Control, National Institute for Occupational Safety and Health. Vol. 1., p. 967.

National Research Council. Committee on Toxicology. 1960. Report to U.S. Navy on Isopropyl Alcohol. 3 p.

Nelson, K.W., Ege, J.F., Jr., Ross, M., Woodman, L.E., and Silverman, L. 1943. Sensory response to certain industrial solvent vapors. J. Ind. Hyg. Toxicol. 25:282-285.

Nixon, G.A., Tyson, C.A., and Wertz, W.C. 1975. Interspecies comparisons of skin irritancy. Toxicol. Appl. Pharmacol. 31: 481-490.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Plaa, G.L., and Traiger, G.J. 1973. Mechanism of potentiation of CC14-induced hepatotoxicity. In Loomis, Ted A., ed. Pharmacology and the Future of Man. Vol. 2. Toxicological Problems. New York: S. Karger. p. 100-113.

Richardson, D.R., Caravati, C.M., Jr., and Weary, P.E. 1969.
Allergic contact dermatitis to "alcohol" swabs. Cutis 5:1115-1118.

Savolainen, H., Pekari, K., and Helojcki, H. 1979. Neurochemical and behavioural effects of extended exposure to isopropanol vapour with simultaneous ethanol intake. Chem. Biol. Interact. 28:237-248.

Scherberger, R.F., Happ, G.P., Miller, F.A., and Fassett, D.W. 1958. A dynamic apparatus for preparing air-vapor mixtures of known concentrations. Am. Ind. Hyg. Assoc. J. 19:494-498.

Scrimgeour, E.M. 1980. Outbreak of methanol and isopropanol poisoning in New Britain, Papua New Guinea. Med. J. Aust. 2:36-38.

Smyth, H.F., Jr., and Carpenter, C.P. 1948. Further experience with the range finding test in the industrial toxicology laboratory. J. Ind. Hyg. Toxicol. 30:63-68.

SRI International. 1982. Directory of Chemical Producers, United States of America. Menlo Park, CA: SRI International. 1132 p.

Tomita, M. 1980. Investigation of volatile matters in oral cavity. An evaluation of exposure levels by concentration of organic solvents in saliva. II. Shike Gakuho 80:1705-1721. [Chem. Abs. 95: 1321d, 1981]

Traiger, G.J., and Plaa, G.L. 1973. Effect of aminotriazole on isopropanol—and acetone—induced potentiation of CC14 hepatotoxicity. Can. J. Physiol. Pharmacol. 51:291-296.

Traiger, G.J., and Flaa, G.L. 1974. Chlorinated hydrocarbon toxicity: potentiation by isopropyl alcohol and acetone. Arch. Environ. Health 28:276-278.

U.S.S.R. Ministry of Public Health Individual Reports. 1971. Maximum Permissible Concentration of Harmful Substances in the Atmospheric Air of Populated Areas. No. 876-71. Environmental Protection Agency Translation TR 175-74. [18] p.

Vermeulen, R. 1966. Isopropyl alcohol and diabetes. Pa. Med. 69(4):53-54.

Wasilewski, C., Jr. 1968. Allergic contact dermatitis from isopropyl alcohol. Arch. Dermatol. 98:502-504.

Wax, J., Ellis, F.W., and Lehman, A.J. 1949. Absorption and distribution of isopropyl alcohol. J. Pharmacol. Exp. Ther. 97:229-237.

ハススと 見い アイマイマン 野道

ļ

Weast, R.C., ed. 1976. CRC Handbook of Chemistry and Physics. 57th ed. Cleveland, Ohio: CRC Press. p. C-459.

Weil, C.S., Smyth, H.F., Jr., and Nale, T.W. 1952. Quest for a suspected industrial carcinogen. AMA Arch. Ind. Hyg. Occup. Med. 5:535-547.

Wickson, E.J. 1968. Isopropyl alcohol. In Kirk-Othmer Encyclopedia of Chemical Technology. 2nd rev. ed., Vol. 16. New York: Interscience Publishers. p. 564-578.

Wills, J.H, Jameson, E.M and Coulston, F. 1969. Effects on man of daily ingestion of small doses of isopropyl alcohol. Toxicol. Appl. Pharmacol. 15: 560-565.

Wise, J.R., Jr. 1969. Alcohol sponge baths. N. Engl. J. Med. 280: 840.

Zakhari, S., Leibowitz, M., Levy, P. and Aviado, D.M. 1977. Isopropanol and Ketones in the Environment. Cleveland, Ohio: CRC Press. 147 p.

PHOSGENE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: COC12 98.9 Molecular weight:

> Chemical name: Carbonyl chloride, carbon

> > oxychloride, chloroformyl

chloride

75-44-5 CAS number: -127.8°C Freezing point: 7.5°C Boiling point:

1.4187 (20°C) Density, liquid: 4.39 (20°C) Density gas:

Specific gravity, gas: $3.4 (2^{\circ}C) (air = 1)$

pecific gravity, liquid: 1.392 (19°C/4°C)

Solubility: Slightly soluble in water

(hydrolyzes to HC1 and CO2); soluble in carbon tetrachloride,

chloroform, acetic acid, and toluene

Easily liquified, colorless, General characteristics:

nonflammable gas; odor, sweet at low

ranning transport of the program of

concentrations and pungent at higher

Conversion factors: 1 ppm = 4 mg/m³ 1 mg/m³ = 0.25 ppm

OCCURRENCE AND USE

Phosgene was first prepared in 1812 by the photochemical reaction of carbon monoxide and chlorine; it is now commercially prepared by passing chlorine and excess carbon monoxide over activated carbon. Depending on the quantity required and the availability of the raw material, numerous variations of the basic synthetic process are used. Continuous processing and a high degree of automation are required for phosgene purification, condensation, and storage.

In its first application, phosgene was the most heavily used chemical-warfare agent during World War I (Moore and Gates, 1946; Stavrakis, 1971). Its utility as a reactive chemical intermediate is of relatively recent origin (Chadwick and Hardy, 1967; Chadwick and Cleveland, 1981; Hardy, 1982; NIOSH, 1976). It is now an important and widely used intermediate; the majority of its production is captive, e.g., in the manufacture of other chemicals within the same plant. The principal use of phosgene is in the polyurethane industry, which consumes over 85% of the world's phosgene output. It is also used in polycarbonate resins, dyes, and pharmaceutical intermediary products (Chadwick and Cleveland, 1981; Hardy, 1982; NIOSH, 1975).

In 1978, demand for phosgene was an estimated 1,630 million pounds. The growth in U.S. demand for phosgene from 1970 to 1979 was 9.2%/yr; future growth in demand is forecast at 7.0%/yr through 1984 (Anonymous, 1980). The capacity for the production of phosgene in the United States (12 producers at 15 sites) in 1983 was estimated to be < 2,101 million pounds (SRI International, 1983).

In 1973, the pattern of phosgene use was as follows: production of toluene diisocyanate (TDI), 61.7%; production of polymethylene polyphenylisocyanate (PMPPI), 23.6%; production of polycarbonate resins, 3.9%; and other uses (including production of acyl chlorides, chloroformate esters, diethylcarbonate, dimethylcarbamylchloride, isocyanates other than TDI and PMFPI, dyes, biocides and pharmaceuticals and use as a chlorinating agent), 10.7% (SRI International, not dated).

In 1977, the polycarbonate industry consumed approximately 6% of the phosgene produced and herbicide manufacture and processing used 9% (Hardy, 1982).

World production of isocyanates in 1978 has been estimated as 635,000 metric tons of TDI and 454,000 metric tons of diphenyl methane-4,4'-diisocyanate (MDI) products. The estimated 1981 capacities of U.S. manufacturers of isocyanates amounted to 318,000 and 444,000 metric tons for TDI and polymeric isocyanates, respectively (Chadwick and Cleveland, 1981).

TDI is a precursor of polyurethane resins, which are widely used to make foams, elastomers, and coatings. Polycarbonate resins based on phosgene find use in appliance and electric-tool housings, electronic parts, and break-resistant glazing. A rapidly growing use of phosgene is in the preparation of PMPPI for the production of rigid polyurethane foams (SRI International, not dated). The reaction of phosgene with primary alkyl and aryl amines, referred to as phosgenation, yields carbamoyl chlorides, which can be dehydrohalogenated readily to isocyanates:

 $RNH_2 + COCl_2 \rightarrow RNHCOCl \rightarrow RN=C=O + HCl.$

This procedure is used almost exclusively for the production of isocyanates (Chadwick and Cleveland, 1981). All commercial manufacturing processes for aromatic isocyanates currently in use appear to have the following steps:

- A solution of an amine in an aromatic solvent—such as xylene, monochlorobenzene, or o-dichlorobenzene—is mixed with a solution of phosgene in the same solvent at a temperature below 60°C.
- The resulting mixture slurry is digested in one to three stages for several hours at progressively increasing temperatures up to 200°C; digestion is accompanied by the injection of additional phosgene.
- The final solution of reaction products is fractionated to recover hydrogen chloride, unreacted phosgene, and solvent for recycling, isocyanate product, and distillation residue.

In addition to commercial production, phosgene can be produced by the thermal decomposition of chlorinated hydrocarbons (Spolyar et al., 1951) and by the photooxidation of chlorinated hydrocarbons in the atmosphere (NIOSH, 1976; Singh, 1976). For example, Singh (1976) demonstrated by field monitoring at four California sites (urban and nonurban) that chloroethylenes (primarily C2Cl4 and C2HCl3), which are emitted worldwide in extremely large quantities (1.5 million metric tons in 1975), photodecompose to form highly toxic species, such as phosgene and chloroacetylchlorides. On the basis of estimates from recent laborarory studies, present C2C14 and C2HC13 emission could result in the formation of about 0.3 million metric tons of phosgene per year (Gay et al., 1976). Because these solvents are typically emitted in urban areas and are relatively reactive (the tropospheric half-lives of C2HCl3 and C2Cl4 are 2 and 4 days, respectively), high concentrations of phosgene could be encountered during adverse meteorologic conditions in and around urban centers. The atmospheric sinks of phosgene have not been fully delineated. Simulated tropospheric irradiation in the presence of water vapor suggests negligible tropospheric loss through photolysis and gas-phase hydrolysis. The two important sinks of phosgene appear to be heterogeneous decomposition and slow liquid-phase hydrolysis (Singh, 1976).

NIOSH estimated that about 10,000 workers have potential occupational exposure to phosgene during its manufacture and use (NIOSH, 1976). Their occupations include those involved in the production of phosgene itself, chlorinated compounds, dyes, glass, herbicides, insecticides, isocyanates, organic chemicals, and resins and firefighting, welding, and brazing (Gafafer, 1966; NIOSH, 1976). Long-term exposure at low concentrations may occur in submarines. The potential source of phosgene in submarines could be leaked Freon undergoing thermal decomposition.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

Three sources of exposure to phosgene in air are readily identifiable: direct emission of phosgene during manufacture, handling, and use (ACGIH, 1980; Polednak, 1980); thermal decomposition of polyvinyl-chloride (PVC) (Brown and Birky, 1980) and chlorinated hydrocarbons (Cucinell, 1974; NIOSH, 1976), such as chloroform, carbon tetrachloride, and trichloroethylene—solvents, paint removers, and nonflammable dry-cleaning fluids containing chlorinated hydrocarbons may decompose to phosgene in the presence of fire or heat (Diller, 1978); and photooxidation of chloroethylenes in air (Gross et al., 1965). The first two sources might result in serious indoor hazard (Diller, 1978; NIOSH, 1976; Spolyar et al., 1951), but their contribution to the total ambient phosgene content is minimal (Spolyar et al., 1951), whereas segments of the general population could be exposed at higher concentrations from atmospheric pollution, e.g., from fuel emission of municipal incinerators (Carotti and Kaiser, 1972).

Human exposure to gaseous phosgene reported in the literature is generally limited to episodes of acute exposure (Diller, 1978). The acute toxicity of phosgene is both dose- and time-dependent. Phosgene at concentrations of 3-5 ppm causes irritation of the eyes and throat with coughing; exposure at 25 ppm for 30-60 min is dangerous; and brief exposure at 50 ppm may be rapidly fatal (Henderson and Haggard, 1943; Hygienic Guide Series, 1968; Patty, 1963; Sax, 1968).

The symptoms of moderate exposure to phosgene are often dryness or a burning sensation in the throat, vomiting, pain in the chest, and dyspnea (Patty, 1963). Phosgene poisoning is characterized by a symptom-free latent period of 2-24 h followed by chest pain, shortness of breath, and increasing difficulty in breathing. Severe respiratory distress may be delayed for up to 72 h; the latent interval depends on the concentration and duration of exposure (Hygienic Guide Series, 1968). The distress is caused by pulmonary edema and is characterized by cough, production of foamy sputum, progressive dyspnea, and severe cyanosis (Patty, 1963). Pulmonary edema may progress to pneumonia, and cardiac failure may intervene. In nonfatal cases, no permanent residual damage is believed to occur (Patty, 1963; Sax, 1968).

The mortality among two groups of workers exposed to phosgene gas at the same uranium-processing plant in 1943-1945 has been described by Polednak (1980). One group consisted of 699 white men with daily exposure at low concentrations or at concentrations above 1 ppm; the second group consisted of 106 men with definite acute exposures and symptoms at exposures equal to or greater than 50 ppm/min. A control group included 9,352 white men who worked at the same plant. Of the group of 105 men, 25 had x-ray evidence of acute pneumonitis, and one death from pulmonary edema due to phosgene occurred less than 24 h after exposure. A total of 30 deaths had occurred in this group as of 1974 (SMR = 113); there were no deaths from lung cancer, but three deaths (vs. 1.37 expected) were due to respiratory disease. Mortality ratios (SMRs) for most causes of death among the 699 chemical workers were similar to those among 9,352 controls employed at the same plant but not exposed to phosgene (or uranium dust).

Table 7 lists a number of cases of human phosgene inhalation exposure (NIOSH, 1976). Splashes of liquid phosgene into the eye may produce severe irritation, and skin contact may cause severe burns (Hygienic Guide Series, 1968).

Fatal exposure to phosgene has resulted in extensive degenerative changes in the epithelium of the trachea, bronchi, and bronchioli and hemorrhagic edematous focal pneumonia (Gerritsen and Buschmann, 1960). Most fatalities due to acute phosgene exposure occur during the first 24-48 h (Chadwick and Hardy, 1967). Most patients who died within the first 72 h died of pulmonary edema or cardiac problems; those who died later had such complications as pulmonary infection, thrombosis, and embolism. The clinical course has also been described by NIOSH (1976), Boyd and Perry (1960), Delephine (1923), Gerritsen and Buschmann (1960), Glass et al. (1971), Long and Hatch (1961), Spolyar et al. (1951), Stavrakis (1971), and Underhill (1920).

No effects other than odor detection were reported (NIOSH, 1976) in 56 military personnel (without upper respiratory problems) who were exposed to phosgene at increasing concentrations until they could all smell it. Of "technically trained" personnel, 50% detected phosgene

· 一般的人的人的人的人

at 1.5 ppm, 39% detected it at 1.2 ppm, and none below 0.4 ppm (Wills et al., 1938).

EFFECTS ON ANIMALS

Acute overexposure to phosgene for short periods has generally produced the same symptoms in all animal species tested (NIOSH, 1976). The lungs appear to be the principal target organ for phosgene, and the characteristic pathologic feature is the development of pulmonary edema of unknown pathogenesis. Survivors of an acute episode exhibit various degrees of bronchopneumonia, benign pneumonia, bronchial plugging, lung collapse, pulmonary consolidation, pneumonia, and emphysema; animals that die after exposure show severe pulmonary edema. For example, Cameron et al. (1942a) exposed mice, rats, guinea pigs, rabbits, cats, monkeys, and goats to phosgene at 0.86 ppm for 5 h. Within 24 h, 10% of the rats and 60% of the sice died. Microscopic examination of the lungs showed severe effects in 39% of the animals, mild effects in 31%, and slight effects in 30%; pulmonary edema was the most common finding. Cameron et al. (1942b) also exposed mice, rats, guinea pigs, rabbits, cats, and goats to phosgene at 0.2 ppm, 5 h/d for 5 d. No deaths occurred from the acute exposure, and few animals showed any evidence of distress. Necropsy disclosed pulmonary lesions in 67% of the animals; an estimated 4-11% displayed moderate to severe lesions. Pulmonary edema was noted in 41% of the animals, but was usually slight. It was concluded that repeated exposure to phosgene at low concentrations induced lung damage, although rarely to a severe degree. However, Cucinell (1974), in reviewing the latter study, stressed the extensive lung lesions present in 4% of the animals. Exposure of the same species at 1 ppm, 5 h/d for 5 d caused pulmonary lesions that were considered "likely to give rise in man to serious clinical symptoms" (Cameron and Foss, 1941). Exposure of cats at the same concentration caused hemoconcentration and leukocytosis (Cucinell, 1974).

人名阿拉勒 经公司的收益的收益

Will and the second

(2) おようである。これではないない。またのできる。 数のののできる 数を含めるがある。

Gross et al. (1965) demonstrated chronic pneumonitis in rats as early as 4 h after exposure to phosgene. The sensitivity of the alveolar epithelium to phosgene is such that the pulmonary reaction can be identified after exposures at concentrations as low as 0.5 ppm for 120 min. The lowest exposure that produced a recognizable typical pulmonary lesion had a Ct value (concentration of the gas, in parts per million, multiplied by the time of exposure, in minutes) of 15 (e.g., 3 ppm for 5 min). Ct values producing pneumonitis are summarized in Table 8. The chronic pneumonitis is initially centered in the respiratory bronchiole and its evaginating alveoli. There is cellular thickening of these structures with the elaboration of new reticulin fibers. In addition to having thickened walls, the alveoli are often filled with desquamated cells (Gross et al., 1965).

Table 9 lists a number of additional phosgene inhalation studies conducted in animals (NIOSH, 1976).

Phosgene is reportedly responsible for the development of longterm lung disease in man, as well as emphysema and obliterative bronchiolitis in dogs (Cucinell, 1974; Galdston et al., 1947; Rossing, 1964). Although no quantitative data are available on the dosage that might cause permanent lung damage in man, it has been shown experimentally that exposure of dogs at 80-160 mg/m³ for 30 min every other day for a week causes interferences in lower airway resistance and, with continued exposure, permanent pathophysiologic changes within the lung (Rossing, 1964). According to Cucinell (1974), these concentrations are toxic and are 10-20 times greater than the dosage needed to produce pneumonitis (Table 8). The exact relation of phosgene exposure at these concentrations to human disease is unknown. Histopathologic ex mination of rats exposed at 0.2 and 1.0 ppm, 4 h/d, 5 d/wk for 2 wk at Haskell Laboratory (1976) showed no effects.

The Committee is unaware of any data on the carcinogenic effects of phosgene. However, exposure at high concentrations could conceivably lead to neoplasia related to scarring or regeneration of damaged lung tissue (NIOSH, 1976).

Cucinell (1974) reviewed aspects of the development of tolerance to phosgene in several species. For example, guinea pigs treated with low doses of phosgene—1.5 ppm (6 mg/m³) for 10 min—for 7 d became relatively resistant to toxic concentrations—35 ppm for 10 min (Cordier and Cordier, 1953b). Repeated exposure of cats to phosgene at 1.5—3.8 ppm or 5—6 ppm for 10 min every day had caused no greater lung damage after 40 d than after 2 d (Cordier and Cordier, 1953a). These animals were able to colerate a total Ct of 9,000 mg·min/m³ (total time, 400 min), even though the LCt50 for cats is about 2,000 mg min/m³ for 1 min (Cucinell, 1974). Tolerance to high loses of phosgene is believed by some to represent a manifestation of pathologic changes in the lungs (Cucinell, 1974). The Committee is unaware of any data on the development of tolerance to phosgene in man.

PHARMACOKINETICS

When phosgene (which is only slightly soluble in water) is inhaled at a moderate concentration, it does not react noticeably with the aqueous mucous film of the upper respiratory tract. Without decomposition, phosgene then reaches the alveolar region and interacts there with components of the blood-air barrier.

Immediate and irreversible damage occurs (Potts et al., 1949)-e.g., the membrane function breaks down, and fluid leaks from the capillaries into the interstitial space and then into the alveolar space, finally spreading to the traches. The duration of this process--the developmental phase or clinical latent period (Diller, 1978) -- depends on the inhaled dosage; the higher the dosage, the shorter the latent period. After moderate dosage, the clinical latent period may be about 6-15 h (Diller, 1978). Much of the current information on the metabolism of phosgene has been gained indirectly through recent studies of the in vitro metabolism of chloroform (Cresteil et al., 1979; Mansuy et al., 1977; Pohl et al., 1977) and carbon tetrachloride (Shah et al., 1979); that shows the intermediary formation of phosgene from both chlorinated hydrorarbons. For example, cysteine can form a stable adduct with chloroform metabolites when added to the incubation mixture (Mansuy et al., 1977; Pohl et al., 1977) and carbon tetrachloride (Shah et al., 1979); that shows

the intermediary formation of phosgene from both chlorinated hydrocarbons. For example, cysteine can form a stable adduct with chloroform metabolites when added to the incubation mixture (Mansuy et al, 1977; Pohl et al., 1977); this adduct is 4-carboxythiazolidine-2one, the reaction product of cysteine with phosgene. The reactions of phosgeve as a chloroform metabolite depend on which nucleophiles are present in the incubation medium (Cresteil et al., 1979; see Figure 3). Direct reaction with water leads to CO2, which is the final chloroform metabolite (Lavigne and Marchand, 1974; Paul and Rubinstein, 1963). Nucleophilic groups of microsomal macromolecules or free amino acids could react with phosgene and generate unstable acyl chlorides--e.g., carbamyl chlorides, chloroformates, or thiochloroformates, depending on whether the reacting group is amine, hydroxyl, or thiol (Cresteil et al., 1979). The reaction of these electrophilic intermediate acyl chlorides with water yields CO2 and regenerates the starting amino acid (or nucleophilic group of macromolecules). Finally, in this pathway, the nucleophilic group of amino acids catalyzes the hydrolysis of phosgene to CO2.

With the evidence that phosgene is the precursor of CO₂ from carbon tetrachloride (Shah et al., 1979), it was suggested that it might also be one of the reactive species that bind to lipids and proteins. Reynolds (1967) reported that ¹⁴COCl₂ given to rats was found in liver protein and to a smaller extent in lipids, but the pattern was quite different from that of ¹⁴CCl₄. Cessi et al., (1966) also reported that [¹⁴C]phosgene administered to rats labeled the liver proteins.

INHALATION EXPOSURE LIMITS

In 1980, ACGIH recommended a TLV for phosgene of 0.1 ppm for an 8-h working day (ACGIH, 1980). This figure is based on data obtained by the Chemical Warfare Service before 1921 that indicated that at 1 ppm phosgene may be safe for prolonged exposure (Cucinell, 1974). It is also based on the studies of Gross et al. (1965) that showed that exposure to phosgene at concentrations as low as 0.5 ppm for 2 h caused definitive pathologic changes in the lungs of rats killed 96 h after exposure. (Some abnormalities were considered to be present 3 mo after rats had been exposed at 2 ppm for 80 min.)

A safe concentration zone of 0.1-0.125 ppm was recommended for international adoption in 1968 by the Joint ILO/WHO Committee on Occupational Health (ILO/WHO, 1968). In 1980, ACGIH recommended a TLV of 0.1 ppm, on the basis of its irritating effects on the respiratory tract at slightly above 0.1 ppm, to which tolerance develops (ACGIH, 1980). Table 10 lists additional maximal allowable concentrations (MACs) for 15 countries (ACGIH, 1983, International Labour Office, 1970; Jpn. Assoc. Ind. Health, 1971; OSHA, 1983; Soc. Ital. Med. Lav., 1975; Winell, 1975). NIOSH recommended in 1976 that occupational exposure to phosgene not exceed 0.1 ppm, determined as a TWA concentration for up to a 10-h workday in a 40-h workweek, or 0.2 ppm, as a ceiling concentration for any 15-min period (NIOSH, 1976).

Cucinell (1974) suggested that the current standard of 0.1 ppm may be too high for 8 h/d, 5 d/wk at room temperature. It is also

believed that ambient concentrations of phosgene in industrial situations in the United States are considerably below the TLV. Industrial intoxication by phosgene has been due primarily to accidental exposure at high concentration. Cucinell (1974) further suggested that some adjustments in the acceptable concentration for the general population must be made for a continuous 24-h/d exposure, as opposed to single or intermittent exposure; as noted previously, animals tolerate intermittent exposures to phosgene better than continuous exposure. It was further noted that, although high doses of phosgene can cause chronic lung disease in man and animals, it is not certain whether low doses can aggravate pre-existing conditions or cause lung disease (Cucinell, 1974). Because the lowest experimental values available for long-term exposure suggest that 0.2 ppm for 5 h/d for 5 d may cause slight changes in the lung (Cameron et al., 1942b), Cucinell (1974) suggests that a value of 0.1 ppm would not be safe enough. If a safety margin of a factor of 10 is used, the environmental concentration should be 0.02 ppm, and that should not be exceeded by working personnel during an 8-h/d, 5-d/wk exposure. In support of the suggested ceiling concentration of phosgene, it was suggested that a Ct of 10 ppm.min is safe and that this application of Haber's law--constant toxic effect = (concentration) (duration of exposure) -- is valid. For an 8-h workday, the ceiling would be 0.02 ppm. Cucinell (1974) further proposed that, for a 24-h/d exposure, the ceiling should be lowered by about one-third to 0.006 ppm and an additional safety factor of 10 was recommended for situations in which the general population may be exposed (0.0006 ppm).

Tentative EELs for phosgene were reviewed by Zielhuis (1970), who suggested that the most relevant animal exposures to consider were those of Rinehart and Hatch (1964) and Gross et al. (1965). For example, when rats were exposed at 0.5-4 ppm for 5-480 min, death occurred when the Ct exceeded 180; at a Ct of 300, the mortality was 60%. There were no respiratory-function effects or observable pathologic changes at autopsy at a Ct of 15 (3 ppm for 5 min). Accordingly, the above information suggests that the Ct for man should remain below 15 when t is less than 60 min. Zielhuis (1970) proposed the following exposure limits:

Duration,	EEL,	EEL/TLV ratio
5	2.0	20
15	0.8	8
30	0.4	4
60	0.2	2

COMMITTEE RECOMMENDATIONS

In 1966, the Committee on Toxicology recommended EELs and CEL for phosgene.

On the basis of what appear to be the most relevant animal-exposure studies (Gross et al., 1965; Cameron et al., 1942a,b; Cameron and Foss, 1941; Rinehart and Hatch, 1964) and the rationale for lowered concentrations of phosgene as described by Cucinell (1974) and Zielhuis (1970), it appears prudent to propose lower EELs than the Committee recommended in 1966. The Committee has based its recommendations on studies done by Cameron and Foss (1941) and Cameron et al. (1942a,b) that show that animals do not tolerate phosgene at 0.2 ppm administered 5 h/d for 5 d (they developed slight pulmonary edema).

The present Committee's recommended EELs and CEL for phosgene and the limits proposed in 1966 are shown below.

	1956	1984
60-min EEL	1.0 ppm	0.2 ppm
24-h EEL	0.1 ppm	0.02 ppm
90-d CEL	0.05 ppm	0.01 ppm

TABLE 7 Phosgene Inhalation Exposure Effects on Humansa

No. Cases ^a	Exposure Variables	Duration of Exposure	Effects	Reference
109	Unknown 1 mo1 of phosgene Unknown ^b	Brief Brief 30 min	Pulmonary edema Pulmonary edema Pulmonary edema, death	Thiess and Goldmann, 1968
2	Unknown ^b Unknown ^b	Indefinite 3 h	Pulmonary edema Pulmonary edema	Gerritsen and Buschmann, 1960
1	Unknown (15 ppm)b,c	3.5 h	Pulmonary edema, death	Spolyar <u>et al.</u> , 1951
1	Unknown ^b	4.5 h	Acute broachitis	Glass <u>et al.</u> , 19
2	Unknown Unknown	Brief Brief	Bronchial irri- tation, death	Delephine, 1923
1	Unknown ^b	Brief	Pulmonary edema	Seidelin, 1961
7	Unknown	Brief	Acute bronchitis, delirium, pul- monary edema	Steel, 1942

a Data from NIOSH, 1976.
b Simultaneous exposure to chlorinated hydrocarbons.
c Recreated exposure and simulating accident.

TABLE 8 Pneumonitis Caused by Phosgene in Ratsa

Ct, ppm-min	Conc.,	Time,	Chronic Pneumonitisb
13	1.3	10	0
15	1.5	10	ŏ
24	0.8	30	+
27	0.9	30	+
33	1.1	30	_
36	1.2	30	+
40	1.0	40	+
48	0.8	60	++
48	0.8	60	+,P
54	0.9	60	++,P
60	0.5	120	+
84	1.4	60	+++,P
88	1.1	80	+
90	0.5	180	+
90	0.5	180	++
90	0.5	180	+
90	1.5	60	+++
96	0.8	120	+++
108	0.9	120	0,P
120	1.0	120	++,P
120	1.0	120	+
120	1.5	80	+
180	0.5	360	+
180	0.5	360	+
180	1.0	180	+
180	1.5	120	+++,P
192	1.2	160	++
198	1.1	180	+++
210	1.0	210	. ++
228	1.9	120	++
240	0.5	480	++,P
240	0.5	480	++
240	0.5	489	+++,P
240	1.0	240	++,P
342	1.9	180	++, <u>P</u>
360	1.0	360	+,F

a Data from Gross et al., 1965.
b 0 = no chronic pneumonitis; + = slight chronic pneumonitis; ++ = moderate chronic pneumonitis; +++ = severe chronic pneumonitis; P = acute pneumonia; and F = fibrinous pneumonia.

TABLE 9 Phosgene Inahalation Exposures and Effects in Animalsa

Species	Concentration, ppm	Duration of Exposure	Duration of Effects	Reference
Rat	55–100	10 min	Reduction in death rate from 74% to 33% by previous challenge	Box and Cullumbine, 1947
Rabbit	50-200	14-25 min	Decrease in sym- pathetic tone	Ivanhoe and Meyers, 1964
Rabbit	67	30 min	Pulmonary edema	Boyd and Perry, 1960
Dog	44-120	30 min	Pulmonary edema, pneumonia, emphysema, death	Underhill, 1920
Dog	72	30 min	Pulmonary conso- lidation, death	Durlacher and Bunting, 1947
Dog	24–40	30 min, 1 or 2 exposures, 1-3/week	Acute bronchio- litis	Clay and Rossing, 1964
Dog	24-40	30 min, 4-10 exposures, 1-3/week	Chronic bron- chiolitis	Clay and Rossing, 1964
Dog	24-40	30 min, 30-40 exposures, 1-3/week	Emphysema	Clay and Rossing, 1964
Cat and guinea pi	2.5-6.25 8	10 min/d, 2-41 d	Pulmonary edema, bronchitis, bronchopneumonia, death	Cordier and Cordier, 1953b

Data from NIOSH, 1976.

b Animals were pregassed at 20 ppm for 10 min to determine effect of pregassing on response to later challenge.

TABLE 10

Maximum Allowable Concentrations (MACs) for Phosgene

Country	<u>Year</u> a	MAC, mg/m ³	Reference
United States	1974	0.4	ACGIH, 1983
United States (OSHA)	1974	0.4	OSHA, 1983
West Germany	1974	0.4	Winell, 1975
East Germany	1973	0.5	Winell, 1975
Sweden	1975	0.3 (ceiling)	Wine11, 1975
CSSR (Czechoslovakia)	1969	0.4	Winell, 1975
USSR	1972	0.5 (ceiling)	Wine11, 1975
Italy	1975	0.4	Soc. Ital. Med. Lav., 1975
Japan	1969	0.4	Jpn. Assoc. Ind. Hlth., 1971
Bulgaria	NG	0.5	Inter. Lab. Off., 1970
Finland	NG	4	Inter. Lab. Off., 1970
Hungary	NG	0.5	Inter. Lab. Off., 1970
Poland	NG	0.5	Inter. Lab. Off., 1970
Rumania	NG	0.5	Inter. Lab. Off., 1970
United Arab Republic	NG	4	Inter. Lab. Off., 1970
Yugoslavia	NG	0.4	Inter. Lab. Off., 1970

a NG = Not given.

REFERENCES

American Conference of Governmental Industrial Hygiensts. 1980. Documentation of the Thresold Limit Values. 4th ed., Clacinnati, Ohio: American Conference of Governmental Industrial Hygienists. pp. 336-337.

American Conference of Governmental Industrial Hygienists. 1983. TLVs^R. Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Hygienists 93 p.

Anonymous. 1980. Phosgene. Chem. Mark. Rep. 217(3):9.

Box, G.E.P., and Cullumbine, H. 1947. The effect of exposure to sub-lethal doses of phosgene on the subsequent L(Ct)50 for rats and mice. Br. J. Pharmacol. 2:38-55.

Boyd, E.M., and Perry, W.F. 1960. Respiratory tract fluid and inhalation of phosgene. J. Pharm. Pharmacol. 12:726-732.

Brown, J.E., and Birky, M.M. 1980. Phosgene in the thermal decomposition products of poly(vinylchoride): Generation, detection and measurement. J. Anal. Toxicol. 4:166-174.

Cameron G.R., and Foss, G.L. 1941. Effects of low concentrations of phosgene repeated for 5 hours on 5 consecutive days in groups of different animals. Porton Report No. 2316, Serial No. 63. Washington, D.C.: British Defense Staff, British Embassy.

Cameron, G.R., Courtice, F.C., and Foss, G.L. 1942a. Effect of exposing different animals to a low concentration of phosgene 1:1,000,000 for 5 hours. Section IX in First Report on Phosgene Poisoning. Porton Report No. 2349, Washington, D.C.: British Defense Staff, British Embassy.

Cameron, G.R., Courtice, F.C., and Foss, G.L. 1942b. Effect of exposed different animals to a low concentration of phosgene 1:1,000,000 for 5 hours on 5 consecutive days. Section VIII in First Report on Phosgene Poisoning. Porton Report No. 2349, Washington, D.C.: British Defense Staff, British Embassy.

Carotti, A.A., and Kaiser, E.R. 1972. Concentrations of twenty gaseous chemical species in the fuel gas of a municipal incinerator. J. Air Pollut. Control Assoc. 22:248-253.

Cessi, C., Colombini, C., and Mameli, L. 1966. The reaction of liver proteins with a metabolite of carbon tetrachloride. Biochem. J. 101:46C-47C.

Ta ta ta ta taun ta taun kata darahar ta mata karangan ka

Chadwick, D.H., and Cleveland, T.H. 1981. Isocyanates, organic. In G.J. Burkey, C.I. Eastman, A. Klingsberg, L. Spiro, and M. Wainwright, eds., Kirk-Othmer Encyclopedia of Chemical Technology. 3rd rev. ed., Vol. 13. New York: Interscience Publishers. p. 789-818.

Chadwick, D.H., and Hardy, E.E. 1967. Isocyanates, organic. In E.A. Parolla, C. Coeman, C. Comiskey, A. Klingsberg, and P. van Reyen, eds., Kirk-Othmer Encyclopedia of Chemical Technology. 2nd rev. ed., Vol. 12. New York: Interscience Publishers p. 54-58.

Clay, J.R., and Rossing, R.G. 1964. Histopathology of exposure to phosgene. Arch. Pathol. 78:544-551.

Compressed Gas Association, Inc. 1981. Handbook of Compressed Gases. 2nd ed. New York: Van Nostrand Reinhold Co. p. 430-435.

Cordier, D., and Cordier, G. 1953a [Do repeated inhalation of weak concentration of phosgene sensitize the organism to a stronger concentration?] Compt. Rend. Sco. Biol. 147:327-330.

Cordier, D., and Cordier, G. 1953b. The toxicity of weak concentration of phosgene after repeated inhalations. J. Physiol. 45:421-428.

Cresteil, T., Beaune, P., Leroux, J.P., Lange, M., and Mansuy, D. 1979. Biotransformation of chloroform by rat and human liver microsomes; in vitro effects on some enzyme activities and mechanism of irreversible binding to micromolecules. Chem. Biol. Interactions 24:153-165.

Cucinell, S.A. 1974. Review of the toxicity of long-term phosgene exposure. Arch. Environ. Health 28:272-275.

Delephine, S. 1923. Summary of notes on two fatalities due to inhaling phosgene (COCl₂). J. Ind. Hyg. 4:433-440.

Diller, W.F. 1978. Medical phosgene problems and their possible solution. J. Occup. Med. 20:189-193.

Durlacher, S.H., and Bunting, H. 1947. Pulmoanry changes following exposure to phosgene. Am. J. Pathol. 23:679-693.

Gafafer, W.M., ed. 1966. Occupational Diseases: A Guide to Their Recognition. Washington, D.C.: U.S. Department of Health, Education, and Welfare, Public Health Service. p. 201 [Public Health Service Publ. No. 1097]

Galdston, M., Luetscher, J.A., Jr., Longcope, W.T., and Ballich, N.L. 1947. A study of the residual effects of phosgene poisoning in human subjects. II. After chronic exposure. J. Clin. Invest. 26:169-181.

Gay, B.W., Jr., Hanst, P.L., Bufalini, J.J., and Noonan, R.C. 176. Atmsopheric oxidation of chlorinated ethylenes. Environ. Sci. Technol. 10:53-67.

Gerritsen, W.B., and Buschmann, C.H. 1960. Phosgene poisoning caused by the use of chemical paint removers containing methylene chroride in ill-ventilated rooms heated by kerosene stoves. Br. J. Ind. Med. 17:187-189.

Glass, W.I., Harris, E.A., and Whitlock, R.M. 1971. Phosgene poisoning: Case report. N.Z. Med. J. 74:386-389.

Gross, P., Rinehart, W.E., and Hatch, T. 1965. Chronic pneumonitis caused by phosgene. Arch Environ. Health 10:768-775.

Hardy, E.E. 1982. Phosgene. In: C.I. Eastman, A. Klingsburg, L. Spiro, G. Wachsman, and M. Wainwright, eds. Kirk-Othmer Encyclopedia of Chemical Technology. 3rd rev. ed., Vol. 17. New York: Interscience Publishers. p. 416-425.

Haskell Laboratory. 1976. Report No. 223-76. Newark, Delaware: E.I. DuPont de Nemours and Co.

Henderson, Y., and Haggard, H.W. 1943. Noxious Gases and the Principles of Respiration Influencing Their Action. Phosgene. 2nd and rev. ed. New York: Reinhold Publishing Corp. p. 137-138.

Hygienic Guide Series. 1968. Phosgene (carbonyl chloride). Am. Ind. Hyg. Assoc. J. 29:308-311.

International Labour Office/World Health Organization. 1968. Permissible Levels of Occupational Exposure to Airborne Toxic Substances. Sixth Report of the Joint IIO/WHO Committee on Occupational Health. World Health Organization Technical Report Series No. 415. Geneva: World Health Organization. 16 p.

International Labour Office. 1970. Permissible Levels of Toxic Substances in the Working Environment. Sixth Session of the Joint ILO/WHO Committee on Occupational Health. Occupational Safety and Health Series 20. Geneva: ILO. 405 p.

Ivannoe, F., and Meyers, F.H. 1964. Phosgene poisoning as an example of neuropralytic acute pulmonary edema: The symathetic vasomotor reflex involved. Dis. Chest 46:211-218.

Japanese Association of Industrial Health. 1971. Recommendations for permissible concentrations, etc. Trans. of Sangyo Igaku (Jpn. J. Ind. Health 13:475-484).

Lavigne, J.-G., and Marchand, C. 1974. The role of metabolism in chloroform hepatotoxicity. Toxicol. Appl. Pharmacol. 29:312-326.

Long, J.E., and Hatch, T.F. 1961. A method for assessing the physiological impairment produced by low-level exposure to pulmonary irritants. Am. Ind. Hyg. Assoc. J. 22:6-13.

Mansuy, D., Beaune, P., Cresteil, T., Lange, M., and Leroux, J.-P. 1977. Evidence for phosgene formation during liver microsomal oxidation of chloroform. Biochem. Biophys. Res. Commun. 79:513-517.

Moore, S., and Gates, M. 1946. Phosgene. In: Summary Technical Report of Division 9, NDRC. Vol. 1 Chemical Warfare Agents, and Related Chemical Problems. Par 1. Washington, D.C.: Office of Scientific Research and Development, N tional Defense Research Committee. p. 17-20.

National Institute for Occupational Safety and Health. 1976. NIOSH Criteria for a Recommended Standard...Occupational Exposure to Phosgene. Cincinnati, Ohio: U.S. Department of Health, Education and Welfare, Public Health Service, Center for Disease Control, National Institute for Occupational Safety and Health. 129 p. [HEW Publ. No. (NIOSH) 76-137]

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air Contaminants 29 CFR 1910.1000.

Patty, F.A. 1963. Phosgene, COC1. In D.W. Fassett, and D.D. Irish, eds.. Patty's Industrial Hygiene and Toxicology. Vol. II Toxicology. 2nd rev. ed. New York: Interscience Publishers. p. 938-940.

Paul, B.B., and Rubinstein, D. 1963. Metabolism of carbon tetrachloride and chloroform by the rat. J. Pharmacol. Exp. Ther. 141:141-148.

Pohl, L.R., Bhooshan, B., Whittaker, N.F., and Krishna, G. 1977. Phosgene: A metabolite of chloroform. Biochem. Biophys. Res. Commun. 79:684-691.

Polednak, A.P. 1980. Mortality among men occupationally exposed to phosgene in 1943-1945. Environ. Res. 22-357-367.

Potts, A.M., Simon, F.P., and Gerard, R.W. 1949. The mechanism of phosgene and diphosgene. Arch. Biochem. 24-329-337.

Reynolds, E.S. 1967. Liver parenchymal cell injury. IV. Pattern of incorporation of carbon and chlorine from carbon tetrachloride into chemical constituents of liver in vivo. J. Pharmcol. Exp. Ther. 155:117-126.

Rinehart, W.E., and Hatch, T. 1964. Concentration-time product (CT) as an expression of dose in sublethal exposures to phosgene. Am. Ind. Hyg. Assoc. J. 25:545-553.

Rossing, R.G. 1964. Physiologic effects of chronic exposure to phosgene in dogs. Am. J. Physiol. 207:265-272.

Sax, N.I. 1968. Dangerous Properties of Industrial Materials. 3rd ed., New York: Reinhold Book Corp. 1251 p.

Seidelin, R. 1961. The inhalation of phosgene in a fire extinguisher accident. Thorax 16:91-93.

Shah, H., Hartman, S.P., and Winhouse, S. 1979. Forestion of carbonyl chloride in carbon tetrachloride metabolism by rat liver in vitro. Cancer Res. 39:3942-3947.

Singh, H.B. 1976. Phosgene in ambient air. Nature 264:428-429.

Societa Italiani di Medicina de Lavoro, Associazione Italiana degli Igienisti Industriali, Valori Limite Ponderati degli Inquinanti Chimici e Particolati degli Ambienti di Laoro per il. 1975. Med. Lav. 66:361-371.

Spolyar, L.W., Harger, R.N. Keppler, J.F., and Bumsted, H.E. 1951. Generation of phosgene during operation of trichloroethylene degreaser. AMA. Arch. Ind. Hyg. 4:156-160.

SRI International. (not dated). Chemical Economics Handbook (CEH). Menlo Park, CA: Standard Research Institute International. [35 vols. loose leaf]

SRI International. 1983. 1983 Directory of Chemical Producers, USA, Menlo Park, CA: Stanford Research Institute International.

Stavrakis, P. 1971. The use of hexamethylenetetramine (HMT) in treatment of acute phosgene poisoning. Ind. Med. Surg. 40:30-31.

Steel, J.P. 1942. Phosgene poisoning. Report of two cases. Lancet 1:316-317.

Theiss, A.M., and Goldmann, P.J. 1968. Ist die Phosgenvergiftung noch ein arbeitsmedizinisches Problem? Zentr. ARbeitsmed. Arbeitsschutz 18:132-141.

Underhill, F.P. 1920. The Lethal War Gases: Physiology and Experimental Treatment. New Haven: Yale University Press. p. 1-21, 40-84.

Weast, R.C., and Astle, M.J., eds. 1978-1979. CRC Handbook of Chemistry and Physics. A Ready-Reference Book of Chemical and Physical Data. 59th ed. West Palm Beach, Florida: CRC Press. p. C-434.

Wills, W.J.H.B., McParland, C.W., and Webster, R.E. 1938. The detection of phosgene by odor. EATR 250. Aberdeen Proving Ground, MD; Edgewood Arsenal, March.

Winell, M. 1975. An international comparison of hygienic standards for chemicals in the work environment. Ambio 4:34-35.

Zielhuis, R.L. 1970. Tentative emergency exposure limits for sulphur dioxide, sulphuric acid, chlorine and phosgene. Ann. Occup. Hyg. 13:171-176.

SODIUM HYDROXIDE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: NaOH Molecular weight: 40.01

Synonyms: White caustic, caustic soda,

soda lye, lye

CAS number: 1310-73-2
Melting point: 318.4°C
Beciling point: 1390°C
Specific gravity: 2.13

Solubility: Soluble in water, ethanol and

glyce.ol; insoluble in acetone

and ether

General characteristics: White deliquescent solid

OCCURRENCE AND USE

Sodium hydroxide is widely used in the manufacture of soaps, paper, rayon, cellophane, mercerized cotton, aluminum, and many chemicals. It is also used in petroleum refining, degreasing, etching, zinc extraction, tin plating, oxide coating, and food processing (for peeling fruits and vegetables). In concentrated form, it is used as a drain cleaner. Sodium hydroxide has been used in the management of pleural effusions.

SUMMARY OF TOXICITY INFORMATION

Sodium hydroxide toxicity depends on the concentration of the sodium hydroxide solution and the duration of its contact with tissue. The chemical acts locally, exerting a strong corrosive action whose mechanism is not known, and causes almost immediate degeneration of the tissue, which can result in rapid absorption of sodium hydroxide into the circulating system and distribution with the body water. It dissociates completely in water, blood, and cytoplasm and is not metabolized.

Because toxicity of sodium hydroxide is determined primarily by concentration of the hydroxyl ion, the total sikalinity is reduced when sodium carbonate (Na₂CO₃) forms. Therefore, measurement of sodium ion concentration is not always an accurate indication of alkalinity and toxic potential (Cooper et al., 1979).

EFFECTS ON HUMANS

Humans can be exposed during the manufacture of sodium hydroxide and in the handling of sodium hydroxide as a solid or concentrated solution.

Sodium hydroxide is corrosive to all body tissues; concentrated vapors cause serious damage to the eyes and respiratory system. Ingestion of sodium hydroxide, which occurs frequently in children, can cause severe necrosis, with stricture of the esophagus and death. Contact with the skin can result in dermetitis, loss of hair, and necrosis due to irritation. Skin types vary in sensitivity to caustic irritation.

Airborne mists of sodium hydroxide around degreasing vats (at 200°F) containing Seco 75 or Tysol 810 (concentrated sodium hydroxide solutions combined with chelating and wetting agents) were associated with irritation of the upper respiratory tract in workers exposed to sodium hydroxide at 0.01-0.7 ag/m³ (mean, 0.1-0.15 mg/m³). The work area also contained vapors of Stoddard solvent, ENSIS 254 oil, Zyglc, and Hagna-flux solutions (Stoddard solvent air concentrations were 13-780 mg/m³) and, at vat-cleaning time, sulfuric acid at 0.1-0.6 mg/m³ (Hervin and Cohen, 1974). The effect of these chelating agents and solvents on sodium hydroxide toxicity is not known.

Ott et al. (1977) assessed mortality rates among chemical-plant employees exposed chronically to caustic dust. Records of acuce exposures (unknown concentrations) indicated that the caustic materials in the plant had caused mild to severe responses (irritation, erythema, and "objective damage to organs") in skin, eyes, and the respiratory system. No indication of purity was given, but it was noted that addium chloride and sodium carbonate were known to be included in the caustic exposures; workers known to be exposed to arsenicals or asbestos, as well as caustic, were excluded from the study. According to the results of a study that measured total alkalinity of air samples in the workplace, sodium hydroxide concentrations of up to 6.7 mg/m^3 in one area of the plant correlated well with subjective response data that indicated increasing respiratory irritation with increasing alkali concentration; in a second area of the plant, where sodium hydroxide content was up to 7.7 mg/m 3 , the correlation was poor. No explanation for the poor correlation could be found. Sodium hydroxide estimated to be as high as 2 mg/m³ (TWA) did appear to cause masal and skin irritation, especially in plant areas with high temperatures. No correction factor for temperature effects on sodium hydroxide toxicity is available, but it is expected that increased temperatures would increase toxicity.

Controlled exposures to sodium hydroxide solutions have been limited to dermal application, usually to the forearms of volunteers. Malten and Spruit (1966) placed 0.12% (0.03 M) or 0.27% (0.0675 M) solutions of sodium hydroxide in cups fixed to the forearms of human volunteers. Erythema was produced within 0.5 h by the stronger solution and within 1 h by the more dilute preparation (see also Spruit and Malten, 1968). Marzulli and Maibach (1975) applied sodium hydroxide solutions to the backs of human subjects in occluded patches renewed daily for 21 d. The lowest concentration studied (0.05%, 0.0125 M) produced no erythema, a 0.5% (0.125 M) solution was mildly irritating, and solutions of 4% or 5% were severely irritating. Similar results were observed in rabbits.

EFFECTS ON ANIMALS

Many toxicologic evaluations of sodium hydroxide have been carried out in animals; the studies have focused on eyes, skin, and lungs as targets. These investigations are summarized in Table 11.

INHALATION EXPOSURE LIMITS

The current ACGIH (1983) ceiling limit for sodium hydroxide is 2 mg/m^3 (TWA); this concentration is also used by NIOSH. Exposure ceilings in West Germany, Finland and Yugoslavia are 2 mg/m^3 . The concentration appears to be based on an undocumented comment by Patty (1949) that 2 mg/m^3 is thought to cause noticeable, but not excessive, respiratory irritation.

COMMITTEE RECOMMENDATIONS

Few inhalation studies to evaluate the toxicity of sodium hydroxide have been reported. The one detailed analysis of workers in a degreasing plant exposed to heated caustic vapor indicated that aerosols containing an average of 0.1 mg/m³ were reversibly irritating to the upper respiratory tract (Hervin and Cohen, 1974); the workers in that study were also exposed to detergent and organic solvents, so evaluation of the toxicity of sodium hydroxide alone is difficult. Sodium hydroxide at 2 mg/m³ (TWA) appeared to cause nasal and skin irritation, especially at high temperatures. Because increased temperature increases toxicity, the Committee concludes that a 1-h exposure to sodium hydroxide at 2 mg/m³ would probably produce no more than a reversible mild irritation of eyes, skin, and respiratory system.

The Committee's previous recommendations for sodium hydroxide exposure limits were made in 1965 on the basis of an undocumented statement that 6.0 mg/m³ produced intolerable respiratory discomfort; the 1965 recommendation was for 10- and 30-min EELs of 4 mg/m³ and a 60-min EEL of 2 mg/m³. Little new information on the toxicity of sodium hydroxide by inhalation is available; experimental-animal studies have concentrated on topical application to eyes and skin, and extrapolation from such studies to human inhalation is difficult. On the basis of the report on workers in a degreasing plant (Hervin and Cohen, 1974), an EEL above 2 mg/m³ may produce much nasal and skin discomfort, especially at high temperature, but no available inhalation report provides a basis on which to establish an EEL for sodium hydroxide with confidence.

The present Committee's recommended EELs for sodium hydroxide and the limits proposed in 1965 are shown below.

	1965	1984
10-min EEL	4 mg/m^3	2 mg/m^3
30-min EEL	4 mg/m^3	2 mg/m^3
60-min EEL	2 mg/m^3	2 mg/m^3

TABLE 11

Summary of Data on Toxicity of NaOH in Animals

Reference	Davidson, 1927	Bromberg et al., 1965	Cosgrove and Hubbard, 1928	Hubbard, 1937; Hubbard, 1938	Hughes, 1946	Grant and Keyn, 1955	Brown, 1971; Brown and Weller, 1970; Brown et al., 1969ª, b, c; 1970	Chiang et al., 1971
Effects	Edema, sloughing Edema, sloughing Edema, limited sloughing	No burn, no dead Burn, 1/5 dead Burn, 2/5 dead Burn, 4/5 dead Burn, 5/7 dead	Necrosis, death	Necrosis	Necrosis	Slight injury Necrosis	Necrosis	Intraocular pressure up Intraocular pressure up Intraocular pressure up
Duration of Exposure	l min, no wash l min, HA _C wash l min, H2 ^O wash	Immediate rinse 30 min, rinse 1 h, rinse 2 h, rinse No wash	NG &	10 s	3 min	15 min 15 min	30 s	Not specified Not specified Not specified
Dose	50%	50%	707	20%	0.2%	pH 11 pH 12	28	0.52b 2.02b 8.02b
Route	Skin	Skin	Eye	Eye	Eye	Eye	Eye	Eye
Species	Pat	Mouse	Rat	Rabbit	Rabbit	Rabbit	Rabb1 t	Rabbí t

TABLE 11 (Continued)

,	,		Duration of		
Species Route	Route	Dose	Exposure	Effects	Reference
Rabbit	Eye	2%	H ₂ 0 rinse, 1 min H ₂ 0 rinse, 4 min	Corneal perforations Corneal perforations	Geeraets et al., 1966
Rabbit	Eye	4%c	1 s 2 s	No injury Corneal swelling	Shapiro, 1956
Rat	Inhala- tion	Inhala- 40% soln. ^d tion	2.5 шое	Degeneration of pulmonary tissue, undefined "tumors"	Dluhos et al., 1969
Rat	Inhala- tion	Inhala- 40% soln. tion aerosolf	l mo	Death	Vyskocil et al., 1966
		20% soln.	1 по	Degeneration of	
		aerosol ^r 10% soln. aerosol ^f	1 по	pulmonary tissue Little change in pulmonary	
		5% soln. aerosolf	1 по	tissue Little change in pulmonary tissue	

a 0.5 ml applied.

b in impregnated paper. c Aerosol; 80% of particles under l $_{\rm pm}$ in diameter. d 30 min twice a day for 2.5 mo with 10-d rest after third week. f Pwice a week.

Twice a week.

REFERENCES

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Bromberg, B.E., Song, I.C., and Walden, R.H. 1965. Hydrotherapy of chemical burns. Plast. Reconstr. Surg. 35:85-95.

Brown, S.I. 1971. Treatment of the alkali-burned cornea. Sight Sav. Rev. 41:83-88. [Ind. Medicus 13:1052, 1972]

Brown, S.I., and Weller, C.A. 1970. Collagenase inhibitors in prevention of ulcers of alkali-burned cornea. Arch. Ophthalmol. 83:352-353.

Brown, S.I., Akiya, S., and Weller, C.A. 1969a. Prevention of the ulcers of the alkali-burned cornea - Preliminary studies with collagenase inhibitors. Arch. Ophthalmol. 82:95-97.

Brown, S.I., Wassermann, H.E., and Dunn, M.W. 1969b. Alkali-burns of the cornea. Arch. Ophthalmol. 82:91-94.

Brown, S.I., Weller, C.A., and Akiya, S. 1970. Pathogenesis of ulcers of the alkali-burned cornea. Arch. Ophthalmol. 83:205-208.

Brown, S.I., Weller, C.A., and Wassermann, H.E. 1969c. Collagenolytic activity of alkali-burned corneas. Arch. Ophthalmol. 81:370-373.

al account to be seen for the second to be seen to be s

Chiang, T.S., Moorman, L.R., and Thomas, R.P. 1971. Ocular hypertensive response following acid and alkali burns in rabbits. Invest. Ophthalmol. 10:270-273.

Cooper, D.W., Underhill, D.W., and Ellenbecker, M.J. 1979. A critique of the U.S. standard for industrial exposure to sodium hydroxide aerosols. Am. Ind. Hyg. Assoc. J. 40:365-371.

Cosgrove, K.W., and Hubbard, W.B. 1928. Acid and alkali burns of the eye - An experimental study. Ann. Surg. 87:89-94.

Davidson, E.C. 1927. The treatment of acid and alkali burns - An experimental study. Ann. Surg. 85:481-489.

Dluhos, M., Sklensky, B., and Vyskocil, J. 1969. Effect of aerosol inhalation of soda hydroxide on the respiratory tract of rats. Vnitr. Lek. 15:38-42. [CA 70:80666e, 1969]

Geeraets, W.J., Aaron, S.D., and Guerry, D., III. 1966. Alkali burns of the cornea and neutral ammonium tartrate - An in vitre and in vivo study. Guildcraft 40:13-16.

Grant, W.M., and Kern, H.L. 1955. Action of alkalies on the corneal stroma. Arch. Ophthalmol 54:931-939.

Hervin, R.L., and Cohen, S.R. 1974. Health hazard evaluation/toxicity determination report 72-97-135, Chicago and North Western Railway, Oelwein, Iowa. Rockville, Maryland: National Institute for Occupational Safety and Health. 18 p. [NIOHS-TR-HHE 72-97-135 and PB-146 442]

Hubbard, W.B. 1937. Treatment of caustic burns of the eye. Arch. Ophthalmol. 18:263-266.

Hubbard, W.B. 1938. Caustic burns of the eye. Arch. Ophthalmol. 19:968-975.

Hughes, W.F., Jr. 1946. Alkali burns of the eye. II. Clinical and pathologic course. Arch. Ophthalmol. 36:189-214.

Malten, K.E., and Spruit, D. 1966. Injury to the skin by alkali and its regeneration. Dermatologica 132:124-130.

Marzulli, F.N., and Maibach, H.I. 1975. The rabbit as a model for evaluating skin irritants: A comparison of results obtained on animals and man using repeated skin exposures. Food Cosmet. Toxicol. 13:533-540.

Ott, M.G., Gordon, H.L., and Schneider, E.J. 1977. Mortality among employees chronically exposed to caustic dust. J. Occup. Med. 19:813-816.

Patty, F.A. 1949. Sodium hydroxide. In: Patty, F.A., ed. Industrial Hygiene and Toxicology, Vol. II. New York: Interscience Publ. p. 560-561.

Shapiro, H. 1956. Swelling and dissolution of the rabbit cornea in alkali. Am. J. Ophthalmol. 42:292-298.

Spruit, D., and Malten, K.E. 1968. Estimation of the injury of human skin by alkaline liquids. Berufs-Dermatosen 16:11-24. [Ind. Medicus 9:8578, 1968]

Vyskocil, J., Tuma, J., and Dluhos, M. 1966. Effect of aerosol inhalations of sodium hydroxide on the elimination of quartz dust from lungs of rats. Scr. Med. Fac. Med. Univ. Brun. Purkynianae 39:25-29. [Chem. Abs. 65:4519d, 1966]

SULFUR DIOXIDE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

Chemical formula: SO2 Molecular weight: 64.07

CAS number: 7446-09-5

Specific gravity (liquid): 1.434 Specific gravity (gas): 2.927

Solubility: Soluble in water, alcohols,

acetic acid, and sulfuric acid

General characteristics: Colorless, nonflammable gas or

liquid; strong suffocating odor

1 ppm = 2.6 mg/m³ 1 mg/m³ = 0.38 ppm Conversion factors:

OCCURRENCE AND USE

Sulfur dioxide is most noteworthy as an environmental pollutant. It is formed when materials containing sulfur are burned, and is thus an important air pollutant, especially in the vicinity of smelters and plants burning soft coal or high sulfur oil. Others are automobile exhaust, wood-burning stoves, pulp mills, and smelters. Note that, in addition to sulfur dioxide itself, many related compounds and decay products of sulfur dioxide -- such as sulfurous and sulfuric acids, sulfates, sulfites, and bisulfites -- are present in the ambient air. It is beyond the scope of this brief review to describe all the information on all these substances; only sulfur dioxide will be addressed.

SUMMARY OF TOXICITY INFORMATION

Within the last 8 yr, three reviews of the toxic effects of sulfur dioxide in humans and animals have been published (Greenfield, Attaway and Tyler, 1976; International Electric Research Exchange, 1981; EYA, 1982). These reviews covered epidemiologic reports, health effects of acute and chronic low-dose exposures of humans, health effects of acute and chronic low- and high-dose exposures of animals, projected estimates of pollutant thresholds for adverse effects of short- and long-term exposures, and correlations of sulfur dioxide exposures with asthmatic attacks and effects on children.

EFFECTS ON HUMANS

Few recent reports have described accidental exposure to sulfur dioxide. Of historical interest is the report of a worker exposed directly to liquid sulfur dioxide who suffered effects of acute freezing of the skin and corneas (Kennon, 1927). A more recent report cited a case of an accidental exposure to sulfur dioxide at a high concentration for 15-20 min that led to the development of a severe, irreversible obstructive syndrome (Woodford et al., 1979). The pulmonary effects of accidental exposure to sulfur dioxide appear to be limited to gaseous sulfur dioxide.

An excellent review of studies involving controlled human exposure is available (Greenfield, Attaway and Tyler, 1976). Briefly, the major health effects of exposure to sulfur dioxide at less than 25 ppm (for various durations) are irritation of mucous membranes, throat, esophagus, and eyes; reflex cough; increase in respiratory rate associated with decrease in depth of respiration; decrease in nasal mucus flow; variable effects on tracheal and bronchial mucus flow; decrease in forced expiratory volume and flow; decrease in airway conductance; and increase in airway resistance.

An estimated 10-20% of the population will respond with hyperreactivity on exposure to sulfur dioxide. Koenig et al. (1980) have shown that asthmatic adolescents are more sensitive than healthy nonsmoking adults to sulfur dioxide at 1 ppm. As demonstrated by Lawther et al. (1975), the changes in airway resistance after exposure to sulfur dioxide at less than 30 ppm are short-lived. Greenfield, Attaway and Tyler, (1976) reviewed similar changes after short-term exposures to sulfur dioxide at concentrations greater than 25 ppm.

Another review (EPA, 1982) indicated that recovery to normal functional values can occur in as little as 5 min after acute exposure of normal resting subjects (Lawther et al., 1975), but may take 30-60 min after exposure during exercise (Bates and Hazucha, 1973), after exposure of exercising asthmatic subjects (Sheppard et al., 1981; Koenig et al., 1981), or after exposure of other sensitive subjects (Gokemeijer et al., 1973; Lawther et al., 1975). Weir and Bromberg (1975) suggested that persons with minimal airway disease are probably not more susceptible than normal persons to the effects of sulfur dioxide. However, this latter conclusion is questionable, in that the presence of pre-existing disease in the subjects interfered with attempts to define threshold concentrations of sulfur dioxide. Asthmatic subjects exposed to sulfur dioxide at 5 ppm for 5 min while exercising have had asthmatic attacks (Sheppard et al., 1980).

Epidemiologic data on sulfur dioxide are characteristically difficult to assess, because of the spectrum of pollutants and particles associated with ambient sulfur dioxide. A number of acute air-pollution episodes in this century have been associated with increased mortality. Incidents in the Meuse Valley, Donora (Pennsylvania), 2 ndcs, and New York City have provided evidence that increased pollution has observable effects on human health (Greenfield, Attava, and Tyler, 1976). Retrospective studies of these events have demonstr. that deaths most often occurred among persons over 45 who were already suffering from chronic heart or lung disease; the effects of sulfur dionice alone were not assessed. Lung-function measurements made daily on four normal subjects and two with bronchitis in London showed that daily variations in lung function were small and were related to respiratory infections (EPA, 1982). Concentrations of sulfur dioxide correlated with variations in peak flow rates and airway resistance. Increased sulfur dioxide pollution has been related to decreases in lung function (FEV1) in children. Several

long-term epidemiologic studies of chronic effects have found that populations living in areas characterized by high concentrations of particulate matter and sulfur dioxide tend to have a higher prevalence of respiratory illness and decreased lung function than groups in areas with lower pollution. However, the influence of particles and other factors associated with ambient sulfur dioxide confounds interpretation (EPA, 1982).

In one occupational study, the frequency of chromosomal aberrations was significantly increased among workers at a sulfite pulp factory in Sweden (Nordenson et al., 1980). This increase was found to be associated mainly with exposure to sulfur dioxide (boiling of sulfite pulp and handling of sulfuric acid) and not with chlorine and dust in other workplaces in the factory.

EFFECTS ON ANIMALS

Alarie et al. (1972) reported an accidental exposure of cynomologus monkeys to sulfur dioxide at an estimated 200-1,000 ppm for 1 h. No animals died, but a deterioration in pulmonary function persisted for the following 48 wk of the study. Histopathologic lesions included thickening of alveolar walls, hyperplasia of bronchial epithelium, and bronchiolar plugging with proteinaceous material, macrophages, and leukocytes.

A comprehensive review (Greenfield, Attaway and Tyler, 1976) of published literature on controlled animal exposures to sulfur dioxide may be summarized as follows: Acute exposure of animals results in diminished pulmonary function similar to that seen in man, decreased tracheal mucus clearance, pneumonia, and lesions of the nasomaxillary turbinates. Acute effects generally are transient at lower sulfur dioxide concentrations (20 ppm). In various species, the effects of exposures at over 100 ppm appear to be time-dependent and range from transient physiologic manifestations to death.

Transient adverse effects on pulmonary function have been demonstrated in rabbits exposed to sulfur dioxide at 200 ppm for 3 h (Davies et al., 1978). Rats exposed at 400 ppm died after 22 h of exposure (Asmundsson et al., 1973). Rats exposed at 567 ppm 6 h/d died after 12 d (Laskin et al., 1970). Adaptation apparently occurs if recovery between exposure periods is sufficient.

Subchronic exposure (less than 90 d) of animals to sulfur dioxide has been reviewed (Greenfield, Attaway and Tyler, 1976). Studies in which animals were exposed at concentrations below 10 ppm have demonstrated functional abnormalities that are reversible. Studies with subacute exposures at higher concentrations are lacking.

Chronic exposures (over 90 d) of a variety of species to sulfur dioxide at both low and high concentrations have been reported. The notable effects of exposure of dogs at 500 ppm 2 h/d for 3-5 mo include hypersecretion of mucus, goblet-cell hyperplasia, bronchial-gland hypertrophy, increase in pulmonary resistance, decrease in airway responsiveness to inhaled mediators, and decrease in tracheal mucus clearance (Drazen et al., 1982; Chakrin and Saunders, 1974; Greene et al., 1982; and Islam et al., 1977). Early exposure effects, such as ocular and mucous-membrane irritation, diminished with

repeated exposure; that indicates the existence of an adaptation mechanism.

Sulfur dioxide is not generally considered to be carcinogenic. One study (Laskin et al., 1970) has demonstrated a higher incidence of squamous-cell carcinomas of the lung in rats that inhaled benzo[a]pyrene in combination with sulfur dioxide at 10 ppm than in rats that inhaled only benzo[a]pyrene. This does not show that sulfur dioxide is carcinogenic, but it does suggest that it may increase the carcinogenic activity of known carcinogens.

The embryotoxic and teratogenic potentials of sulfur dioxide have been evaluated in mice and rabbits (Murray et al., 1979). Exposure of mice at 25 ppm and of rabbits at 70 ppm for days 6-15 and days 6-18 of gestation, respectively, yielded no evidence of a teratogenic effect related to sulfur dioxide exposure; however, significant increases in the incidence of minor skeletal variants were observed in both species.

PHARMACOKINETICS

The penetration of sulfur dioxide to the lungs is greater during mouth breathing than during nose breathing. Sulfur dioxide is readily removed during passage through the upper respiratory tract. At concentrations higher than 1 ppm, removal is over 90% (Strandberg, 1964). Sulfur dioxide is converted to bisulfite ion (HSO3) on mixing with water. It has been shown that inhaled sulfuric acid mists with droplets of 0.4-1.1 µm in mass median aerodynamic diameter have deposition patterns similar to those of dry aerosols and that the effect of hygroscopicity is not dominant in determining the site of deposition (Dahl et al., 1983). Studies using [35S] sulfur dioxide have shown that inhaled sulfur dioxide is readily distributed throughout the body. Inhaled sulfur dioxide is only slowly removed from the lower respiratory tract. Radioactivity from inhaled labeled sulfur dioxide can be detected a week or more after inhalation and may be due to 35S binding with protein. Recent work has shown that [35S] sulfuric acid is cleared faster from smaller-diameter than from larger-diameter airways of dogs (A.R. Dahl et al., unpublished manuscript). In addition, species differences have been noted: clearance is slower in guinea pigs than in dogs and slower in dogs than in rats.

ANALYTIC METHODS

Much information is available on ambient sulfur dioxide concentrations, and the assay system is well standardized and reasonably accurate. Among the procedures used are acidimetry, colorimetry, electrochemistry, fluorimetry, flame photometry, and emission and absorption spectroscopy (Cheremisinoff and Morresi, 1981; Perry and Young, 1977). It is important to remember that sulfur dioxide is dynamically related to ${\rm SO_4}^{2-}$ (sulfate). The rates of oxidation of sulfur dioxide to sulfuric acid and conversion to sulfate are greatly increased in polluted air (Rall, 1974). The atmospheric chemistry of these reactions is complex and not completely understood.

INHALATION EXPOSURE LIMITS

OSHA (1983) recommended a TWA limit of 2 ppm. ACGIH (1980, 1983) recommended a TLV-TWA of 2 ppm for an 8-h period and a STEL of 5 ppm for 15 min. The reduction of the TLV from 5 to 2 ppm was based on available data and, in the opinion of the ACGIH committee, effects reported below 2 ppm were not serious enough to justify a lower limit.

COMMITTEE RECOMMENDATIONS

The Committee on Toxicology previously recommended EELs and CEL in 1966. After reviewing the current literature, the Committee recommended no changes in the EELs and CEL.

The present Committee's recommended EELs and CEL for sulfur dioxide and the limits proposed in 1966 are shown below.

		1966	1984
10-min	EEL	30 ppm	36 ppm
30-min	EEL	20 ppm	20 ppm
60-min	EEL	10 ppm	10 ppm
24-h	EEL	5 ppm	5 ppm
90-d	CEL	1 ppm	1 ppm

REFERENCES

Alarie, Y., Ulrich, C.F., Busey, W.M., Krumm, A.A., and MacFarland, H.N. 1972. Long-term continuous exposure to sulfur dioxide in cynomolgus monkeys. Arch. Environ. Health 24:115-128.

American Conference of Governmental Industrial Hygienists. 1980. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 375-376.

American Conference of Governmental Industrial Hygienists. 1983. TLVs(R): Threshold Limit Values for Chemical Substances an Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Hygienists. p. 93.

Asmundsson, T., Kilburn, K.H., and McKenzie, W.N. 1973. Injury and metaplasia of airway cells due to SO₂. Lab. Invest. 29:41-53.

Bates, D.V., and Hazucha, M. 1973. The short-term effects of ozone on the human lung. Proceedings of the Conference on Health Effects of Air Pollutants. National Academy of Sciences-National Research Council. Assembly of Life Sciences, October 3-5, 1973. Prepared for the Committee on Public Works, United States Senate. Serial No. 93-15. Washington, D.C. p. 507-540.

Chakrin, L.W., and Saunders, L.Z. 1974. Experimental chronic bronchitis: Pathology in the dog. Lab. Invest. 30:145-154.

Cheremisinoff, P.N., and Morresi, A.C. 1981. Air Follution Sampling and Analysis Deskbook. Ann Arbor, MI: Ann Arbor Science Publishers, Inc. 490 p.

Dahl, A.R., Felicetti, S.A., and Muggenburg, B.A. 1983. Clearance of sulfuric acid-introduced ³⁵S from the respiratory tracts of rats, guinea pigs and dogs following inhalation or instillation. Fundam. Appl. Toxicol. 3:293-297.

Dahl, A.R., Snipes, M.B., Muggenburg, B.A., and Young, T.C. 1983. Deposition of sulfuric acid mists in the respiratory tract of beagle dogs. J. Toxicol. Environ. Health 11:141-149.

Davies, A., Dixon, M., Penman, R., Widdicombe, J.G., and Wise, J.C.M. 1978. Effect of repeated exposures to high concentrations of sulphur dioxide on respiratory reflexes in rabbits. Bull. Eur. Physiopathol. Respir. 14:41-52.

Drazen, J.M., O'Cain, C.F., and Ingram, R.H., Jr. 1982. Experimental induction of chronic bronchitis in dogs: Effects on airway obstruction and responsiveness. Am. Rev. Respir. Dis. 126:75-79.

Environmental Protection Agency. 1982. Review of the National Ambient Air Quality Standards for Sulfur Oxides: Assessment of Scientific and Technical Information. Report No. EPA-450/5-82-007. Research Triangle Park, NC: U.S. Environmental Protection Agency, Office of Air Quality and Program Standards. 225 p. [Available from National Technical Information Service, Springfield, Va., as PB84-102920.]

Gökemeijer, J.D., DeVries, K., and Orie, N.G. 1973. Response of the bronchial tree to chemical stimuli. Rev. Inst. Hyg. Mines 28:195-197.

Greene, S.A., Wolff, R.K., Hahn, F.F., Mauderly, J.L., and Lundgren, D.L. 1982. Chronic bronchitis in dogs exposed to SO₂. In Inhalation Toxicology Research Institute Annual Report 1981-1982, Report no. LMF-102. Albuquerque, N.M.: Lovelice Biomedical and Environmental Research Institute. p. 439-444.

Greenfield, Attaway and Tyler. 1976. Sulfur oxides: Current Status of Knowledge. FINAL REPORT. EPRI EA-316. Palo Alto, Calif.: Electric Power Research Institute. 155 p.

International Electric Research Exchange. 1981. Effects of SO₂ and its Derivatives on Health and Ecology. Vols. 1 and 2. Palo Alto, CA: Electric Power Research Institute.

Islam, M.S., Oellig, W.-P., and Weller, W. 1977. Respiratory damage casued by long-term inhalation of high concentration of sulphur-dioxide in dogs. Res. Exp. Med. 171:211-218.

Kennon, B.R. 1927. Report of a case of injury to the skin and eyes by liquid sulphur dioxide. J. Ind. Hyg. 9:486-487.

Koenig, J.Q., Pierson, W.E., and Frank, R. 1980. Acute effects of inhaled SO₂ plus NaCl droplet aerosol on pulmonary function in asthmatic adolescents. Environ. Res. 22:145-153.

Koenig, J.Q., Pierson, W.E., Horike, M., and Frank, R. 1981. Effects of SO₂ plus NaCl aerosol combined with moderate exercise on pulmonary function in asthmatic adolescents. Environ. Res. 25:340-348.

Laskin, S., Kuschner, M., and Drew, R.T. 1970. Studies in pulmonary carcinogenesis. Pp. 321-350 in Hanna, M.G., Jr., Nettesheim, P., and Gilbert, J.R., eds. Inhalation Carcinogenesis. AEC Symposium Series 18. Oak Ridge: U.S. Atomic Energy Commission. Division of Technical Information.

Lawther, P.J., Macfarlane, A.J., Waller, R.E., and Brooks, A.G.F. 1975. Pulmonary function and sulphur dioxide, some preliminary fixlings. Environ. Res. 10:355-367.

Murray, F.J., Schwetz, B.A., Crawford, A.A., Henck, J.W., Quast, J.F., and Staples, R.E. 1979. Embryotoxicity of inhaled sulfur dioxide and carbon monoxide in mice and rabbits. J. Environ. Sci. Health C13:233-250.

Nordenson, I., Beckman, G., Beckman, L., Rosenhall, L. and Stjernberg, N. 1980. Is exposure to sulphur dioxide clastogenic? Chromosomal aberrations among workers at a sulphite pulp factory. Hereditas 93:161-164.

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air Contaminants. 29 CFR 1910.1000.

Perry, R., and Young, R.J. 1977. Handbook of Air Pollution Analysis. New York: John Wiley & Sons. Distributed in the U.S.A. by Halsted Press. 506 p.

Rall, D.P. 1974. Review of the health effects of sulfur oxides. Environ. Health Perspect. 8:97-121.

Sheppard, D., Wong, W.S., Uehara, C.F., Nadel, J.A., and Boushey, H.A. 1980. Lower threshold and greater bronchomotor responsiveness of asthmatic subjects to sulfur dioxide. Am. Rev. Respir. Dis. 122:873-878.

Sheppard, D., Saisho, A., Nadel, J.A., and Boushey, H.A. 1981. Exercise increases sulfur dioxide-induced bronchoconstriction in asthmatic subjects. Am. Rev. Respir. Dis. 123:486-491.

Strandberg, L.G. 1964. SO₂ absorption in the respiratory tract. Studies on the absorption in rabbit, its dependence on concentration and breathing phase. Arch. Environ. Health 9:160-166.

Weir, F.W., and Bromberg, P.A. 1975. Effects of sulfur dioxide on healthy and peripheral airway impaired subjects. Recent Advances in the Assessment of the Health Effects of Environmental Pollution. Vol. IV: 1989-2000. EUR 5360. Luxembourg: Commission of the European Communities.

Woodford, D.M., Coutu, R.E., and Gaensler, E.A. 1979. Obstructive lung disease from acute sulfur dioxide exposure. Respiration 38:238-245.

VINYLIDENE CHLORIDE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROPERTIES

CH2=CC12 Chemical formula: 96.96 Molecular weight:

Chemical names: 1,1-Dichloroethylene,

dichloroacetylene, 1,1-dichloroethane

Synonym: Boiling point: 37°C -10°C Flash point:

Saturated vapor pressure: 591 torr (20°C)

Sparingly soluble in water; soluble Solubility:

in most organic solvents

General characteristics: Volatile, colorless liquid that

polymerizes readily; mild sweet odor

1 ppm = 3.97 mg/m^3 Conversion factors:

 $1 \text{ mg/m}^3 = 0.25 \text{ ppm}$

OCCURRENCE AND USE

Vinylidene chloride (VDC) is prepared from ethylene chloride or by passing trichloroethylene over any of several alkaline materials at a temperature above 70°C. It is an intermediate in production of Saran and Velon polymeric plastics for films and coatings. At high concentrations, it may decompose explosively to form CO and phosgene (Stecher et al., 1968). VDC has been reported to be a contaminant of space vehicles and submarines (Saunders, 1967).

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

An uncontrolled 4-d exposure to VDC and many other compounds at a low (but variable and unknown) concentration occurred in a NASA-sponsored Manned Environmental Systems Assessment project involving a five-man crew. Signs and symptoms included loss of appetite, extreme nausea, vomiting, facial-muscle symptoms, and headache. Herpes-like lesions developed on the faces of all participants shortly after the 4-d exposure (Saunders, 1967).

An epidemiologic survey of 138 workers industrially exposed to VDC at 5-70 ppm (TWA) for periods of less than 1 yr and up to over 10 yr revealed no substantial adverse health effects related to exposure (Ott et al., 1976).

EFFECTS ON ANIMALS

LT₅₀s for rats were determined for VDC by Andersen et al. (1979):

Concentration, ppm	Time, h
100	8.0
200	4.1
500	3.0
1,000	2.4
2,000	1.4

Inhalation of VDC by the rat can result in irritation of the eyes and nose, excessive salivation, respiratory distress, tremors, convulsions, incoordination, prostration, narcosis, and death due to vascular collapse and shock (Carpenter et al., 1949; Jaeger et al., 1973). Exposure to VDC at 200 ppm in air for 4 h produced severe liver damage in rats (Carpenter et al., 1949). Mild liver damage resulted from a 23-h exposure at 60 ppm (Short et al., 1977). Mice exposed at 50 ppm showed progressive renal necrosis in the first 24 h (Reitz et al., 1979); severe tubular necrosis has been reported in mice exposed at 15 ppm for 23 h (Short et al., 1977).

Dogs, rats, guinea pigs, rabbits, and monkeys exposed to VDC by inhalation at 99 ppm, 8 h/d, 5 d/wk for 6 wk showed no mortality or evidence of toxicity. However, in continuous exposure at 15 ppm, deaths occurred in 7 of 15 guinea pigs (between days 4 and 9) and 3 of 9 monkeys (days 26, 60, and 64) (Prendergast et al., 1967).

Liver damage and kidney damage were seen in most of the animals exposed continuously at 189 mg/m³. Nonspecific lung damage was also seen in most animals (Prendergast et al., 1967).

VDC caused epinephrine-induced arrhythmias in rats. Epinephrine in doses as low as 0.5 $\mu g/kg$ produced a series of premature ventricular contractions in rats exposed to VDC at about 25,000 ppm for 47 min. The cardiac effects were completely reversible on discontinuation of VDC inhalation. Pretreatment of the animals with phenobarbital increased the cardiac effects; that suggests they were due to a metabolite of VDC, in that phenobarbital induces microsomal enzymes (Siletchnik and Carlson, 1974).

Rampy et al. (1977) published an interim report of a 2-yr study on VDC toxicity. Sprague Dawley rats that inhaled VDC at 25 or 75 ppm, 6 h/d, 5 d/wk, for 18 mo failed to develop VDC-related tumors, as judged by gross examination 24 mo after the beginning of exposure.

Maltoni et al. (1977) exposed Sprague Dawley rats to VDC at 10, 25, 50, 100, and 150 ppm, 4 h/d, 4-5 d/wk, for 12 mo and, under a similar protocol, Swiss mice at 10 and 25 ppm. Early result3, 30 wk after the last exposure (animal ages, 91-98 wk), showed an increase in the incidence of mammary tumors in treated groups, but the increase was not dose-related; one Zymbal's-gland carcinoma was seen in a rat exposed at 100 ppm. VDC did produce renal adenocarcinomas without metastases in

mice exposed at 25 ppm, but not at 10 ppm. Oral administration of VDC to rats (25 mg/kg) and inhalation exposure of Chinese hamsters (25 ppm, 4 h/d, for 52 wk) did not result in tumor formation (Maltoni et al., 1977). A carcinogenesis bioassay of VDC conducted on rats (1 or 5 mg/kg) and mice (2 or 10 mg/kg) in which the chemical was given by gavage was negative (National Toxicology Program, 1982).

Another carcinogenesis study produced different results (Lee et al., 1978). CD-1 mice were exposed to VDC at 55 ppm, 6 h/d, 5 d/wk, for 12 mo; 3 of 70 mice tested developed hemangiosarcomas of the liver, and 6 of 70 had bronchioalveolar adenomas. Those results are comparable with those of exposure to vinyl chloride. Similar treatment of rats produced a hemangiosarcoma in a lymph node of one animal and another in subcutaneous tissue of a second rat; 71 animals were tested. The VDC was reported to be 99% pure, but analysis was not provided. In the Maltoni et al. study, the material used reportedly contained 99.95% VDC, $0.04\sqrt[3]{1,2}$ -dichloroethylene, 0.01% acetone, 0.005% methylene chloride, and 0.002% monochloroacetylene and dichloroacetylene; p-methoxyphenol was added at 200 ppm as a stabilizer. The Lee et al. report commented on gross lesions in several organs in mice, but did not mention any changes in the kidneys; an earlier report from the same laboratory indicated that VDC was renotoxic at 15 ppm in just 23 h.

Hong et al. (1981) conducted a 1-yr followup of neoplastic changes after VDC exposure of rats and mice at 50, 250, and 1,000 ppm as a sequel to the work of Lee et al. (1978). Results showed that cumulative tumor incidence in liver, lung, and mammary gland increased with dose and duration of exposure. Evidence of a VDC exposure threshold was presented. Chu and Milman (1981) reviewed carcinogenesis results on VDC and related compounds.

Reitz et al. (1980) studied the effects of VDC on DNA synthesis and DNA repair in rats and mice. These animals were exposed to VDC at 10 and 50 ppm for 6 h; for comparison purposes, dimethylnitrosamine was also studied in other animals. DNA alkylation was minimal (one or two orders of magnitude lower than dimethylnitrosamine). However, DNA repair in the mouse kidney was much less than in the liver. Tissue damage and increased DNA repair (by a factor of 25) occurred at 50 ppm in the mouse kidney, but not at 10 ppm. This suggests that the tumors observed in mice exposed to VDC arise primarily through effects of the chemical on nongenetic components of the cells.

PHARMACOKINETICS

Exposure of rats to [14C]VDC for several hours has demonstrated that most of the label is excreted in the urine, and the balance is expired as \$14CO_2\$ and unchanged VDC, excreted in the feces, or retained in the tissues (McKenna et al., 1977). VDC apparently is metabolized to an intermediate that can bind covalently to protein and nucleic acids (McKenna et al., 1977, 1978; Reitz et al., 1979). In a study on the pharmacokinetics of [14C]VDC in rats, McKenna et al. (1978) found that fasting had no effect on its metabolism when rats were exposed to vapor at 10 ppm for 6 h; but fasted rats exposed at 200 ppm for 6 h showed a reduced capacity to metabolize VDC, and liver damage and

kidney damage were also noted. The two major urinary metabolites detected--N-acetyl-S-(2-hydroxyethyl)cysteine and thiodiglycolic acid--indicate that a major pathway for detoxification of VDC is conjugation with liver glutathione. Reitz et al. (1979) reported that the extent to which DNA alkylation occurs in VDC-treated rats and mice (10-50 ppm) correlates well with the carcinogenicity results of the Maltoni et al. study: alkylation of rat liver and kidney DNA and mouse liver DNA is low, compared with alkylation of DNA in mouse kidney, the target organ for VDC. These investigators suggested that VDC may act as a carcinogen, not only because its administration can lead to alkylation of mouse kidney DNA, but also because of its frank toxicity to mouse kidney, which forces an increase in the rate of DNA replication as regeneration takes place in the kidney. Hence, the toxicity of VDC to the kidney could promote the initiation effect of DNA alkylation. Such a correlation between toxicity and carcinogenicity is well established with a variety of chemical compounds.

The importance of pharmacokinetics needs to be considered. The carcinogenicity of VDC in the mouse kidney can be hypothetically explained by a metabolic transformation of VDC to an alkylating agent that initiates the carcinogenic process by covalent binding to DNA and promotes the process by stimulating cellular regeneration. If such initiation and promotion can occur only when VDC exposures are high enough to deplete tissue stores of glutathione (which detoxifies activated VDC), then it can be argued that there is some degree of exposure to VDC that would result in a low probability of tumor formation. Although not yet substantiated, this is a plausible hypothesis. Therefore, it seems reasonable that, in the case of VDC, EELs can be set if sufficient data are available to permit the Committee to judge what might be an exposure unlikely to result in a toxic effect in humans.

INHALATION EXPOSURE LIMITS

The ACGIH has recommended a TLV-TWA for VDC of 5 ppm and a 15-min TLV-STEL of 20 ppm (ACGIH, 1983). The TLV-TWA of 5 ppm was considered "low enough to prevent overt toxicity in exposed workers" (ACGIH, 1982). OSHA adopted 10 ppm as the federal workplace standard for VDC (OSHA, 1983).

COMMITTEE RECOMMENDATIONS

In 1966, the Committee recommended a 24-h EEL and a 90-d CEL for VDC; however, considerable data have since accumulated (Table 12) that indicate that VDC is a potent hepatotoxin and is also a hepatocarcinogen.

The data most helpful in recommending an EEL for VDC come from the study of Ott et al. (1976), who examined the mortality and health-examination findings of 138 company employees exposed to VDC at measured concentrations in the absence of vinyl chloride. No occupation-related disorders could be detected among workers exposed to VDC at estimated concentrations of 5-70 ppm (TWA) over several years.

The Committee notes that, although the effects of single short-term exposures to carcinogens cannot be predicted, the probability of tumor formation under these conditions would be low--and less than that associated with repeated exposures. Because of the possibility of liver and kidney damage from acute exposure, the Committee recommends a lowering of the 24-h EEL to 10 ppm. The Committee notes that this concentration is approximately 20 times greater than that suggested by the Committee for short-term exposures to VDC as a drinking-water contaminant. However, as pointed out in the introduction to this report, the EELs given here are for a narrowly defined, healthy, adult working population, whereas drinking water containing VDC could be consumed for up to a week by a more heterogeneous population, including young children and other persons with increased sensitivity to the effects of this chemical.

The Committee's recommendation for the 90-d CEL is based on animal studies (continuous and repeated exposure of rats, rabbits, guinea pigs, dogs, and monkeys) and the previously cited (8 h/d) occupational exposure of humans. Continuous exposure at 15 ppm produced mortality among some guinea pigs and monkeys, but surviving animals displayed no organ toxicity. Applying a 100-fold uncertainty factor to the concentration fatal to these species, i.e., guinea pigs and monkeys, the Committee recommends a CEL of 0.15 ppm.

The present Committee's recommended EEL and CEL for VDC and the limits proposed in 1966 are shown below.

1966 1984 24-h EEL 25 ppm 10 ppm 90-d CEL 2 ppm 0.15 ppm

TABLE 12
Summary of Results of Recent Inhalation-Toxicology Studies on VDC

Species	Concentration, ppm	Duration of Exposure	Toxic Effects	Reference
Human	5-70	Several years	No clinical signs	Ott et al., 1976
Monkey	25, 47	24 h/d, 90 d	Death	Prendergast et al., 1967
Guinea pig	; 15, 25, 47	24 h/d, 90 d	Death	Prendergast et al., 1967
Rat	500	6 h/d, 20 d	Decreased body weight, liver toxicity	Gage, 1970
Rat	150	4 h	Increased serum alamine-α-keto-glutarate transamin-ase 24 h later	Jaeger <u>et al</u> ., 1975
Rat	100	4 h	None	Jaeger <u>et al.</u> , 1975
Rat	75, 100	4 h/d, 5 d/wk, 12 mo	No dose-related tumors	Viola and Caputo, 1977
Rat	25, 75	6 h/d, 5 d/wk, 18 mo	Cytoplasmic vacuo- lization in hepatocytes after 1 mc, no gross tumors at 24 mo	Rampy ct al., 1977
Rat	60	22-23 h/d, 2 d	Increased serum glutamic oxaloacetic transaminase and serum glutamic pyruvic transaminase	Short <u>et al</u> ., 1977
Rat	55	6 h/d, 5 d/wk, 12 mo	Hemangiosarcoma in lymph nodes and sub- cutaneous tissue	Lee et al., 1978
Rat	47	24 h/d, 90 d	Kidney injury	Prendergast et al., 1967
Rat	10, 40	6 h/d, 5 d/wk, 4 wk	No vacuolization	Rampy et al., 1977

Table 12 (continued)

Species	Concentration, ppm	Duration of Exposure	Toxic Effects	Reference
Mouse	100	4 h/d, 2 d	Death	Maltoni et al., 1977
Mouse	60	22-23 h/d, 2 d	Death	Short et al., 1977
Mouse	55	6 /d, 5 d/wk, 12 mo	Liver herangiosarcoma and bronchioalveolar adenoma	Lee et al., 1978
Моиве	50	4 h/d, 4 d	Death	Maltoni <u>et al</u> ., 1977
Mouse	15	1 d	Increased serum glutamic oxaloacetic transaminase and serum glutamic pyruvic transaminase	Short <u>et al</u> ., 1977
Dog, rat, monkey	47	24 h/d, 90 d	Liver injury	Prendergast et al., 1967
Not stated	1 50, 100	8 h/d, 5 d/wk, several months	Liver and kidney injury	Torkelson and Rowe, 1981
Not stated	25	8 h/d, 5 wk, several months	"Minimal" liver and kidney injury	Torkelson and Rowe, 1981

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Vinylidene chloride. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 433(82)-434(82).

American Conference of Governmental Industrial Hygienists. 1983. TLVs (R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

Andersen, M.E., French, J.E., Gargas, M.L., Jones, R.A., and Jenkins, L.J., Jr. 1979. Saturable metabolism and the acute toxicity of 1,1-dichloroethylene. Toxicol. Appl. Pharmacol. 47:385-393.

Carpenter, C.P., Smyth, H.F., Jr., and Pozzani, U.C. 1949. The assay of acute vapor toxicity, and the grading and interpretation of results on 96 chemical compounds. J. Ind. Hyg. Toxicol. 31:343-346.

Chu, K.C., and Milman, H.A. 1981. Review of experimental carcinogenesis by compounds related to vinyl chloride. Environ. Health Perspect. 41:211-220.

Gage, J.C. 1970. The subscute inhalation toxicity of 109 industrial chemicals. Br. J. Ind. Med. 27:1-18.

Hong, C.B., Winston, J.M., Thornburg, L.P., Lee, C.C., and Woods, J.S. 1981. Follow-up study on the carcinogenicity of vinyl chloride and vinylidene chloride in rats and mice: Tumor incidence and mortality subsequent to exposure. J. Toxicol. Environ. Health 7:909-924.

Jaeger, R.J., Trabulus, M.J., and Murphy, S.D. 1973. The interaction of adrenalectomy, partial adrenal replacement therapy, and starvation with hepatotoxicity and lethality after 1,1-dichloroethylene intoxication. Toxicol. Appl. Pharmacol. 25:491, Abstr. no. 133.

Jaeger, R.J., Conolly, R.B., and Murphy, S.D. 1975. Short term inhalation toxicity of halogenated hydrocarbons. Effects on fasting rats. Arch. Environ. Health 30:26-31.

Lee, C.C., Bhandari, J.C., Winston, J.M., House, W.B., Dixon, R.L., and Woods, J.S. 1978. Carcinogenicity of vinyl chloride and vinylidene chloride. J. Toxicol. Environ. Health 4:15-30.

Maltoni, C., Cotti, G., Morisi, L., and Chieco, P. 1977. Carcinogenicity bioassays of vinylidene chloride. Research plan and early results. Med. Lav. 68:241-262. McKenna, M.J., Watanabe, P.G., and Gehring, P.J. 1977. Pharmacokinetics of vinylidene chloride in the rat. Environ. Health Perspect. 21:99-105.

McKenna, M.J., Zempel, J.A., Madrid, E.O., and Gehring, P.J. 1978. The pharmacokinetics of [14C] vinylidene chloride in rats following inhalation exposure. Toxicol. Appl. Pharmacol. 45:599-610.

National Toxicology Program. 1982. Carcinogenesis Bioassay of Vinylidene Chloride in F344 Rats and B6C3F₁ Mice (Gavage Study). NTP Tech. Rept. Series No. 228. Bethesda, Maryland: National Toxicology Program, U.S. Department of Health and Human Services, Public Health Service, National Institutes of Health. 184 p. [NTP-80-82; NIH Publ. No. 82-1784]

Occupational Safety and Health Administration. 1983. Toxic and Hazardous Substances. Air contaminants. 29 CFR 1910.1000.

Ott, M.G., Fishbeck, W.A., Townsend, J.C., and Schneider, E.J. 1976. A health study of employees exposed to vinylidene chloride. J. Occup. Med. 18:735-738.

Prendergast, J.A., Jones, R.A., Jenkins, L.J., Jr., and Siegel, J. 1967. Effects on experimental animals of long-term inhalation of trichloroethylene, carbon tetrachloride, 1,1,1-trichloroethane, dichlorodifluoromethane, and 1,1-dichloroethylene. Toxicol. Appl. Pharmacol. 10:270-289.

Rampy, L.W., Quast, J.F., Humiston, C.G., Balmer, M.F., and Schwetz, B.A. 1977. Interim results of two-year toxicological studies in rats of vinylidene chloride incorporated in the drinking water or administered by repeated inhalation. Environ. Health Perspect. 21:33-43.

Reitz, R.H., Watanabe, P.G., McKenna, M.J., Quast, J.F., and Gehring, P.J. 1979. Effects of vinylidene chloride on DNA synthesis and DNA repair in the rat and mouse: A comparative study with dimethylnitrosamine. (Report to Manufacturing Chemists Assoc.) Midland, Michigan: Dow Chemical Co., Toxicology Research Laboratory. 31 p.

Reitz, R.H., Watanabe, P.G., McKenna, M.J., Quast, J.F., and Gehring, P.J. 1980. Effects of vinylidene chloride on DNA synthesis and DNA repair in the rat and mouse: A comparative study with dimethylnitrosamine. Toxicol. Appl. Pharmacol. 52:357-370.

Saunders, R.A. 1967. A new hazard in closed evironmental atmospheres. Arch. Environ. Health 14:380-384.

Short, R.D., Winston, J.M., Minor, J.L., Hong, C.-B., Seifter, J., and Lee, C.-C. 1977. Toxicity of vinylidene chloride in mice and rats and its alteration by various treatments. J. Toxicol. Environ. Health 3:913-921.

Siletchnik, L.M., and Carlson, G.P. 1974. Cardiac sensitizing effects of 1,1-dichloroethylene: Enhancement by phenobarbital pretreatment. Arch. Int. Pharmacodyn. Ther. 210:359-364.

Stecher, P.G., Windholz, M., and Leahy, D.S., eds. 1968. The Merck Index: An Encyclopedia of Chemicals and Drugs. 8th ed. Rahway, New Jersey: Merck & Co., Inc. p. 1109.

Torkelson, T.R., and Rowe, V.K. 1981. Halogenated aliphatic hydrocarbons containing chlorine, bromine, and iodine. Vinylidene chloride. In: Clayton, G.D., and Clayton, F.E., eds. Patty's Industrial Hygiene and Toxicology. Vol. IIB. Toxicology. 3rd rev. ed. New York: John Wiley & Sons. p. 3545-3550.

Viola, P.L., and Caputo, A. 1977. Carcinogenicity studies on vinylidene chloride. Environ. Health Perspect. 21:45-47.

XYLENE

BACKGROUND INFORMATION

PHYSICAL AND CHEMICAL PROFERTIES*

	o-Xylene	m-Xylene	<u>p-Xylene</u>
CAS number: Chemical formula:	1330-20-7 C6H4(CH3)2	1330-20-7	1330-20-7 C6H4(CH3)2
C6H4(CH3)2	904 (9 3 / 2		00.4(0.0)/2
Molecular weight:	106.16	106.16	106.16
Boiling point (760 mm Hg):	144.411°C	139.103°C	138.351°C
Freezing point:	-25.182°C	-47.872°C	13.263°C
Density (25°C):	0.87596 g/ml	0.85990 g/ml	0.85669 g/m1
Vapor density (air = 1):	3.7	3.7	3.7
Flash point (closed cup):	17.2°C	25°C	25°C
Vapor pressure (25°C,760 mm Hg):	6.6	8.39	8.87
Conversion factors for three		2	
isomeric xylenes:	1 ppm = 4.34 mg/m^3 $1 \text{mg/m}^3 = 0.23 \text{ ppm}$		
General characteristics:	Clear flammable liquid; aromatic hydrocarbon odor; commercial form is mixture of three isomers, with meta form usually principal component; insoluble in water; miscible with organic solvents		

OCCURRENCE AND USE

The xylenes are major commodity chemicals in this country and around the world. The National Research Council report The Alkyl Benzenes (1981) showed that in the United States the annual production of xylenes over the last decade averaged approximately 2.7 million metric tons; about 15.3 million metric tons has been used per year by the chemical industries. Because of the impetus to remove lead from gasoline and the excellent antiknock properties of these compounds, they are found in increasing concentrations in gasoline. In a gasoline sample reported in the NRC document, m- and p-xylenes constituted 6.73% and o-xylene 2.86% by weight. Reports on the use of these compounds in solvents have suggested that as much as 500,000 metric tons of xylenes is used for this purpose per year. Because of their high volatility and ubiquitous use, these compounds pervade the environment; one result is extensive human exposure. Xylenes are monitored as possible contaminants of submarine atmosphere.

^{*} Adapted from NRC, 1980.

SUMMARY OF TOXICITY INFORMATION

EFFECTS ON HUMANS

Evidence of brain hemorrhages and axonic snoxia was observed in a person who died as a result of exposure to xylene at approximately 10,000 ppm over an 18-h period (Morley et al., 1970). In another case, a man was exposed at 60-350 ppm to mixed solvents containing 75% xylene and experienced giddiness, anorexia, and vomiting (Glass, 1961). In a similar report of exposure to a mixed solvent containing 80% xylene, a person who apparently suffered from latent epilepsy displayed seizures; that suggested that xylene might exacerbate seizures in susceptible people (Goldie, 1960). In each of these instances, either mixtures were involved or the extent of exposure was not well documented.

Neither flicker fusion nor reaction time was affected in 23 volunteers exposed at 100 or 200 ppm for 3-7 h (Ogata et al.,1970). In a similar study, Gusev (1968) reported changes in brain electric activity at 9.07 ppm, but not at 0.05 ppm. Fifteen male subjects were exposed to xylene vapor at approximately 100 or 300 ppm for 70 min (Gamberale et al., 1978). No noticeable change in test performance (numerical ability, reaction time, short-term memory, and critical flicker fusion) was a n. However, subjects exposed to xylene at 300 ppm for 70 min when the exposure period began with 30 min of reactise on a bicycle showed significant impairment of performance in tests for numerical ability, short-term memory, and choice reaction time. The difference was apparently due to an increase in the uptake of xylene in subjects that exercised during the first part of the exposure period.

Savolainen et al. (1979b), Riihimaki and Savolainen (1980), and Savolainen et al. (1980) exposed six volunteers to m-xylene at 100 or 200 ppm, 6 h/d, 3 d/wk for 2 wk. In one part of the experiment, constant exposure concentrations were used; in another, the concentration was varied so that peak concentrations of 400 ppm were obtained. During the first week, significant increases in reaction time and some impairment of equilibrium were observed at 100 ppm, but these effects were transient, perhaps because tolerance developed. The effects reappeared during the second week at the higher concentration. No changes in manual dexterity or visual functions were observed. The authors suggested that light exercise reverses the effects of xylene. At 200-400 ppm, the report suggested, the subjects displayed decreased vigilance, as indicated by EEG changes.

Carpenter et al. (1975) studied the odor threshold for mixed xylenes. They estimated it to be about 1 ppm. They suggested that discomfort and dizziness would ensue at 460 ppm and concluded that 110 ppm would be tolerable for working.

^{*} Flicker fusion refers to the frequency at which a flickering light no longer appears to flicker.

EFFECTS ON ANIMALS

Cameron et al. (1938) reported that, in rats and mice, the lethal doses by subcutaneous injection were 5-10 ml/kg for p- and m-xylenes and 2.5-5.0 ml/kg for o-xylene. The lethal doses by intraperitoneal injection were 2-2.5 ml/kg for p- and m-xylenes and 1.5-2 ml/kg for o-xylene. The acute LD50 in male rats receiving a single peroral administration of 95% pure xylene (19% o-, 52% m-, and 24% p-) was 4.3 g/kg (Wolf et al., 1956). The impurities were not identified. Repeated skin contact with the undiluted xylene solution led to erythema and slight necrosis in rabbits. Instillation into the rabbit eye led to conjunctival irritation and very slight, transient corneal injury.

Exposure of rats and mice to o- and m-xylenes (separately) by inhalation at 2,000-3,000 ppm (8.7-13.0 g/m³) for 24 h resulted in deaths, and mice were reportedly more sensitive than rats to m-xylene (Cameron et al., 1938). Deaths were not observed in either rats or mice that had been exposed to o-xylene at 4,912 ppm (21.3 g/m³) for 24-28 h, but they did occur after exposure at 19,650 ppm (85.3 g/m³) for 12 h. Exposure of rats to the various pure isomers of xylene separately at 1,000-1,500 ppm (4.3-6.5 g/m³) 8 h/d for 14 d produced no deaths, and no specific organ changes were observed on autopsy. It should be noted that, generally, only small groups of animals were studied in this relatively early report, and the source and purity of the compounds used were not specified.

In a later study (Carpenter et al., 1975), the LC50 in male rats for a 4-h inhalation exposure to a mixture of xylenes was 29 (22-37) g/m^3 . Pathologic findings in the 16 rats that died after exposure at 43 g/m^3 included atelectasis, hemorrhage, and interlobular edema of the lung (two cases each). At concentrations as low as 5.8 g/m^3 , the investigators observed transient irritation, protraction of the eyes, and lack of coordination of the extremities. Rats exposed at g/m^3 displayed a slight loss of coordination by the second hour of exposure. No unusual effects were observed after exposures at approximately 2 g/m^3 . Respiratory-tract irritation, as evidenced by a decrease of 50% or more in respiratory rate, was observed in mice exposed at 5.6 g/m^3 or more for 1 min. This did not occur at 2 g/m^3 or during the 15-min period after exposure.

TANGET ESTABLE BOSTON POSTER FOR SOME BOSTON FOR SOME BOSTON POSTER POST

Exposure of cats at 41 g/m³ for 2 h resulted in a classic nervous-system effect: the sequential development of salivation, ataxia, tonic and clonic spasms, anesthesia, and death. Pathologic examination revealed no lesions that were obviously related to the exposure. Dogs exposed at approximately 4 g/m³ experienced an increase in lacrimation that began after 1 h and persisted throughout the 4-h exposure period.

LC50s of 5,267 ppm (22.8 g/m³) for m-xylene, 4,595 ppm (19.9 g/m³) for o-xylene, and 3,907 ppm (17 g/m³) for p-xylene were reported in mice (Bonnet et al., 1979).

Carpenter et al. (1975) exposed 4 male beagles and 25 male rats at 3.5, 2.0, or 0.77 g/m^3 or to control air 6 h/d, 5 d/wk, for up to 66 d. There was no evidence of toxicity of xylene.

Jenkins et al. (1970) exposed rats, guinea pigs, monkeys, and dogs to o-xylene at either 780 ppm (3358 mg/m³) for 8 h/d, 5 d/wk for 30 exposures or 78 ppm (389 mg/m³) continuously for 90 d. Mortality was 3 of 15 rats, none of 15 guinea pigs, none of 2 dogs, and 1 of 4 monkeys. One dog was tremulous and one rat died during the continuous exposure. Hematologic studies and evaluation of necropsy material did not show any effect of o-xylene.

Male guinea pigs were given intraperitoneal injections of American Chemical Society reagent-grade xylene (isomer not specified) at 1,000 mg/kg (DiVincenzo and Krasavage, 1974). The investigators observed an increase in serum ornithine carbamoyl transferase, which is said to be an indicator of hepatocellular damage. Histologic examination revealed a moderate degree of hepatic lipid accumulation, but no tissue necrosis. After intraperitoneal injections at 2,000 mg/kg, three of four guinea pigs died.

In behavioral studies, Battig and Grandjean (1964) failed to detect alterations in a conditioned-avoidance protocol with rats exposed to xylene by inhalation at 800 ppm initially and then 550-750 ppm for the balance of the 2.5-h exposure. Desi et al. (1967) injected xylene into rats at 0.02, 0.05, or 0.1 ml/100 g of body weight subcutaneously and compared them with control animals for ability to run a maze. Treatment continued for 28 d, after which the rats given the highest dose died. Other treated animals lost weight. The authors concluded that xylene prevents the acquisition of learned behavior, but does not facilitate loss of acquired behavior. The authors' conclusions require substantiation.

Carpenter et al. (1975) exposed rats to mixed xylenes at 2,800 ppm (12.2 g/m³ and reported irritation and prostration within 2-3 h. Exposure at 1,300 ppm (5.6 g/m³) led to ataxia. Exposure of dogs and rats at 580 ppm did not produce these effects. Batchelor (1927) had previously reported that exposure at 980 and 620 ppm (4.3 and 2.7 g/m³) caused no central effects. Lazarev (1929) reported that the production of narcosis in mice by o-, m-, and p-xylenes required 3,846, 1,923, and 7,885 ppm, (16.7, 8.3, and 34.2 g/m³), respectively.

Intravenous infusion of a 10% solution of m-xylene (commercial-grade) as a lipid emulsion into rabbits produced a concentration of 30 ppm (0.130 g/m³) in blood and caused positional nystagmus, but no vestibular effects, at a steady-state blood content of 10 ppm (Aschan et al., 1977).

Savolainen er al. (1979a) studied behavioral and neurochemical changes in young rats exposed to xylene at 300 ppm (1.3 g/m³) 6 h/d, 5 d/wk, for 5-18 wk with and without simultaneous ingestion of ethanol. Although the xylene alone had no behavioral effects, the mixed exposure resulted in increases in preening and ambulation, which suggested a synergistic effect; when ethanol was given alone, there was a transient decrease in preening. Minor changes in brain enzymes were observed, including an increase in superoxide dismutase activity after 14 wk of exposure to xylene alone and an increase in brain proteolytic activity in animals given both ethanol and xylene at 9-14 wk. There was no indication of a cause-effect relation between the enzyme changes and changes in preening and ambulation, nor was there evidence that these minor changes in enzyme activity constituted serious toxic impairment.

Savolainen and Pfaffli (1980) studied neurochemical changes in rat brain after inhalation of m-xylene at 48.8, 393, and 739 ppm $(0.21, 1.7, and 3.2 \text{ g/m}^3)$ 6 h/d, 5 d/wk, for 2 wk. Xylene in brain and peripheral fat increased from the first to the second week. Brain content of NADPH diaphorase increased, whereas brain superoxide dismutase decreased. Biochemical analyses on rats withdrawn from exposure for 2 wk indicated that the biochemical effects were largely abolished within that time, although cerebral RNA was above the control value at the two higher exposures. The latter data disagree with the results of the earlier study, however, and the difference in duration of exposure may account for the discrepancy. The most important finding was the increase in brain RNA, which persisted after the termination of the treatment period.

Andersson et al. (1981) exposed rats to commercial xylene and to its component o-, m-, and p-xylenes and ethylbenzene at 2,000 ppm (8.7 g/m³) 6 h/d for 3 d. Within 16-18 h after exposure stopped, they found increases in norepinephrine turnover in the hypothalamus caused by all the compounds and increases in dopamine in the median eminence of the forebrain caused by all but ethylbenzene and c-xylene. Ethylbenzene selectively reduced norepinephrine in the paraventricular region of the hypothalamus. Prolactin secretion was reduced by p- and o-xylenes. Although no overt behavioral signs were found in these experiments, the authors suggested that xylene (by disturbing dopamine neurotransmission) can produce motor disturbances and motivational deficits.

AND PROPERTY TO COMPANY THE COURT TO COMPANY THE COMPANY THE COMPANY TO COMPANY THE COMPAN

A connection between the occurrence of sacral aplasia and occupational exposure to organic solvents has been demonstrated by Kucera (1968), who also observed a developmental malformation analogous to sacral aplasia in chick embryos treated with an unspecified concentration of xylene, which was positively correlated with time or exposure and negatively correlated with embryonic age. Krotov and Chebotar (1972) did not find any developmental defects in fetuses of rats that had inhaled xylene. Hudak and Ungvary (1978) evaluated the embryotoxic effects of xylene (a mixture of 10% o-xylene, 50% m-xylene, 20% p-xylene, and 20% ethylbenzene) at 230 ppm $(\overline{1} \text{ g/m}^3)$ given by inhalation to rats for 24 h/d on days 1-21 of pregnancy. Untreated groups of animals that inhaled pure air served as controls. No evidence of teratogenicity was found, but skeletal anomalies (extra ribs and fused sternebrae) increased, although they were not significant (p < 0.05 for both). Mirkova et al. (1983) studied the effect of daily exposure (6 h/d, 5 d/wk) to xylene at 10, 50, and 500 mg/m3 on pregnant white Wistar rats during days 1 through 21 of gestation. The authors observed that concentrations of 50 and 500 mg/m³ exerted pronounced embryotoxic and teratogenic effects. Xylene increased the incidence of anomolies of internal organs (hydroephalus, microphthalmia, intracerebral hematomas and hemorrhages in the liver). It impaired the processes of ossification of sternum and skull. At concentrations of 50 and 500 mg/m3, xylene causes disturbances in postnatal development of the F1 generation.

Xylene was not mutagenic in a battery of short-term tests: a test for mitotic gene conversion in yeast <u>Saccharomyces cerevisiae</u> D4, gene-mutation tests in bacteria (with <u>Salmonella typhimurium</u> strains TA 98, TA 100, TA 1535, TA 1537, and TA 1538) with and without activation,

and specific-locus forward-mutation induction in the L5178Y thymidine kinase Fischer mouse lymphoma cell assay. Xylene did not produce significant increases in chromosomal aberrations in rat bone marrow cells at 0.044, 0.157, and 0.441 ml/kg (Litton Bionetics, 1978).

PHARMACOKINETICS

All the xylene isomers enter the body rapidly by inhalation and less rapidly by absorption from the gastrointestinal tract (Gerarde, 1960) or through the skin (Dutkiewicz and Tyras, 1968). The blood-to-gas partition ratio for xylene has been calculated to be 29:1 (Astrand, 1975) and 42:1 (Sherwood, 1975). The rate at which xylene is absorbed through human skin that is immersed in liquid xylene has been estimated to be 4.5-9.6 mg/cm² per hour (Dutkiewicz and Tyras, 1968).

Xylene is distributed rapidly into all tissues of the body, especially into the adrenals, bone marrow, brain, spleen, and adipose tissue (Fabre et al., 1960). The pharmacokinetics of m-xylene in rats and mice fit a three-compartment model (Bergman, 1979), and more than 90% of an administered dose should be eliminated within 24 h (NRC, 1981). Comparable studies have not been conducted in humans, but Sedivec and Flek (1976) reported that only 5% of a dose of a mixture of xylenes was exhaled. Their finding that 95% of the urinary metabolites was excreted within 10 h indicates that the total body clearances of the xylenes are rather high.

The xylenes undergo side-chain oxidation to methylbenzoic acids, which in turn are conjugated with glycine and excreted into ur'ne as toluic acids in all mammals studied, including dogs, humans, rabbits, rats, and guinea pigs. In addition to side-chain oxidation, the xylenes undergo ring hydroxylation, but only to a minor extent (Bakke and Scheline, 1970). In rats, approximately 1% of a dose (100 mg/kg) of p-xylene was converted to 2,5-xyleno1, 0.9% of m-xylene was converted to 2,4-xylenol, and 0.1% of o-xylenol was converted to 3.4-xylenol. Pretreatment of rats with phenobarbital or 3-methylcholanthrene markedly increased the activity of the hepatic enzyme that converts p-xylene to p-methylbenzyl alcohol, but did not alter the activity of the enzyme in the lungs (Harper et al., 1977). Patel et al. (1978, 1979) reported that pretreatment of mice with individual xylene isomers did not alter the activity of cytochrome P-450 in the liver. However, the administration of p-xylene to rabbits inactivated cytochrome P-450 in the lung.

INHALATION EXPOSURE LIMITS

The American Conference of Governmental Industrial Hygienists (1980, 1983) recommended a TLV-TWA of 100 ppm and a 15-min TLV-STEL of 150 ppm. It believed that irritant effects will be minimal, and that no substantial degree of narcosis or chronic injuries will result from continued occupational exposure at 100 ppm. OSHA (1983) recommended a PEL of 100 ppm. Other recommendations were as follows: ANSI (1970), 100 ppm; West Germany (1974), 200 ppm; Sweden (1975), 100 ppm; Czechoslavakia (1969) and East Germany (1973), 45 ppm; and USSR (1972), 11 ppm (ACGIH, 1980).

COMMITTEE RECOMMENDATIONS

The Committee on Toxicology suggested limits for exposure to xylene in 1966: a 60-min EEL of 200 ppm, a 24-h EEL of 100 ppm, and a 90-d CEL of 50 ppm.

The Committee on Alkyl Benzene Derivatives (NRC, 1981) reviewed the toxicologic data on xylene and included the following statement in its report:

The acute toxicity of the xylenes predominantly reflects effects on the central nervous system similar to those produced by other alkyl benzenes and related compounds. Irritant effects on mucous membranes have been reported, particularly upon direct contact. There is negligible evidence of acute or chronic effects in organ systems other than the central nervous system.

Xylene is a major commodity solvent and component of gasoline, and the potential for human exposure is large. The major effects are on the central nervous system and appear to be reversible. There is no indication of genetic toxicity or carcinogenesis.

The lowest doses at which any biologic effects have been observed in humans were reported by Ogata et al. (1970) and Gusev (1968), who detected changes in electric activity of brain in humans exposed at 0.05-0.07 ppm. These changes were not related to any functional impairment. Although Savolainen et al. (1979b), Riihimaki and Savolainen (1980), and Savolainen et al. (1980) described changes in psychophysiologic functions, such as reaction time and body balance, that were also accompanied by EEG changes in humans exposed to xylene at 90-200 ppm, one of the authors (Seppolainen, personal communication) has questioned the wisdom of using these results to set exposure limits. Chronic exposure does not seem to increase sensitivity to these effects. The available evidence does not warrant revision of the exposure limits recommended in 1966.

The present Committee's recommended EELs and CEL for xylene and the limits proposed in 1966 are shown below.

	1966	1984	
60 min EEL	200 ppm	200 ppm	
24-h EEL	100 ppm	100 ppm	
90-d CEL	50 pra	50 ppm	

REFERENCES

American Conference of Governmental Industrial Hygienists. 1980. Trichlorofluoromethane. Documentation of the Threshold Limit Values. 4th ed. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. p. 439-440.

American Conference of Governmental Industrial Hygienists. 1983. TLVs^(R): Threshold Limit Values for Chemical Substances and Physical Agents in the Work Environment with Intended Changes for 1983-1984. Cincinnati, Ohio: American Conference of Governmental Industrial Hygienists. 93 p.

American Petroleum Institute. 1978. Mutagenicity Evaluation of Xylene. Washington, D.C.: American Petroleum Institute. Med. Res. Publ. 26-60018. 150 p.

Andersson, K., Fuxe, K., Nilsen, O.G., Toftgard, R., Eneroth, P., and Gustafsson, J-A. 1981. Production of discrete changes in dopamine and noradrenaline levels and turnover in various parts of the rat brain following exposure to xylene, ortho-, meta-, and para-xylene, and ethylbenzene. Toxicol. Appl. Pharmacol. 60:535-548.

Aschan, G., Bunnfors, I. Hydén, D., Larsby, B., Odkvist, L.M. and Tham, R. 1977. Electronystagmographic and gas chromatographic studies in rabbits. Acta Otolaryngol. 84:370-376.

Astrand, I. 1975. Uptake of solvents in the blood and tissues of man: A review. Scand. J. Work Environ. Health 1:199-218.

Bakke, O.M., and Scheline, R.R. 1970. Hydroxylation of aromatic hydrocarbons in the rat. Toxicol. Appl. Pharmacol. 16:691-700.

Batchelor, J.J. 1927. The relative toxicity of benzol and its higher homologues. Am. J. Hyg. 7:276-298.

Battig, K., and Grandjean, E. 1964. Industrial solvents and avoidance conditioning in rats. A comparison of the effects of acetone, ethyl alcohol, carbon disulfide, carbon tetrachloride, toluene, and xylene on acquisition and extinction. Arch. Environ. Health 9:745-749.

Bergman, K. 1979. Whole-body autoradiography and allied tracer techniques in distribution and elimination studies of some organic solvents: Benzene, toluene, xylene, styrene, methylene chloride, chloroform, carbon tetrachloride and trichloroethylene. Scand. J. Work Environ. Health 5(Suppl 1):1-263.

Bonnet, P., Raoult, G., and Gradiski, D. 1979. Lethal concentration of 50 main aromatic hydrocarbons. Arch. Mal. Prof. Med. Trav. Secur. Soc. 40:805-810. (in French)

Cameron, G.R., Paterson, J.L.H., DeSaram, G.S.W., and Thomas, J.C. 1938. The toxicity of some methyl derivatives of benzene with special reference to pseudocumene and heavy coal tar naphtha. J. Pathol. Bacteriol. 46:95-107.

Carpenter, C.P., Kinkead, E.R., Geary, D.L., Jr., Sullivan, L.J., and King, J.M. 1975. Petroleum hydrocarbon toxicity studies. V. Animal and human response to vapors of mixed xylenes. Toxicol. Appl. Pharmacol. 33:543-558.

Dési, I., Kovács, F., Zahumenszky, Z., and Balogh, A. 1967. Maze learning in rats exposed to xylene intoxication. Psychopharmacologia 11:224-230.

DiVincenzo, G.D., and Krasavage, W.J. 1974. Serum ornithine carbamyl transferase as a liver response test for exposure to organic solvents. Am. Ind. Hyg. Assoc. J. 35:21-29.

Dutkiewicz, T., and Tyras, H. 1968. Skin absorption of toluene, styrene, and xylene by man. Br. J. Ind. Med. 25:243.

Fabre, R., Truhaut, R., and Laham, S. 1960. Recherches sur le métabolisme comparé des xylènes ou dimethylbenzènes. Arch. Mal. Prof. Med. Trav. Secur. Soc. 21:314-328. (in French)

Gamberale, F., Annwall, G., and Hultengren, M. 1978. Exposure to xylene and ethylbenzene. III. Effects on central nervous functions. Scand. J. Work Environ. Health 4:204-211.

Gerarde, H.W. 1960. Toxicology and Biochemistry of Aromatic Hydrocarbons. New York: Elsevier Publishing Co. 329 p.

Glass, W.I. 1961. Annotation: A case of suspected xylol poisoning. N. Z. Med. J. 60:113.

Goldie, I. 1960. Can mylene (mylol) provoke convulsive seizures? Ind. Med. Surg. 29:33-35.

Gusev, I.S. 1968. Comparative toxicity studies of benzene, toluol, and xylol by the reflex activity method. Pp. 60-67 in Levine, B.S., U.S.S.R. Literature on Air Pollution and Related Occupational Diseases. Volume 17. Springfield, VA.: National Technical Information Service. PB-180 522-T.

Harper, C., Drew, R.T., and Fouts, J.R. 1977. Benzene and p-xylene: A comparison of inhalation toxicities and in vitro hydroxylations. Pp. 302-311 in Jollow, D.J., Kocsis, J.J., Snyder, R., and Vainio, H., eds. Biological Reactive Intermediates: Formation, Toxicity, and Inactivation. Proceedings of an International Conference on Active Intermediates: Formation, Toxicity, and Inactivation, held at the University of Turku, Turku, Finland, July 26-27, 1975. New York: Plenum Press.

Hudak, A., and Ungvary, G. 1978. Embryotoxic effects of benzene and its methyl derivatives: Toluene, xylene. Toxicology 11:55-63.

Jenkins, L.J., Jr., Jones, R.A., and Siegel, J. 1970. Long-term inhalation screening studies of benzene, toluene, o-xylene, and cumene on experimental animals. Toxicol. Appl. Pharmacol. 16:818-823.

Krotov, Yu.A., and Chebotar, N.A. 1972. Embryotoxic and teratogenic action of some industrial substances formed during production of dimethyl terephthalate. Gig. Tr. Prof. Zabol. 16(6):40-43.

Kucera, J. 1968. Exposure to fat solvents: A possible cause of sacral agenesis in man. J. Pediatr. 72:857-859.

Lazarev, N.W. 1929. Toxicity of various hydrocarbon vapors. Arch. Exp. Pathol. Pharmakol. 143:223-233.

Mirkova, E., Zaikov C., Antov, G., Mikhailova, A., Khinkova, L., and Benchev, I. 1983. Prenatal toxicity of xylene. J. Hyg. Epidemiol. Microbiol. Immunol. 27:337-343.

Morley, R., Eccleston, D.W., Douglas, C.P., Greville, W.E.J., Scott, D.J., and Anderson, J. 1970. Xylene poisoning: A report on one fatal case and two cases of recovery after prolonged unconsciousness. Br. Med. J. 3:442-443.

National Research Council. 1980 The Alkyl Benzenes. Washington, D.C.: National Academy Press. 384 p.

Ogata, M., Tomokuni, K., and Takatsuka, Y. 1970. Urinary excretion of hippuric acid and m- or p-methylhippuric acid in the urine of persons exposed to vapours of toluene and m- and p-xylene as a test of exposure. Br. J. Ind. Med. 27:43-50.

Patel, J.M., Harper, C., and Drew, R.T. 1978. The biotransformation of p-xylene to a toxic aldehyde. Drug Metab. Dispos. 6:368-374.

Patel, J.M., Wolf, C.R., and Philpot, R.M. 1979. Interaction of 4-methylbenzaldehyde with rabbit pulmonary cytochrome P-450 in the intact animal, microsomes, and purified systems. Destructive and protective reactions. Biochem. Pharmacol. 28:2031-2036.

Riihimaki, V., and Savolainen, K. 1980. Human exposure to m-xylene. Kinetics and acute effects on the central nervous system. Ann. Occup. Hyg. 23:411-422.

Savolainen, H., Pfaffli, P., Helojoki, M., and Tengén, M. 1979a. Neurochemical and behavioural effects of long-term intermittent inhalation of xylene vapour and simultaneous etha. 1 intake. Acta Pharmacol. Toxicol. 44:200-207.

Savolainen, H., Riihimaki, V., and Linnoila, M. 1979b. Effects of short-term xylene exposure on psychophysiological functions in man. Int. Arch. Occup. Environ. Health 44:201-211.

Savolainen, H., Riihimaki, V., Seppäläinen, A.M., and Linnoila, M. 1980. Effects of short-term m-xylene exposure and physical exercise on the central nervous system. Int. Arch. Occup. Environ. Health 45:105-121.

Savolainen, H., and Pfaffli, P. 1980. Dose-dependent neurochemical conges during short-term inhalation exposure to m-xylene. Arch. loxicol. 45:117-122.

Sedivec, V., and Flek, J. 1976. The absorption, metabolism, and excretion of xylenes in man. Int. Arch. Occup. Environ. Health 37:205-217.

Sherwood, R.J. 1976. Ostwald solubility coefficients of some industrially important substances. Br. J. Ind. Med. 33:106-107.

Wolf, M.A., Rowe, V.K., McCollister, D.D., Hollingsworth, R.L., and Oyen, F. 1956. Toxicological studies of certain alkylated benzenes and benzene. Experiments on laboratory animals. AMA Arch. Ind. Health 14:387-398.